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Drivers and limits of phenotypic responses in vulnerable seagrass populations: *Zostera marina* in the intertidal

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Abstract :

1. Intertidal seagrass meadows are exposed to both marine and terrestrial environmental constraints. Seagrass vulnerability to climate changes in these highly dynamic and thermally stressful environments is concerning.

2. Using broad scale monitoring data covering contrasted intertidal environments, this study aims to provide a comprehensive view of the extent, drivers and potential limits of Zostera marina phenotypic responses. The links between phenotypic and reproductive strategies are also explored.

3. Across 500km of coastline, Z. marina exhibited extensive variations of density, morphology, aboveand belowground biomass, and rates of clonality. Variance partitioning of phenotypic traits confirmed the strong link between intertidal seagrass populations and broad-scale climate variability. However, it also highlighted a non-negligeable role of local factors such as exposure regime, substrate and tidal cycles. In its response to the environment, Z. marina displayed a trade-off between the density and size of shoots, leading to two distinct phenotypic types: high densities and low above-to-belowground biomass ratios (Type 1) in response to both high hydrodynamic and temperature stress; and low densities, high aboveground biomass per shoot and developed leaves and sheaths (Type 2) in the most stable and less stressful intertidal environments. We argue that Type 1 maximizes self-facilitation whereas Type 2 minimizes intra-specific competition, and that their occurrence matches predictions from the stressgradient hypothesis (SGH). Building on the SGH, we propose a generalized response of seagrass to environmental changes and discuss the role of light as a potential limiting resource for intertidal meadows.

4. Synthesis – Here, we show how the SGH can explain seagrass phenotypic responses, drawing on previous experimental results to provide relevant predictions across different stress gradients. We also show that Zostera marina responds to strong hydrodynamics and thermal constraints, both likely to increase in the intertidal with climate changes, with a facilitation-maximizing phenotypic type (Type 1). This strategy appears incompatible with the competition-minimizing Type 2 found when seagrass face resource limitation, such as light limitation induced by water quality degradation. This potential limit to the resilience of intertidal seagrass populations in the face of cumulative stressors raises concern about their vulnerability regarding future climate scenarios.

Keywords : aquatic plant ecology, marine phanerogams, stress-gradient hypothesis, cumulative stressors, resilience, phenotypic traits, clonality, intraspecific variability, eelgrass, foundation species

3. INTRODUCTION

Seagrasses are marine angiosperms able to form extensive meadows that are critically important for the biodiversity and functioning of marine coastal ecosystems (Duffy, 2006). They act as foundation species and thereby define many aspects of ecosystems (Ellison, 2019). In particular, they fulfil key roles in carbon sequestration and nutrient cycling, in coastal protection and erosion control, and in the support of commercial fisheries (Barbier et al., 2011). However, the rapid and profound degradation of coastal environments over the past decades (Lotze et al., 2006) has put seagrasses under high pressure and led to massive shrinkages of their distribution worldwide (Orth et al., 2006; Waycott et al., 2009).

Temperate North-East Atlantic meadows have suffered severe losses over the past decades (Airoldi & Beck, 2007). Recently, rates of seagrass losses seem to be decelerating throughout Europe and recovery of fast-growing species have been documented in several locations (de los Santos et al., 2019). Yet, challenges to the recovery of European seagrass meadows still remain. In this region, seagrass diversity is limited, especially in coastal and estuarine areas where only two seagrass species are found: Zostera marina, which is the most common and abundant, and Zostera noltii (Hily et al., 2003; Short et al., 2007). Although both species may co-occur, most meadows are monospecific stands (Moore & Short, 2006; Short et al., 2007). They are therefore acutely vulnerable to environmental stressors and susceptible to pandemic diseases (Waycott et al., 2009). In Western Europe, for instance, "wasting disease" outbreaks have led to the die-back of almost 90% of Z. marina populations in the 1930's (Godet et al., 2008). This remain the major driver of seagrass loss throughout Europe and signs of Z. marina recovery remain limited (de los Santos et al., 2019), impeded by the emergence of new challenges, such as the impact of warming waters on shallow meadows (Krause-Jensen et al., 2021).

In these conditions, resilience of North-East Atlantic seagrass meadows cannot lean on community-based processes and almost entirely relies on the adaptive capacity and response plasticity of the two species present, and in particular of the most common one: *Z. marina* (Unsworth et al., 2015). Intra-specific phenotypic (Maxwell et al., 2014) and genotypic (Hughes & Stachowicz, 2011; Reusch et al., 2005) variability of *Zostera*

species was shown to confer some resilience to the seagrass meadows. However, the capacity and limits of these response mechanisms remain to be fully determined, especially in the face of multiple interacting stressors that are likely to affect seagrass species differently depending on their life history strategies (Kilminster et al., 2015; O'Brien et al., 2018).

A continuum of ecological strategies exists across seagrass taxa, which generally follow a functional trade-off between two extremes (Kilminster et al., 2015; O'Brien et al., 2018). On the one hand, K-strategists (e.g. *Posidonia oceanica*) tend to be large and persistent species that mostly invest in clonal reproduction and on resistance traits, such as large energy storage. They often have long resistance and recovery times. On the other hand, r-strategists are smaller, colonizing species (e.g. *Ruppia maritima*) that tend to rely more on sexual reproduction and recovery traits, such as seed banks, and less on resistance traits. They therefore decline but also recover more quickly. *Zostera marina* occupies an intermediate position along this resistance-recovery continuum (Kilminster et al., 2015; O'Brien et al., 2018). Its investment between sexual and clonal reproduction (i.e. rate of clonality) can be extremely variable among and even within meadows (Becheler et al., 2014; Jarvis et al., 2012). As such, *Z. marina* seems able to tune its strategy across a wide range of the functional trade-off between r- and K-extrema and both annual (opportunistic-like) and perennial strategies have been reported depending on environmental conditions (Kim et al., 2014; Van Lent & Verschuure, 1994a).

Zostera marina phenotypic variability represents a challenge from a management perspective to interpret the temporal trajectories of the meadows, identify the drivers behind their changes and adequately evaluate their status and resilience (Kilminster et al., 2015). Variation of *Z. marina* phenotypic traits, density and reproductive strategy have been documented in relation to a wide range of environmental variables including substrate type, nutrient availability, temperature, salinity, depth, light availability and current regime (Jarvis et al., 2012; Kim et al., 2014; Moore & Short, 2006; Van Lent & Verschuure, 1994b). However, these different drivers may exert similar or opposite pressures on seagrass traits (e.g. de Los Santos et al., 2010) and current knowledge of how these multiple responses are integrated and expressed in natural conditions remains sparse (Stockbridge et al., 2020; Yang et al., 2013). In particular, the compound effects of

climate and hydrodynamic conditions, two major factors through which climate change is expected to affect coastal ecosystems (Harley et al., 2006), with local drivers, such as substrate type and tidal variation, remain to be thoroughly explored in natural intertidal populations.

There is now compelling evidence that the strong physiological constraints associated with intertidal environments may be exacerbated by interactive effects with other environmental factors and anthropogenic pressures (Lefcheck et al., 2017). These interactive effects makes shallow seagrass populations particularly vulnerable in the face of climate changes worldwide and may be a key limit to their recovery in the North-East Atlantic (Krause-Jensen et al., 2021; Unsworth et al., 2015). Hence, important experimental efforts have been devoted to understand the responses of shallow and intertidal Z. marina populations to cumulative stressors (e.g. Moreno-Marín et al., 2018; Zimmerman et al., 2017). Yet, long-term and large-scale monitoring efforts are needed to confirm, generalize and upscale available experimental knowledge in order to guide conservation efforts (Lefcheck et al., 2017; Potouroglou et al., 2014; Witman et al., 2015; Yang et al., 2013). Disentangling the relative and cumulative effects of multiple concomitant factors remains challenging in observational studies (e.g. Kaldy & Lee, 2007) and long-term monitoring across large environmental gradients is necessary for this purpose (Verheven et al., 2017). Currently, most published long-term studies of shallow Z. marina populations phenotypic responses have focused on a limited geographic area or on a single driver (Kaldy & Lee, 2007; Kim et al., 2014; Potouroglou et al., 2014) while those covering large environmental gradients have lacked temporal replication and focused on gradients of sediment and tidal conditions (Yang et al., 2013).

To fill this gap, we study the phenotypic responses of intertidal *Z. marina* across a region (1) that is a highly diverse environmental mosaic (Gallon et al., 2017), covering most of the intertidal habitats of *Z. marina* in the North-East Atlantic (Hily et al., 2003), and (2) was shown to support high genetic and phenotypic diversity for *Z. marina* (Becheler et al., 2010). We use a regional monitoring of eight intertidal *Z. marina* beds over five non-consecutive years along the 500 km coasts of Brittany (France). Monitored variables covered the density, morphology, above- and belowground biomass as well as epiphytic load and were complemented by genotypic data. The aims of this study are to (1)

disentangle and quantify the relative importance of a wide range of potential drivers (water temperature, salinity, exposure regime, tidal emersion, sediment and climatic conditions) on *Z. marina* phenotypic variability, (2) identify potential traits trade-offs constraining *Z. marina* morphological and structural responses to environmental changes in the intertidal and (3) explore the potential mechanisms underlying the morphological and reproductive strategies of *Z. marina* to address their implications for conservation purposes.

4. MATERIAL AND METHODS

4.1. Study area and sampling of Zostera marina

Eight intertidal monospecific *Zostera marina* beds were monitored along the coasts of Brittany (France; Figure 1) for five non-consecutive years, in 2007 and from 2009 to 2012, using data collected in the context of the *RÉseau BENThique* (REBENT) monitoring programme (http://www.rebent.org). These eight meadows were chosen to encompass the spectrum of environmental settings in which intertidal *Z. marina* meadows can be found in Brittany: from sheltered bays with fine sediments and turbid waters to exposed areas with coarse sediments and fully marine conditions, through semi-opened habitats (Hily et al., 2003; Moore & Short, 2006). Details on the environmental conditions encompassed in this study can be found in Table S1, Figure S7, as well as in Boyé et al. (2017) for substrate composition.

Sampling was consistently performed for all beds around the spring equinox of each year, between the end of February and the end of April, to limit the influence of the seasonal variability of the descriptors measured (Martínez-Crego et al., 2008). In order to account for within-meadow variability when interpreting differences among meadows (Balestri et al., 2003), three fixed sampling points distributed 200 metres apart were visited within each seagrass bed as shown with the Glénan meadow (Figure 1). At each sampling point, all shoots in two 0.05 m² quadrats were collected to measure densities, above and belowground biomass, and describe each shoot's morphology with measures of sheath height, leaves length and width as well as the number of leaves per shoot. Associated drifting algae biomass were also measured in each quadrat. Additionally, ten shoots were randomly collected at each sampling point to estimate epiphytic load. These

shoots were sampled as described above and measured for all variables characterising their morphology and above- and belowground biomass.

4.1.1. Morphological and structural traits measurements

Sheath height was measured from the first node to the separation mark of the leaves. The length of each leaf was measured from the first node to the apex. The number of broken leaves was counted and expressed as a percentage of the total number of leaves found in each quadrat. One leaf of median length was used to estimate the leaf width for each shoot. Leaves, roots/rhizomes, and brown, red and green algae biomass were estimated as dry weight after 24 hours desiccation at 60°C for each quadrat. Epiphytes were collected using razor blades and their biomass were measured with the same procedure. The ratio between epiphyte and leaf biomass was then computed and averaged across the 10 shoots randomly collected at each sampling point. Total Z. marina leaf and root/rhizome biomass, Z. marina densities and algae biomass were scaled up and expressed per square metre for the two quadrats. To assess the relative investment of Z. marina between its aboveground and belowground parts, a leaves-toroots/rhizomes biomass ratio was calculated within each quadrat based on the total leaf and root/rhizome biomass measured in each. Mean leaf biomass and root/rhizome biomass per shoot were estimated by dividing the total biomass by the shoot densities within each quadrat. For all other variables (sheath height, leaf length and width, number of leaves per shoot, proportion of broken leaves), mean values were calculated for each of the two quadrats separately. Broken leaves were kept in the calculation of mean leaf length to reflect both the physiological and mechanical impacts of the seagrass environment. The two per quadrat values of each sampling point were then averaged to estimate all the variables at the level of the sampling point. All Z. marina morphological and structural traits and their units are summarised in Table1.

4.1.2. Estimation of the rate of clonality

The rate of clonality *c*, corresponding to the relative frequency of the descendants resulting from clonal reproduction within a population (Marshall & Weir, 1979; Stoeckel & Masson, 2014), was inferred in six of the eight meadows in this study (all but Glénan and Sept-Îles). These six meadows were part of genetic surveys in 2009 and 2012 (Becheler

et al., 2014, 2010) where two quadrats, positioned in the central part of the *Z. marina* stand and separated by about tens metres, were sampled. Rates of clonality were inferred from the transition of genotypic frequencies between the two sampling years using the Bayesian approach ClonEstiMate (Becheler et al., 2017). This rate varies between 0 (purely sexual population) and 1 (for strict clonality).

4.2. Environmental variables

A 25 mm width and 50 mm long sediment core was collected at each sampling point for grain size distribution and organic matter content assessment. Sediments were dried in an oven (24 h at 60 °C) and separated into 15 fractions (<63 μ m, 63, 80, 100, 125, 160, 200, 315, 500, 800, 1250, 2000, 3150, 5000 and >10000 μ m) whose masses were measured. Fractions were afterwards grouped into gravel (> 2 mm), sand (63 μ m to 2 mm) and silt and clay (<63 μ m; Fournier et al. 2012). Organic matter content was estimated by mass loss on ignition at 450 °C for 5 hours.

Water temperatures, salinities and currents were obtained from the publicly available database MARC (https://marc.ifremer.fr/en) using the MARS3D model (2.5 km grain, 40 depth levels), previously validated in the study area (Lazure et al., 2009; Poppeschi et al., 2021). All variables were extracted daily for the years under study at midday near the sediment surface. Linear interpolation on neighbouring grids of the beds coordinates was used to estimate the variables at the site level (no difference for the three points within each site). Meadows being intertidal, and thus close to the land, it was necessary to shift the extraction coordinates away from the coast to avoid model edge effect (Figure S1). When needed, salinity and temperature data from the literature and environmental monitoring programme such as the Service d'Observation en Milieu Littoral (SOMLIT; Roscanvel meadow) were used to adjust the extraction coordinates. Temperature and salinity HOBO U24-002-C data loggers were also deployed for one month within three meadows to ensure the adequacy between the MARS3D model data and in situ finescale measures. Overall, this model was shown to adequately reproduce coastal hydrology in the study area (Lazure et al., 2009) but while broad-scale variation of hydrology across meadows and years may be well represented, we note that fine-scale temporal variation and extreme events remain underestimated with these data (Figure

S2; Poppeschi et al., 2021). Extracted water temperature, salinity and current time series were characterised by their distribution over the six months preceding the sampling dates of each meadows (in order to match with the response timescale of seagrass structural and demographic traits; O'Brien et al., 2018; Roca et al., 2016) with the maximum, minimum and the first, second (median) and third quartile.

The same characterisation was applied to tidal coefficients retrieved from the SHOMAR software (SHOM, 2014) in order to describe the tidal cycle during the six-month period preceding the sampling. The frequencies of spring (tidal coefficient superior to 100) and neap (inferior to 40) tides were also recorded. Tidal variations are common to all sites and reflect solely temporal changes over the whole study area. In addition, estimations of tidal range, exposure time and meadows depth are provided in Table S1 as indication of the overall tidal environment of the meadows. They were not used for the modelling of *Zostera marina* traits in the redundancy analysis (see Table 1). Indeed, all these meadows are in macrotidal environments (tidal range >4m; Whitfield & Elliott, 2012) and exposure time and depth is similar across the meadows. Moreover, these meadows being intertidal, depth is well above reported depth limits for *Zostera marina* (Duarte et al., 2007; Krause-Jensen et al., 2011) and was not considered limiting here.

Meteorological conditions were also retrieved from the nearest meteorological stations and integrated over the six months preceding the sampling dates. The meteorological variables comprised the minimum and maximum air temperature and the maximum wind velocity recorded over the six months, the total number of frost days and the cumulative rainfall over this period. Furthermore, the monthly mean of temperature, the monthly mean of the daily range of temperature, and the monthly mean of wind velocity were averaged over the six months. A North-Atlantic Oscillation (NAO) index based on the months preceding sampling (December, January and February) was retrieved from the National Center for Atmospheric Research Staff (http://climatedataguide.ucar.edu, PCbased index accessed in March 2015).

Light is generally not expected to be strongly limiting for intertidal meadows (Yang et al., 2013) and was therefore not surveyed during the monitoring. However, given the hypothesis of potential light limitation advanced by Yang et al. (2013) after the

monitoring, photosynthetically active radiation (PAR) data were retrieved *a posteriori* from EuSeaMap models (Populus et al., 2017). These PAR data are estimated from MERIS satellite data and available at a 1km resolution as the average PAR (mol photons m⁻² day⁻¹) per month over the 2005-2009 period, thus covering the beginning of this study period. The values extracted at the meadows coordinates (Figure S3) were all well above the minimum light requirement thresholds reported in the literature for *Z. marina* (Bertelli & Unsworth, 2018). Thus, they were unlikely to explain any potential light limitation of the meadows under study, and were not included in the redundancy analysis modelling the response of *Zostera marina* to environmental changes (see Table 1).

4.3. Statistical analyses

Balanced datasets are desirable for statistical tests such as multivariate analysis of variance (MANOVA) so that the year 2008, initially monitored in the REBENT programme, was discarded due to missing data. For the same reason, missing organic matter data for two points of Roscanvel in 2007 and one point of Glénan in 2010 as well as root biomass for the three points of Sainte-Marguerite in 2007 were estimated using k-Nearest neighbour imputation. This prevented the removal of entire sites or years while giving neutral weights to these observations. Epiphytes data were missing for the whole year 2011 and thus could not be estimated in this way. Therefore, they were not included as an explanatory variable for the modelling of *Zostera marina* attributes in the redundancy analysis (see Table 1).

Environmental explanatory variables were divided into five groups to distinguish between the effects of 1) associated flora, 2) substrate conditions, 3) hydrological and climatic conditions, 4) exposure regime and 5) tidal cycles (Table 1). Within each subset, collinear variables were removed using variance inflation factors (VIF) with a threshold of 5 (Legendre & Legendre, 2012), leading to the removal of the gravel fraction of the sediment and of some of the variables characterising the distributions of temperature, salinity, currents and tidal coefficient values.

To test the null hypothesis of no variation of *Z. marina* morphological and structural traits through space and time, a multivariate analysis of variance (MANOVA) was computed by redundancy analysis (RDA; Rao 1964) on the standardized biometric variables and

tested by permutations (Legendre & Anderson, 1999; Legendre & Legendre, 2012). Sites and years were coded by Helmert contrasts (Legendre & Gauthier, 2014) and homogeneity of multivariate dispersions was tested at the α = 0.05 significance level prior to this analysis (Anderson, 2006). This method allows testing and quantifying spatial and temporal variations along with their interaction, which estimates if temporal variations are similar across all sites; or expressed differently, if the spatial patterns are constant through time.

Sampling units in which *Z. marina* shared similar morphological and structural traits were identified using K-means clustering (Hartigan & Wong, 1979; MacQueen, 1967). Clustering was performed on the standardized *Z. marina* data and optimal number of groups between 2 and n/2 (n=120 samples) was identified using the Caliński and Harabasz criterion (1974). Variation of *Z. marina*'s characteristics in space and time were illustrated through the mapping of the groups so defined in an interaction map as proposed by Legendre et al. (2010). Patterns of trait variation were also visualised using Principal Component Analysis (PCA) on the standardized variables. A particular focus was given to the co-variation between *Zostera marina* densities and its above- and belowground biomass, as it can be indicative of the processes underlying seagrass demography (Cabaço et al., 2013, 2007; Vieira et al., 2015, 2018).

Redundancy analysis (RDA) was used on the standardized *Z. marina* traits to relate the variation of *Z. marina* morphological and structural features to the environmental variables. Stepwise selection based on adjusted coefficient of multiple determination (R^2_{adj}) was applied to the explanatory data within each of the five sets of environmental variables previously described (Table 1; Blanchet et al., 2008). Variation partitioning was used to quantify the individual and shared effects of each set of environmental variables (Peres-Neto et al., 2006). A MANOVA was then performed on the residuals of the RDA model using all sets of selected environmental variables together to evaluate if all the spatial and temporal structures of the *Z. marina* attributes were explained by our environmental dataset or if we missed important variables constraining *Zostera marina* development.

MARS3D hydrological data were extracting using Python. All other statistical analyses were conducted using R (R Core Team, 2018) and relied on the G2Sd (Fournier et al., 2014), VIM (Templ et al., 2015) and vegan (Oksanen et al., 2016) packages. All the data used in the study, in addition to R scripts to reproduce the analyses and the figures, can be found online at http://doi.org/10.5281/zenodo.5018108.

5. RESULTS

5.1. Spatial and temporal variations of Zostera marina traits

Statistical evidence for a strong spatial and temporal structuring of *Z. marina* traits was found among the eight meadows and over the five years of the study (Table 2). Sites, years, and their interaction explained 72% of the total variation of *Z. marina* traits. Temporal variations, although significant, were of lesser extent (5.9%) compared to the strong signal exclusively attributable to the sites, which explained 43.2% of the total variation of *Z. marina* traits. The significant interaction between space and time explained 22.9% and indicated that differences among meadows were not constant through the years or, in other words, that temporal changes of the meadows did not concern all sites equally. The site-specific dynamics that underlie this interaction term are illustrated by the spatio-temporal map of the two K-means clusters (Figure 2A).

Separating the sampling units into two groups maximized the Caliński-Harabasz criterion compared to more groups. Saint-Malo, Sept-Iles and Glénan meadows were almost constantly in a first group (Type 1) throughout the study whereas Sainte-Marguerite, Molène and Roscanvel were generally classified in a second group (Type 2). Arcouest and Callot meadows alternated between the two states and were the main contributors to the previously mentioned space-time interaction. Type 1 (Figure 2A; light grey square) was characterised by high shoot densities, low investment in aboveground parts compared to belowground system (both in terms of biomass and length/width of the shoots and leaves) and greater root systems at the square metre scale on average (Figure 2B and S4). Type 2 (Figure 2A; black diamond) comprised sampling units with, on average, lower *Z. marina* densities and a plant investment more orientated to aboveground parts with higher above to belowground biomass ratios, higher aboveground biomass per shoot, higher leaf and sheath lengths as well as higher leaf

width (Figure 2B and S4). The number of leaves and the belowground biomass per shoot showed less variation between the two groups (Figure S4).

Overall, two main configurations emerged from K-means clustering: cases where meadows were constituted by developed shoots in low densities, and cases where meadows displayed higher densities but of smaller shoots. This conspicuous compromise between aboveground biomass per shoot and shoot density was further supported by the absence of any meadow combining high densities with high leaf biomass per shoot (Figure 3). As a result, a negative relationship between the two features of the plant arose at the regional scale. Such relationship was also suggested by the opposite direction theses variables displayed in the PCA (Figure S5). Either one or the other of these variables, but never both, seemed to be favoured according to the sites, which constitutes the main factor discriminating the meadows under study.

Rates of clonality inferred for 11 quadrats distributed across 6 meadows (Table S2) were highly variable, ranging from 0.1 to 0.8. The most clonal meadows were Sainte-Marguerite and Molène (assigned to Type 2) and Arcouest (that switched between the two types over the years) with assessed rates reaching up to 0.7 and 0.8. The lowest rates of clonality (0.1 < c < 0.3) were found in Roscanvel (also assigned to the Type 2) and Callot (assigned either to Type 1 or 2 depending on the year). Intermediate values were found in Saint-Malo, the only representative of the Type 1 for which we have genotypic data (*c* values of 0.5 and 0.6).

5.2. Effects of environmental constraints on Zostera marina traits

Among the five sets of explanatory variables, all but the one describing the associated flora (F = 1.47, p = 0.14 using 9999 permutations) explained a significant amount of *Z. marina* trait variation (R^2_{adj} between 10.4 and 27.8%; p < 0.05). Stepwise selection was then used within the four significant sets to assess the most important forcing variables in explaining the patterns of variation of *Z. marina* morphological and structural traits. Variables selected concerned sediment characteristics, seawater salinity and rainfall, atmospheric and seawater temperature, average wind and current conditions, and influence of tidal emersions (Table 1). Overall, the 13 selected variables significantly explained around 40% (R^2_{adj}) of the total variation of *Z. marina* attributes (F = 7.1, p < 0.05).

0.001 using 9999 permutations). Hydrological and climatic conditions (see Table 1) emerged as the prevailing factors governing *Z. marina* traits in variation partitioning, explaining 6.2% alone and 24.2% when considering all shared portions of variance (Figure 4). Exposure regime explained 15.1% of *Z. marina* variations, of which only 0.6% was shared with substrate characteristics. By contrast, exposure regime shared 8.8% of explained variance with hydrological and climatic variables. Substrate characteristics and tidal cycles (the latter only explaining pure temporal variations) equally explained 13.4% of *Z. marina* variations.

The variation explained by our set of environmental variables (Figure 5) matched to a large extent with the main patterns emphasized by the unconstrained ordination of Z. *marina* traits (Figure S5). In particular, sites were well discriminated by the RDA model, with the main predicted gradient corresponding to the two groups delineated by K-means clustering i.e. Type 2 (such as Roscanvel and Molène) versus Type 1 (such as Sept-Iles, Saint-Malo and Glénan; Figure 5A & B). Meadows belonging to Type 2 were associated with higher mud content, mean wind velocities, rainfall and median seawater temperatures (Figure 5C). Among Type 1 meadows, the RDA identified different environmental drivers depending on the meadows, suggesting that the same Type 1 trait combination occurs in different environments. The RDA also highlighted subtle trait differences within these Type 1 meadows according to the environment. On the one hand, higher root/rhizome biomass per square metre were found in the Sept-Iles meadow, this Type 1 being best explained by the higher average currents velocities this site is exposed to. Higher percentage of broken leaves, as well as higher epiphytic load (Figure S6) were found in Saint-Malo and Glénan on the other hand. The Type 1 trait combinations found in these two sites were best explained by the high temperature variability (number of days with frost and average daily range of temperature; Figure S7) and by the (mobile) sandy conditions experienced by the plant in these sites. Characteristics displayed by Z. marina in these two meadows were also correlated with more spring tides (tidal coefficient superior to 100) and bigger tides (higher maximum) tidal coefficient) over the six preceding months.

All spatial and temporal signals in the data were not extracted with the set of selected explanatory variables (Table 2, Residuals). Within the 59.8% of variance left unexplained,

46% (significant at α = 0.05 level) remained structured in space and time, representing 27.5% of the total variance (46% of 59.8%). Most of the spatial and temporal structure left unexplained concerned the interaction term, which remained significant within the residuals and represented 18.5% of total variance, while it represented 22.9% in the raw data. Year-to-year variations on the other hand were well accounted for by our statistical model as no pure temporal signal was found significant in the residuals. Likewise, pure spatial signal was relatively well explained by the selected variables. Indeed, residual sites variation only represented 7.7% of the original total variance, which, albeit significant, is 6 times lower that the initially detected spatial signal. Overall, if the RDA failed to accurately model the site-specific variations at the origin of the significant space-time interaction (e.g. one-off variation occurring in a specific site; see Figure 2A), it explained most of the persistent spatial differences as well as the year-to-year variations of *Z. marina* phenotypic traits over the region.

6. DISCUSSION

6.1. Extent and drivers of Zostera marina trait variability

This study provides a comprehensive view of the extent and drivers of *Zostera marina* trait variation across Brittany. This region's 500 km coast is a highly diverse environmental mosaic (Boyé et al., 2017; Gallon et al., 2017). Although its macrotidal shores (Whitfield & Elliott, 2012) span a limited thermal gradient, generally lying within a suitable range for *Z. marina* growth (Table S1 & Figure S7; Lee et al., 2007; Nejrup & Pedersen, 2008; Wilson & Lotze, 2019), this region is representative of the diversity of *Z. marina* intertidal habitats in the North-East Atlantic in terms of substrate, salinity and hydrodynamic conditions (Hily et al., 2003; Moore & Short, 2006). Brittany has been described as a hot-spot of genetic and phenotypic diversity for *Z. marina* (Becheler et al., 2010) and this is in agreement with the extensive variability in morphology, meadow structure and rates of clonality reported here at the regional and within-bed scales.

Overall, most of the persistent spatial differences and year-to-year variations of *Z. marina* traits were explained by our set of abiotic variables but the low resolution of the available environmental data likely explains the limited ability of our model to capture site-specific one-off variation (Fig S2; Gohin et al., 2020; Poppeschi et al., 2021). In agreement with

other North-East Pacific intertidal populations, substrate type and tidal variations were important predictors of Z. marina traits (Yang et al., 2013). However, trait variation was primarily explained by temperature-related variables. Despite experimental evidence (Gustafsson & Boström, 2014), algae did not appear to have a significant effect at such broad scales. This hierarchy of drivers therefore supports the tight link existing between broad-scale climate variability and intertidal seagrass populations (Rasheed & Unsworth, 2011; Thom et al., 2014). Nonetheless, it also highlights a non-negligeable role of localscale factors such as local tidal variation that may reinforce climatic constraints (Helmuth et al., 2002; Thom et al., 2014). These are often neglected in broad-scale seagrass distribution models that mostly focus on temperature and light availability (Jayathilake & Costello, 2018; Wilson & Lotze, 2019). As for other foundation species (Firth et al., 2021), better accounting for local drivers may lead to more accurate projections of seagrass distribution changes. In the future, combining finer-scale *in situ* environmental data with higher-resolution models will also be needed to improve our understanding and predictions of seagrass changes in the face of more frequent and intense extreme events (Oliver et al., 2018; Poppeschi et al., 2021).

6.2. Phenotypic strategies of *Zostera marina* in the intertidal: facilitation-maximizing versus competition-minimizing

Two main trait configurations ("Types") were delineated among the studied meadows. While most meadows were consistently in the same configuration, some (e.g. Callot and Arcouest) changed over time. The two types are opposite ends of a continuum defined by a trade-off between density and size of shoots (Figure 3). Type 1, found for instance in Glénan, Sept-Iles and Saint-Malo, matches with the typical description of intertidal or shallow *Z. marina* meadows, namely, a combination of high densities, small shoots and leaves, and a biomass investment orientated towards below- rather than aboveground parts (Krause-Jensen et al., 2000; Park et al., 2016). This trait configuration appears particularly suited to intertidal life, as it is known to enhance moisture retention, thereby reducing thermal stress (Maxwell et al., 2017), provide shade and photoprotection against high-light stress (Park et al., 2016; Schubert et al., 2015), as well as greater resistance to the erosive forces and sediment instability typical of intertidal areas (Peralta

et al., 2008). Type 1 is therefore generally considered as a facilitation-maximizing trait configuration (Fischman et al., 2019).

At the other end of the continuum, the Type 2 meadows of Sainte-Marguerite, Molène and Roscanvel have low densities of large shoots (in terms of length and aboveground biomass) and a biomass production mainly devoted to photosynthetic parts. Type 2 is more typical of deeper meadows (Krause-Jensen et al., 2000) as it maximizes photosynthetic capacity and light harvesting (Ferguson et al., 2016; Ralph et al., 2007). Indeed, larger shoots and density together increase self-shading. Hence, as light becomes more limited with depth, seagrass density tends to decrease whereas shoot length and aboveground biomass per individual exhibit the opposite pattern (Beca-Carretero et al., 2019; Enríquez et al., 2019; Ralph et al., 2007). This process is traditionally referred to as a "self-thinning" response from the plant (Yoda et al., 1963). It results from high intra-specific competition increasing ramet spacing through densitydependent mortality, density-dependent recruitment, or through density-dependent modular growth in clonal plant such as seagrass (Brun et al., 2006; de Kroon, 1993; Johnson et al., 2020; Marbà & Duarte, 2003; Olesen & Sand-Jensen, 1994; Yang et al., 2016). Self-thinning alleviates competition among remaining shoots. Type 2 therefore corresponds to a competition-minimizing trait configuration (Fischman et al., 2019).

6.3. Potential mechanisms underlying the two phenotypic strategies: an explanation through the 'stress-gradient hypothesis'

Here, we report a negative relationship between shoot size and density across meadows of similar depth as well as the unexpected presence in the intertidal of a Type 2 (competition-minimizing) strategy, where a Type 1 (facilitation-maximizing) is generally favoured (Krause-Jensen et al., 2000; Park et al., 2016). Based on similar observations in intertidal *Zostera marina* populations of the Salish sea, Yang et al. (2013) hypothesized the role of unexpected light limitations in intertidal environments. Nonetheless, such negative relationships have been reported for seagrass species along various environmental gradients, such as physical disturbance, nutrient load, temperature and salinity (Barry et al., 2017; Ferguson et al., 2016; McDonald et al., 2016; Peralta et al., 2005). More generally, the 'stress-gradient hypothesis' (SGH) predicts that facilitative

effects prevail over competitive ones under harsh conditions, while competitive effects become more important under milder environmental conditions (Callaway & Walker, 1997). Therefore, Type 1 (facilitation-maximizing) is expected under any "harsh environment" while Type 2 (competition-minimizing) would be expected in any "mild environment", where growth is not constrained by the general environment but by the single most limiting resource (Liebig, 1840). This is often light or nutrients for seagrass (Lee et al., 2007; Ralph et al., 2007).

In agreement with the SGH, Type 1 (facilitation-maximizing) was associated in our study to two sources of stress: (i) temperature/desiccation (Saint-Malo and Glénan) and (ii) hydrodynamics (Sept-Iles). Those are two of the major factors limiting the distribution of seagrass populations in shallow environments (Koch, 2001). Type 1 was associated to maximum and minimum temperatures falling outside *Z. marina* optimal range (Figure S7; Lee et al., 2007; Nejrup & Pedersen, 2008; Wilson & Lotze, 2019). It was also linked to higher occurrences and bigger spring tides that may locally reinforce climatic constraints (Helmuth et al., 2002; Thom et al., 2014). These results are therefore in line with previous studies that described the self-facilitative feedbacks promoted by a Type 1 configuration in the face of thermal stress and strong erosive forces (Maxwell et al., 2017; Peralta et al., 2008).

In contrast, Type 2 (competition-minimizing) was found in meadows exposed to lower hydrodynamic (measured by median current velocities) and lower temperature variability. Type 2 meadows were exposed to temperatures falling within the optimal range for *Z. marina* (Lee et al., 2007; Nejrup & Pedersen, 2008; Wilson & Lotze, 2019). Hence, these beds did not seem to be constrained by any other environmental variables than those with a strong link with light availability. While *in situ* light availability was quantified in this study from broad-scale satellite data, these 4-year average PAR data available at 1km resolution were all above the minimum light requirement thresholds reported in the literature for *Z. marina* (Fig. S3; Bertelli & Unsworth, 2018). However, in support of Yang's et al. (2013) hypothesis, Type 2 was found in more turbid waters (see Figure 5): it is associated to muddier sediment, higher average wind velocities potentially promoting re-suspension, and higher precipitations, which are strongly linked with riverine inputs in the area (Tréguer et al. 2014). In combination, these environmental factors may

contribute to reduce light availability, while increasing seagrass light requirements due to muddier conditions (Zabarte-Maeztu et al., 2021). However, such fine-scale variability in *in situ* water clarity is notoriously hard to retrieve from currently available satellite data (Gohin et al., 2020). This interpretation of light as the most limiting factor of these Type 2 meadows is also congruent with their high above-to-belowground biomass ratio (de Los Santos et al., 2010; Ferguson et al., 2016). Indeed, seagrass investment in above- and belowground structures is proportional to resource supply. Belowground-parts are favoured for nutrient acquisition and storage whereas aboveground-parts help maximize photosynthetic capacity under nutrient-sufficient conditions but limiting light-levels (Herbert & Fourqurean, 2009).

Overall, the distribution of Type 1 and Type 2 meadows in this study match with the predictions of the SGH. This is in agreement with previous studies supporting (1) the occurrence of self-thinning processes in *Z. marina* and other seagrass species (Brun et al., 2006; Enríquez et al., 2019; Marbà & Duarte, 2003; Olesen & Sand-Jensen, 1994; Yang et al., 2013), and (2) the congruence of *Z. marina* dynamics with the SGH (Fischman et al., 2019; Yang et al., 2016). Predictions of the SGH allow to unify previous observations of seagrass responses across different stress gradients (e.g. Barry et al., 2017; Ferguson et al., 2016; McDonald et al., 2016; Peralta et al., 2005). The SGH can therefore provide a useful simplifying scheme to predict seagrass complex context-dependent feedbacks (Maxwell et al., 2017; Yang et al., 2016) and can help generalize seagrass responses to environmental changes (Figure 6), with several implications in terms of conservation and restoration strategies.

6.4. Implications for monitoring, conservation and restoration

As the proposed phenotypic typology is directly related to the SGH, it can be used to adapt restoration efforts to environmental conditions (see Fischman et al., 2019). In terms of monitoring, the convergence toward the Type 1 (facilitation-maximizing) in markedly different environments confirms that seagrass structural traits are sensitive to changes but are not stressor-specific indicators (Roca et al., 2016). Therefore, they might be used to give an integrative portrait of seagrass status but need to be coupled to other indicators, such as remote sensing data (cover and fragmentation), physiological,

biochemical and genetic indicators, in order to identify stressor-specific responses across different spatial and temporal scales (Roca et al., 2016). Moreover, recent experimental work suggests that a salinity-stress gradient can result in a Stress-Induced Morphometric Response (SIMR) in several seagrass species with shoot density increasing with stress intensity up to a mortality-threshold (Collier et al., 2014). Here, the concordance of our results with the predictions from the SGH suggests that this SIMR can be generalized to a broad range of other stressors (Figure 6). As a consequence, shoot-proliferation should not be considered, alone, as a consistent indicator of good ecological status for seagrass species.

The design and efficiency of management actions are contingent on the reproductive strategy of the species and the transitory or persistent nature of the meadows (Kilminster et al., 2015). Indeed, rates of clonality (r) deeply influence the resilience and evolution of seagrass (O'Brien et al., 2018). The r ranges from a theoretical minimum of 0 for purely sexual population to a maximum of 1 for strictly clonal populations. Here, r ranged from 0.1 to 0.8, confirming the high reproductive variability of Z. marina, even across a 500km coastline (Becheler et al., 2014; Jarvis et al., 2012). Together with the significant space-time interaction highlighting substantial site-specific phenotypic variation, this high variability of r suggests that conservation efforts should focus on site-specific actions, accounting for the local specificities of the meadows, rather than broad scale measures.

Rates of clonality (r) are influenced by both investment in sexual reproduction and seedling success. Theoretically, high investment in sexual reproduction is expected for Type 1 meadows in stressful environments (Cabaço & Santos, 2010; Kim et al., 2014). Conversely, Type 2 meadows in stable conditions are expected to follow an Initial Seedling Recruitment (ISR) that should favour high clonality (Eriksson 1993), with low seedling success due to competition (Johnson et al., 2020; Yang et al., 2016). Accordingly, the prevalence of persistent clones was highest in some Type 2 meadows (e.g. Molène) but unexpectedly high contribution of sexual reproduction was found in other Type 2 meadows (e.g. Roscanvel). Combination of muddy sediments and calm hydrodynamic conditions (Figure 5) that favour seed germination (Yang et al., 2013) may contribute to the low r of these meadows, but the drivers of these unexpected values warrant further research. The Type 1 Saint-Malo population showed intermediate r and

was not the meadow with the highest contribution of sexual reproduction. This intermediate r could partly reflect the low survival of seedlings in this harsh intertidal environment (Valdemarsen et al., 2010; Yang et al., 2013), despite expectations from the SGH that facilitative effects could enhance seedling success in stressful conditions (Yang et al., 2016). It matches with the mixed-annual reproductive strategy observed in some thermally stressed environment (Jarvis et al., 2012), where only a fraction of the individuals are renewed through seed recruitment and coexists with persistent clones (Becheler et al., 2014). Overall, while variation of both *Z. marina* recruitment success (Yang et al., 2016) and phenotypic traits (this study) were shown to match predictions from the SGH, observed variation of clonality rates did not match expectations (Figure 6). If observed r did not appear randomly distributed (extremes in Type 2 meadows, and intermediate in the Type 1 Saint-Malo), more studies are needed to clarify how reproductive modes and phenotypic types are linked across stress-gradients.

Overall, we highlight two distinct phenotypic types with a strong theoretical support from the 'stress-gradient hypothesis' (Callaway & Walker, 1997). While their link with reproductive strategies needs to be elucidated, these two types already provide a useful typology to guide conservation and restoration efforts (Fischman et al., 2019). In particular, the trade-off between shoot size and density underlying these two meadow types seems to be one of the key constraints governing Z. marina phenological plasticity in intertidal environments. Indeed, the trait configuration allowing to deal with the strong hydrodynamics and climatic constraints associated with intertidal life (Type 1, facilitationmaximizing) seem incompatible with the responses usually involved in dealing with light limitation (Type 2, competition-minimizing) due to water quality degradation. This can explain the synergistic effects often reported on seagrass when increased temperature co-occur with reduced light availability and increased competition (Stockbridge et al., 2020). Here, we suggest, through predictions of the SGH, that this apparent incompatibility can be generalized to multiple stress gradients. However, how this incompatibility is resolved for different stressors combinations warrants further experimental work. For example, de los Santos et al. (2010) showed that light effects prevailed over hydrodynamic ones in intertidal Z. noltii. More experimental work is therefore needed to understand how and when the SGH can help explain and predict synergistic impacts on seagrass (Stockbridge et al., 2020). Moreover, occurrence of a given phenotypic type is likely to affect (positively or negatively) the resilience of shallow seagrass populations in the face of extreme events (e.g. Maxwell et al., 2014). As these are becoming more frequent and intense (Oliver et al., 2018), future seagrass monitoring programmes will require finer-scale environmental data in order to better apprehend the effect of these drivers of seagrass changes. Increasing hydrodynamics and climatic constraints (Harley et al., 2006) are already leading to strong negative interactive effects in conjugation with water quality degradation (Krause-Jensen et al., 2021; Lefcheck et al., 2017). Water quality policies are therefore key pressure points that need to be used to ensure the resilience of seagrass meadows (Unsworth et al., 2015), especially for intertidal and shallow populations as our results suggest they face high risks of strong cumulative effects under global change stressors.

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8. CONFLICT OF INTEREST STATEMENT

The authors declare no conflict of interest.

9. AUTHORS' CONTRIBUTIONS

A.B., O.G., R.B., C.H. and J.G. conceived the ideas; C.H., R.B., V.L.G., M.M. and J.G. collected the field data; A.B. analysed the data with substantial contributions from O.G., and from R.B. for the genetic data; A.B led the writing with contributions from all authors. All authors gave final approval for publication.

10. DATA AVAILABILITY

All the data used in the study, in addition to R scripts to reproduce the analyses and the figures, can be found online at http://doi.org/10.5281/zenodo.5018108. The complete

data from the REBENT monitoring programme (http://www.rebent.org) are available in the Quadrige database (http://envlit.ifremer.fr/resultats/base_de_donnees_quadrige) and in the database of the marine observatory of the IUEM (available upon request: https://www-iuem.univ-brest.fr/observatoire). The complete time series are available for each meadow at:

- Saint-Malo: https://wwz.ifremer.fr/surval/Donnees/Graphes-30140#/sensor/60003702
- Arcouest: https://wwz.ifremer.fr/surval/Donnees/Graphes-30140#/sensor/60003701
- Sept-Iles: https://wwz.ifremer.fr/surval/Donnees/Graphes-30140#/sensor/60003700
- Callot: https://wwz.ifremer.fr/surval/Donnees/Graphes-30140#/sensor/60003703
- Sainte-Marguerite: https://wwz.ifremer.fr/surval/Donnees/Graphes-30140#/sensor/60003704
- Molène: https://wwz.ifremer.fr/surval/Donnees/Graphes-30140#/sensor/60003705
- Roscanvel : https://wwz.ifremer.fr/surval/Donnees/Graphes-30140#/sensor/60003706
- Glénan : https://wwz.ifremer.fr/surval/Donnees/Graphes-30140#/sensor/60003707

11. REFERENCES

- Airoldi, L., & Beck, M. W. (2007). Loss, status and trends for coastal marine habitats of Europe. *Oceanography and Marine Biology: An Annual Review*, 45, 357–405. doi: 10.1201/9781420050943.ch7
- Anderson, M. J. (2006). Distance-based tests for homogeneity of multivariate dispersions. *Biometrics*, 62(1), 245–253. doi: 10.1111/j.1541-0420.2005.00440.x
- Balestri, E., Cinelli, F., & Lardicci, C. (2003). Spatial variation in *Posidonia oceanica* structural, morphological and dynamic features in a northwestern Mediterranean coastal area: A multi-

scale analysis. Marine Ecology Progress Series, 250, 51–60. doi: 10.3354/meps250051

- Barbier, E. B., Hacker, S. D., Kennedy, C., Koch, E. W., Stier, A. C., & Silliman, B. R. (2011). The value of estuarine and coastal ecosystem services. *Ecological Monographs*, *81*(2), 169–193. doi: 10.1890/10-1510.1
- Barry, S. C., Jacoby, C. A., & Frazer, T. K. (2017). Environmental influences on growth and morphology of *Thalassia testudinum*. *Marine Ecology Progress Series*, *570*, 57–70.
- Beca-Carretero, P., Stanschewski, C. S., Julia-Miralles, M., Sanchez-Gallego, A., & Stengel, D. B. (2019). Temporal and depth-associated changes in the structure, morphometry and production of near-pristine *Zostera marina* meadows in western Ireland. *Aquatic Botany*, *155*, 5–17. doi: 10.1016/j.aquabot.2019.02.003
- Becheler, R., Benkara, E., Moalic, Y., Hily, C., & Arnaud-Haond, S. (2014). Scaling of processes shaping the clonal dynamics and genetic mosaic of seagrasses through temporal genetic monitoring. *Heredity*, *112*(2), 114–121. doi: 10.1038/hdy.2013.82
- Becheler, R., Diekmann, O., Hily, C., Moalic, Y., & Arnaud-Haond, S. (2010). The concept of population in clonal organisms: mosaics of temporally colonized patches are forming highly diverse meadows of *Zostera marina* in Brittany. *Molecular Ecology*, *19*, 2394–2407. doi: 10.1111/j.1365-294X.2010.04649.x
- Becheler, R., Masson, J. P., Arnaud-Haond, S., Halkett, F., Mariette, S., Guillemin, M. L., Valero,
 M., Destombe, C., & Stoeckel, S. (2017). ClonEstiMate, a Bayesian method for quantifying rates of clonality of populations genotyped at two-time steps. *Molecular Ecology Resources*, 17(6), e251–e267. doi: 10.1111/1755-0998.12698
- Bertelli, C. M., & Unsworth, R. K. F. (2018). Light stress responses by the eelgrass, *Zostera marina* (L). *Frontiers in Environmental Science*, *6*, 1–13. doi: 10.3389/fenvs.2018.00039
- Blanchet, F. G., Legendre, P., & Borcard, D. (2008). Forward selection of explanatory variables. *Ecology*, *89*(9), 2623–2632. doi: 10.1890/07-0986.1

Boyé, A., Legendre, P., Grall, J., & Gauthier, O. (2017). Constancy despite variability: Local and

regional macrofaunal diversity in intertidal seagrass beds. *Journal of Sea Research*, 130, 107–122. doi: 10.1016/j.seares.2017.06.004

- Brun, F. G., Vergara, J. J., Peralta, G., García-Sánchez, M., Hernández, I., & Pérez-Lloréns, J. L. (2006). Clonal building, simple growth rules and phylloclimate as key steps to develop functional-structural seagrass models. *Marine Ecology Progress Series*, 323, 133–148. doi: 10.3354/meps323133
- Cabaço, S., Apostolaki, E. T., García-Marín, P., Gruber, R., Hernández, I., Martínez-Crego, B., Mascaró, O., Pérez, M., Prathep, A., Robinson, C., Romero, J., Schmidt, A. L., Short, F. T., van Tussenbroek, B. I., & Santos, R. (2013). Effects of nutrient enrichment on seagrass population dynamics: evidence and synthesis from the biomass-density relationships. *Journal of Ecology*, *101*(6), 1552–1562. doi: 10.1111/1365-2745.12134
- Cabaço, S., Machás, R., & Santos, R. (2007). Biomass-density relationships of the seagrass *Zostera noltii*: A tool for monitoring anthropogenic nutrient disturbance. *Estuarine, Coastal and Shelf Science*, *74*, 557–564. doi: 10.1016/j.ecss.2007.05.029
- Cabaço, S., & Santos, R. (2010). Reproduction of the eelgrass *Zostera marina* at the species southern distributional limit in the Eastern Atlantic. *Marine Ecology*, *31*(2), 300–308. doi: 10.1111/j.1439-0485.2009.00331.x
- Caliński, T., & Harabasz, J. (1974). A dendrite method for cluster analysis. *Communications in Statistics-Theory and Methods*, *3*(1), 1–27.
- Callaway, R. M., & Walker, L. R. (1997). Competition and facilitation: a synthetic approach to interactions in plant communities. *Ecology*, *78*, 1958–1965. doi: 10.1890/0012-9658(1997)078[1958:CAFASA]2.0.CO;2
- Collier, C. J., Villacorta-Rath, C., van Dijk, K., Takahashi, M., & Waycott, M. (2014). Seagrass Proliferation Precedes Mortality during Hypo-Salinity Events: A Stress-Induced Morphometric Response. *PLoS ONE*, *9*(4), e94014. doi: 10.1371/journal.pone.0094014

de Kroon, H. (1993). Competition between Shoots in Stands of Clonal Plants. Plant Species

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Biology, 8(2–3), 85–94. doi: 10.1111/j.1442-1984.1993.tb00060.x

- de Los Santos, C. B., Brun, F. G., Bouma, T. J., Vergara, J. J., & Pérez-Lloréns, J. L. (2010). Acclimation of seagrass *Zostera noltii* to co-occurring hydrodynamic and light stresses. *Marine Ecology Progress Series*, *398*, 127–135. doi: 10.3354/meps08343
- de los Santos, C. B., Krause-Jensen, D., Alcoverro, T., Marbà, N., Duarte, C. M., van Katwijk, M.
 M., Pérez, M., Romero, J., Sánchez-Lizaso, J. L., Roca, G., Jankowska, E., Pérez-Lloréns, J. L.,
 Fournier, J., Montefalcone, M., Pergent, G., Ruiz, J. M., Cabaço, S., ... Santos, R. (2019).
 Recent trend reversal for declining European seagrass meadows. *Nature Communications*, *10*(1). doi: 10.1038/s41467-019-11340-4
- Duarte, C. M., Marbà, N., Krause-Jensen, D., & Sánchez-Camacho, M. (2007). Testing the predictive power of seagrass depth limit models. *Estuaries and Coasts*, *30*(4), 652–656. doi: 10.1007/BF02841962
- Duffy, J. E. (2006). Biodiversity and the functioning of seagrass ecosystems. *Marine Ecology Progress Series*, *311*, 233–250. doi: 10.3354/meps311233
- Ellison, A. M. (2019). Foundation Species, Non-trophic Interactions, and the Value of Being Common. *IScience*, *13*, 254–268. doi: 10.1016/j.isci.2019.02.020
- Enríquez, S., Olivé, I., Cayabyab, N., & Hedley, J. D. (2019). Structural complexity governs seagrass acclimatization to depth with relevant consequences for meadow production, macrophyte diversity and habitat carbon storage capacity. *Scientific Reports*, *9*(1), 1–14. doi: 10.1038/s41598-019-51248-z
- Eriksson, O. (1993). Dynamics of genets in clonal plants. *Trends in Ecology and Evolution, 8*(9), 313–316. doi: 10.1016/0169-5347(93)90237-J
- Ferguson, A. J. P., Gruber, R. K., Orr, M., & Scanes, P. (2016). Morphological plasticity in *Zostera* muelleri across light, sediment, and nutrient gradients in Australian temperate coastal lakes.
 Marine Ecology Progress Series, 556, 91–104. doi: 10.3354/meps11830

Firth, L. B., Harris, D., Blaze, J. A., Marzloff, M. P., Boyé, A., Miller, P. I., Curd, A., Vasquez, M.,

Nunn, J. D., O'Connor, N. E., Power, A. M., Mieszkowska, N., O'Riordan, R. M., Burrows, M. T., Bricheno, L. M., Knights, A. M., Nunes, F. L. D., ... Hawkins, S. J. (2021). Specific niche requirements underpin multidecadal range edge stability, but may introduce barriers for climate change adaptation. *Diversity and Distributions*, *27*(4), 668–683. doi: 10.1111/ddi.13224

- Fischman, H. S., Crotty, S. M., & Angelini, C. (2019). Optimizing coastal restoration with the stress gradient hypothesis. *Proceedings of the Royal Society B: Biological Sciences, 286*(1917), 20191978. doi: 10.1098/rspb.2019.1978
- Fournier, J., Bonnot-Courtois, C., Paris, R., Voldoire, O., & Le Vot, M. (2012). Analyses granulométriques, principes et méthodes. *CNRS, Dinard*, 99.
- Fournier, J., Gallon, R., & Paris, R. (2014). G2Sd: a new R package for the statistical analysis of unconsolidated sediments. *Geomorphologie: Relief, Processus, Environnement*, Vol. 20, pp. 73–78. doi: 10.4000/geomorphologie.10513
- Gallon, R. K., Lavesque, N., Grall, J., Labrune, C., Gremare, A., Bachelet, G., Blanchet, H., Bonifácio, P., Bouchet, V. M. P., Dauvin, J. C., Desroy, N., Gentil, F., Guerin, L., Houbin, C., Jourde, J., Laurand, S., Le Duff, M., ... Gauthier, O. (2017). Regional and latitudinal patterns of soft-bottom macrobenthic invertebrates along French coasts: Results from the RESOMAR database. *Journal of Sea Research*, *130*, 96–106. doi: 10.1016/j.seares.2017.03.011
- Godet, L., Fournier, J., Van Katwijk, M. M., Olivier, F., Le Mao, P., & Retière, C. (2008). Before and after wasting disease in common eelgrass *Zostera marina* along the French Atlantic coasts: A general overview and first accurate mapping. *Diseases of Aquatic Organisms*, *79*(3), 249– 255. doi: 10.3354/dao01897
- Gohin, F., Bryère, P., Lefebvre, A., Sauriau, P. G., Savoye, N., Vantrepotte, V., Bozec, Y., Cariou, T.,
 Conan, P., Coudray, S., Courtay, G., Françoise, S., Goffart, A., Fariñas, T. H., Lemoine, M.,
 Piraud, A., Raimbault, P., & Rétho, M. (2020). Satellite and in situ monitoring of chl-a,
 turbidity, and total suspended matter in coastal waters: Experience of the year 2017 along
 the french coasts. *Journal of Marine Science and Engineering*, 8(9), 665. doi:

This article is protected by copyright. All rights reserved

10.3390/jmse8090665

- Gustafsson, C., & Boström, C. (2014). Algal mats reduce eelgrass (*Zostera marina* L.) growth in mixed and monospecific meadows. *Journal of Experimental Marine Biology and Ecology*, 461, 85–92. doi: 10.1016/j.jembe.2014.07.020
- Harley, C. D. G., Hughes, A. R., Hultgren, K. M., Miner, B. G., Sorte, C. J. B., Thornber, C. S.,
 Rodriguez, L. F., Tomanek, L., & Williams, S. L. (2006). The impacts of climate change in coastal marine systems. *Ecology Letters*, 9(2), 228–241. doi: 10.1111/j.1461-0248.2005.00871.x
- Hartigan, J. A., & Wong, M. A. (1979). Algorithm AS 136: A k-means clustering algorithm. *Applied Statistics*, 100–108.
- Helmuth, B., Harley, C. D. G., Halpin, P. M., O'Donnell, M., Hofmann, G. E., & Blanchette, C. A. (2002). Climate change and latitudinal patterns of intertidal thermal stress. *Science*, 298(5595), 1015–1017. doi: 10.1126/science.1076814
- Herbert, D. A., & Fourqurean, J. W. (2009). Phosphorus availability and salinity control productivity and demography of the seagrass *Thalassia testudinum* in Florida Bay. *Estuaries and Coasts*, *32*(1), 188–201. doi: 10.1007/s12237-008-9116-x
- Hily, C., van Katwijk, M. M., & Den Hartog, C. (2003). The seagrasses of Western Europe. In Green, E. P. & Short, F. T. (Eds.), *World atlas of seagrasses* (pp. 38–47). University of California Press.
- Hughes, A. R., & Stachowicz, J. J. (2011). Seagrass genotypic diversity increases disturbance response via complementarity and dominance. *Journal of Ecology*, *99*(2), 445–453. doi: 10.1111/j.1365-2745.2010.01767.x
- Jarvis, J. C., Moore, K. A., & Kenworthy, W. J. (2012). Characterization and ecological implication of eelgrass life history strategies near the species' southern limit in the western North Atlantic. *Marine Ecology Progress Series*, 444, 43–56. doi: 10.3354/meps09428

Jayathilake, D. R. M., & Costello, M. J. (2018). A modelled global distribution of the seagrass

biome. *Biological Conservation*, 226, 120–126. doi: https://doi.org/10.1016/j.biocon.2018.07.009

- Johnson, A. J., Orth, R. J., & Moore, K. A. (2020). The role of sexual reproduction in the maintenance of established *Zostera marina* meadows. *Journal of Ecology*, *108*(3), 945–957. doi: 10.1111/1365-2745.13362
- Kaldy, J. E., & Lee, K. S. (2007). Factors controlling *Zostera marina* L. growth in the eastern and western Pacific Ocean: Comparisons between Korea and Oregon, USA. *Aquatic Botany*, *87*(2), 116–126. doi: 10.1016/j.aquabot.2007.03.008
- Kilminster, K., McMahon, K., Waycott, M., Kendrick, G. A., Scanes, P., McKenzie, L., O'Brien, K. R.,
 Lyons, M., Ferguson, A., Maxwell, P., Glasby, T., & Udy, J. (2015). Unravelling complexity in seagrass systems for management: Australia as a microcosm. *Science of the Total Environment*, 534, 97–109. doi: 10.1016/j.scitotenv.2015.04.061
- Kim, S. H., Kim, J. H., Park, S. R., & Lee, K. S. (2014). Annual and perennial life history strategies of *Zostera marina* populations under different light regimes. *Marine Ecology Progress Series*, 509, 1–13. doi: 10.3354/meps10899
- Koch, E. W. (2001). Beyond light: Physical, geological, and geochemical parameters as possible submersed aquatic vegetation habitat requirements. *Estuaries*, *24*(1), 1–17. doi: 10.1007/BF02693942
- Krause-Jensen, D., Carstensen, J., Nielsen, S. L., Dalsgaard, T., Christensen, P. B., Fossing, H., & Rasmussen, M. B. (2011). Sea bottom characteristics affect depth limits of eelgrass *Zostera marina*. *Marine Ecology Progress Series*, 425, 91–102. doi: 10.3354/meps09026
- Krause-Jensen, D., Duarte, C. M., Sand-Jensen, K., & Carstensen, J. (2021). Century-long records reveal shifting challenges to seagrass recovery. *Global Change Biology*, 27(3), 563–575. doi: https://doi.org/10.1111/gcb.15440
- Krause-Jensen, D., Middelboe, A. L., Sand-Jensen, K., & Christensen, P. B. (2000). Eelgrass, *Zostera marina*, growth along depth gradients : upper boundaries of the variation as a

powerful predictive tool. Oikos, 91(2), 233-244. doi: 10.1034/j.1600-0706.2001.910204.x

- Lazure, P., Garnier, V., Dumas, F., Herry, C., & Chifflet, M. (2009). Development of a hydrodynamic model of the Bay of Biscay. Validation of hydrology. *Continental Shelf Research*, *29*(8), 985–997. doi: 10.1016/j.csr.2008.12.017
- Lee, K. S., Park, S. R., & Kim, Y. K. (2007). Effects of irradiance, temperature, and nutrients on growth dynamics of seagrasses: A review. *Journal of Experimental Marine Biology and Ecology*, *350*(1–2), 144–175. doi: 10.1016/j.jembe.2007.06.016
- Lefcheck, J. S., Wilcox, D. J., Murphy, R. R., Marion, S. R., & Orth, R. J. (2017). Multiple stressors threaten the imperiled coastal foundation species eelgrass (*Zostera marina*) in Chesapeake Bay, USA. *Global Change Biology*, *23*(9), 3474–3483. doi: 10.1111/gcb.13623
- Legendre, P., & Anderson, M. J. (1999). Distance-Based Redundancy Analysis: Testing Multispecies Responses in Multifactorial Ecological Experiments. *Ecological Monographs*, 69(1), 1–24. doi: 10.2307/2657192
- Legendre, P., Cáceres, M. De, & Borcard, D. (2010). Community surveys through space and time: testing the space--time interaction in the absence of replication. *Ecology*, *91*(1), 262–272. doi: 10.1890/09-0199.1
- Legendre, P., & Gauthier, O. (2014). Statistical methods for temporal and space-time analysis of community composition data. *Proceedings of the Royal Society of London B: Biological Sciences*, 281, 20132728. doi: 10.1098/rspb.2013.2728

Legendre, P., & Legendre, L. (2012). Numerical ecology, 3rd English edition (Elsevier, Ed.).

- Liebig, J. (1840). Die organische Chemie in ihrer Anwendung auf Agricultur und Physiologie. In *Braunschweig*. F. Vieweg und Sohn.
- Lotze, H. K., Lenihan, H. S., Bourque, B. J., Bradbury, R. H., Cooke, R. G., Kay, M. C., Kidwell, S. M., Kirby, M. X., Peterson, C. H., & Jackson, J. B. C. (2006). Depletion, Degradation, and Recovery Potential of Estuaries and Coastal Seas. *Science*, *312*(5781), 1806–1809. doi: 10.1126/science.1128035

- MacQueen, J. (1967). Some methods for classification and analysis of multivariate observations. *Proceedings of the Fifth Berkeley Symposium on Mathematical Statistics and Probability*, 1, 281–296.
- Marbà, N., & Duarte, C. M. (2003). Scaling of ramet size and spacing in seagrasses: Implications for stand development. *Aquatic Botany*, 77(2), 87–98. doi: 10.1016/S0304-3770(03)00079-2
- Marshall, D. R., & Weir, B. S. (1979). Maintenance of genetic variation in apomictic plant populations. *Heredity*, *42*(2), 159–172. doi: 10.1038/hdy.1979.20
- Martínez-Crego, B., Vergés, A., Alcoverro, T., & Romero, J. (2008). Selection of multiple seagrass indicators for environmental biomonitoring. *Marine Ecology Progress Series*, *361*, 93–109. doi: 10.3354/meps07358
- Maxwell, P. S., Eklöf, J. S., van Katwijk, M. M., O'Brien, K. R., de la Torre-Castro, M., Boström, C., Bouma, T. J., Krause-Jensen, D., Unsworth, R. K. F., van Tussenbroek, B. I., & van der Heide, T. (2017). The fundamental role of ecological feedback mechanisms for the adaptive management of seagrass ecosystems a review. *Biological Reviews*, *92*(3), 1521–1538. doi: 10.1111/brv.12294
- Maxwell, P. S., Pitt, K. A., Burfeind, D. D., Olds, A. D., Babcock, R. C., & Connolly, R. M. (2014).
 Phenotypic plasticity promotes persistence following severe events: Physiological and morphological responses of seagrass to flooding. *Journal of Ecology*, *102*(1), 54–64. doi: 10.1111/1365-2745.12167
- McDonald, A. M., Prado, P., Heck Jr, K. L., Fourqurean, J. W., Frankovich, T. A., Dunton, K. H., &
 Cebrian, J. (2016). Seagrass growth, reproductive, and morphological plasticity across environmental gradients over a large spatial scale. *Aquatic Botany*, *134*, 87–96.
- Moore, K. A., & Short, F. T. (2006). *Zostera*: Biology, Ecology and Management. In *Seagrasses: biology, ecology and conservation* (pp. 361–386). doi: 10.1007/978-1-4020-2983-7_16
- Moreno-Marín, F., Brun, F. G., & Pedersen, M. F. (2018). Additive response to multiple environmental stressors in the seagrass *Zostera marina* L. *Limnology and Oceanography*,

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63(4), 1528–1544. doi: 10.1002/lno.10789

- Nejrup, L. B., & Pedersen, M. F. (2008). Effects of salinity and water temperature on the ecological performance of *Zostera marina*. *Aquatic Botany*, *88*(3), 239–246. doi: 10.1016/j.aquabot.2007.10.006
- O'Brien, K. R., Waycott, M., Maxwell, P., Kendrick, G. A., Udy, J. W., Ferguson, A. J. P., Kilminster,
 K., Scanes, P., McKenzie, L. J., McMahon, K., Adams, M. P., Samper-Villarreal, J., Collier, C.,
 Lyons, M., Mumby, P. J., Radke, L., Christianen, M. J. A., & Dennison, W. C. (2018). Seagrass ecosystem trajectory depends on the relative timescales of resistance, recovery and disturbance. *Marine Pollution Bulletin*, 134, 166–176. doi: 10.1016/j.marpolbul.2017.09.006
- Oksanen, J., Blanchet, F. G., Kindt, R., Legendre, P., O'Hara, R. B., Simpson, G. L., Solymos, P., Stevens, M. H. H., & Wagner, H. H. (2016). *vegan: Community Ecology Package. R package version 2.3-4. http://CRAN.R-project.org/package=vegan.* Retrieved from http://cran.r-project.org/package=vegan
- Olesen, B., & Sand-Jensen, K. (1994). Biomass-density patterns in the temperate seagrass *Zostera marina*. *Marine Ecology Progress Series*, *109*, 283–292. doi: 10.3354/meps111283
- Oliver, E. C. J., Donat, M. G., Burrows, M. T., Moore, P. J., Smale, D. A., Alexander, L. V, Benthuysen, J. A., Feng, M., Gupta, A. Sen, Hobday, A. J., & others. (2018). Longer and more frequent marine heatwaves over the past century. *Nature Communications*, *9*(1), 1324.
- Orth, R. J., Carruthers, T. J. B., Dennison, W. C., Duarte, C. M., James, W., Heck Jr., K. L., Hughes, A. R., Kendrick, G. A., Kenworthy, W. J., Olyarnik, S., Short, F. T., Waycott, M., & Williams, S. L. (2006). Global Crisis for Seagrass Ecosystems. *BioScience*, 56(12), 987–996. doi: 10.1641/0006-3568(2006)56[987:AGCFSE]2.0.CO;2
- Park, S. R., Kim, S., Kim, Y. K., Kang, C.-K., & Lee, K.-S. (2016). Photoacclimatory Responses of *Zostera marina* in the Intertidal and Subtidal Zones. *PLOS ONE*, *11*(5), e0156214. doi: 10.1371/journal.pone.0156214

Peralta, G., Brun, F. G., Hernández, I., Vergara, J. J., & Pérez-Lloréns, J. L. (2005). Morphometric

variations as acclimation mechanisms in *Zostera noltii* beds. *Estuarine, Coastal and Shelf Science, 64*(2–3), 347–356. doi: 10.1016/j.ecss.2005.02.027

- Peralta, G., Van Duren, L. A., Morris, E. P., & Bouma, T. J. (2008). Consequences of shoot density and stiffness for ecosystem engineering by benthic macrophytes in flow dominated areas: a hydrodynamic flume study. *Marine Ecology Progress Series, 368*, 103–115.
- Peres-Neto, P. R., Legendre, P., Dray, S., & Borcard, D. (2006). Variation partitioning of species data matrices: estimation and comparison of fractions. *Ecology*, *87*(10), 2614–2625. doi: 10.1890/0012-9658(2006)87[2614:VPOSDM]2.0.CO;2
- Poppeschi, C., Charria, G., Goberville, E., Rimmelin-Maury, P., Barrier, N., Petton, S., Unterberger, M., Grossteffan, E., Repecaud, M., Quéméner, L., Theetten, S., Le Roux, J.-F., & Tréguer, P. (2021). Unraveling Salinity Extreme Events in Coastal Environments: A Winter Focus on the Bay of Brest . *Frontiers in Marine Science*, Vol. 8, p. 966. Retrieved from https://www.frontiersin.org/article/10.3389/fmars.2021.705403
- Populus, J., Vasquez, M., Albrecht, J., Manca, E., Agnesi, S., Al Hamdani, Z., Andersen, J., Annunziatellis, A., Bekkby, T., Bruschi, A., Doncheva, V., Drakopoulou, V., Duncan, G., Inghilesi, R., Kyriakidou, C., Lalli, F., Lillis, H., ... Tunesi, L. (2017). *EUSeaMap. A European broad-scale seabed habitat map.* doi: 10.13155/49975
- Potouroglou, M., Kenyon, E. J., Gall, A., Cook, K. J., & Bull, J. C. (2014). The roles of flowering, overwinter survival and sea surface temperature in the long-term population dynamics of *Zostera marina* around the Isles of Scilly, UK. *Marine Pollution Bulletin*, *83*(2), 500–507. doi: 10.1016/j.marpolbul.2014.03.035
- R Core Team. (2018). R: A Language and Environment for Statistical Computing. *R Foundation for Statistical Computing, Vienna, Austria.*, R version 3.5.0 (2018-04-23). Retrieved from https://www.r-project.org/
- Ralph, P. J., Durako, M. J., Enríquez, S., Collier, C. J., & Doblin, M. A. (2007). Impact of light limitation on seagrasses. *Journal of Experimental Marine Biology and Ecology*, *350*(1–2), 176–193. doi: 10.1016/j.jembe.2007.06.017

This article is protected by copyright. All rights reserved

Rao, R. (1964). The Use and Interpretation of Principal Component Analysis in Applied Research. *The Indian Journal of Statistics*, *26*(4), 329–358.

- Rasheed, M. A., & Unsworth, R. K. F. (2011). Long-term climate-associated dynamics of a tropical seagrass meadow: implications for the future. *Marine Ecology Progress Series*, *422*, 93–103. doi: 10.3354/meps08925
- Reusch, T. B. H., Ehlers, A., Hämmerli, A., & Worm, B. (2005). Ecosystem recovery after climatic extremes enhanced by genotypic diversity. *Proceedings of the National Academy of Sciences*, *102*(8), 2826–2831. doi: 10.1073/pnas.0500008102
- Roca, G., Alcoverro, T., Krause-Jensen, D., Balsby, T. J. S., van Katwijk, M. M., Marba, N., Santos,
 R., Arthur, R., Mascara, O., Fernandez-Torquemada, Y., Perez, M., Duarte, C. M., & Romero,
 J. (2016). Response of seagrass indicators to shifts in environmental stressors: A global
 review and management synthesis. *Ecological Indicators*, 63, 310–323. doi: 10.1016/j.ecolind.2015.12.007
- Schubert, N., Colombo-Pallota, M. F., & Enríquez, S. (2015). Leaf and canopy scale characterization of the photoprotective response to high-light stress of the seagrass *Thalassia testudinum. Limnology and Oceanography*. doi: 10.1002/lno.10024
- SHOM. (2014). SHOMAR Édition 2014-2015, disponible sur http://www.shom.fr/lesproduits/bases-de-donnees-numeriques/maree-et-courant/shomar/.
- Short, F., Carruthers, T., Dennison, W., & Waycott, M. (2007). Global seagrass distribution and diversity: A bioregional model. *Journal of Experimental Marine Biology and Ecology*, 350(1–2), 3–20. doi: 10.1016/j.jembe.2007.06.012
- Stockbridge, J., Jones, A. R., & Gillanders, B. M. (2020). A meta-analysis of multiple stressors on seagrasses in the context of marine spatial cumulative impacts assessment. *Scientific Reports*, *10*(1), 1–11. doi: 10.1038/s41598-020-68801-w
- Stoeckel, S., & Masson, J. P. (2014). The exact distributions of FIS under partial asexuality in small finite populations with mutation. *PLoS ONE*, *9*(1). doi: 10.1371/journal.pone.0085228

Templ, M., Alfons, A., Kowarik, A., & Prantner, B. (2015). VIM: Visualization and Imputation of Missing Values. R package version 4.4.1. http://CRAN.R-project.org/package=VIM.

- Thom, R., Southard, S., & Borde, A. (2014). Climate-linked Mechanisms Driving Spatial and Temporal Variation in Eelgrass (*Zostera marina* L.) Growth and Assemblage Structure in Pacific Northwest Estuaries, U.S.A. *Journal of Coastal Research*, *68*(sp1), 1–11. doi: 10.2112/si68-001.1
- Tréguer, P., Goberville, E., Barrier, N., L'Helguen, S., Morin, P., Bozec, Y., Rimmelin-Maury, P., Czamanski, M., Grossteffan, E., Cariou, T., Répécaud, M., & Quéméner, L. (2014). Large and local-scale influences on physical and chemical characteristics of coastal waters of Western Europe during winter. *Journal of Marine Systems*, *139*(November), 79–90. doi: 10.1016/j.jmarsys.2014.05.019
- Unsworth, R. K. F., Collier, C. J., Waycott, M., Mckenzie, L. J., & Cullen-Unsworth, L. C. (2015). A framework for the resilience of seagrass ecosystems. *Marine Pollution Bulletin*, 100(1), 34–46. doi: 10.1016/j.marpolbul.2015.08.016
- Valdemarsen, T., Canal-Verges, P., Kristensen, E., Holmer, M., Kristiansen, M. D., & Flindt, M. R. (2010). Vulnerability of *Zostera marina* seedlings to physical stress. *Marine Ecology Progress Series*, 418, 119–130. doi: 10.3354/meps08828
- Van Lent, F., & Verschuure, J. M. (1994a). Intraspecific variability of *Zostera marina* L. (eelgrass) in the estuaries and lagoons of the southwestern Netherlands. I. Population dynamics. *Aquatic Botany*, *48*(1), 31–58. doi: 10.1016/0304-3770(94)90072-8
- Van Lent, F., & Verschuure, J. M. (1994b). Intraspecific variability of *Zostera marina* L. (eelgrass) in the estuaries and lagoons of the southwestern Netherlands. II. Relation with environmental factors. *Aquatic Botany*, *48*(1), 59–75. doi: 10.1016/0304-3770(94)90072-8
- Verheyen, K., De Frenne, P., Baeten, L., Waller, D. M., Hédl, R., Perring, M. P., Blondeel, H., Brunet, J., Chudomelová, M., Decocq, G., De Lombaerde, E., Depauw, L., Dirnböck, T., Durak, T., Eriksson, O., Gilliam, F. S., Heinken, T., ... Bernhardt-Römermann, M. (2017). Combining biodiversity resurveys across regions to advance global change research. *BioScience*, *67*(1),

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73-83. doi: 10.1093/biosci/biw150

- Vieira, V. M. N. C. S., Leitão, F., & Mateus, M. (2015). Biomass-density data analysis: A comment on Cabaço et al. (2013). *Journal of Ecology*, *103*(3), 537–540. doi: 10.1111/1365-2745.12294
- Vieira, V. M. N. C. S., Lopes, I. E., & Creed, J. C. (2018). The biomass-density relationship in seagrasses and its use as an ecological indicator. *BMC Ecology*, 18(1), 1–16. doi: 10.1186/s12898-018-0200-1
- Waycott, M., Duarte, C. M., Carruthers, T. J. B., Orth, R. J., Dennison, W. C., Olyarnik, S., Calladine, A., Fourqurean, J. W., Heck Jr., K. L., Hughes, A. R., Kendrick, G. A., Kenworthy, W. J., Short, F. T., & Williams, S. L. (2009). Accelerating loss of seagrasses across the globe threatens coastal ecosystems. *Proceedings of the National Academy of Sciences*, *106*(30), 12377–12381. doi: 10.1073/pnas.0905620106
- Whitfield, A., & Elliott, M. (2012). Ecosystem and Biotic Classifications of Estuaries and Coasts. In *Treatise on Estuarine and Coastal Science* (Vol. 1, pp. 99–124). doi: 10.1016/B978-0-12-374711-2.00108-X
- Wilson, K. L., & Lotze, H. K. (2019). Climate change projections reveal range shifts of eelgrass *Zostera marina* in the Northwest Atlantic. *Marine Ecology Progress Series*, 620, 47–62. doi: 10.3354/meps12973
- Witman, J. D., Lamb, R. W., & Byrnes, J. E. K. (2015). Towards an integration of scale and complexity in marine ecology. *Ecological Monographs*, *85*(4), 475–504. doi: 10.1890/14-2265.1
- Yang, S., HilleRisLambers, J., & Ruesink, J. L. (2016). Reversal of intraspecific interactions by an ecosystem engineer leads to variable seedling success along a stress gradient. *Marine Ecology Progress Series*, 543, 163–171. doi: 10.3354/meps11561
- Yang, S., Wheat, E. E., Horwith, M. J., & Ruesink, J. L. (2013). Relative Impacts of Natural Stressors on Life History Traits Underlying Resilience of Intertidal Eelgrass (*Zostera marina* L.). *Estuaries and Coasts*, 36(5), 1006–1013. doi: 10.1007/s12237-013-9609-0

- Yoda, K., Kira, T., Ogawa, H., & Hozumi, K. (1963). Self-thinning in overcrowded pure stands under cultivated and natural conditions (Intraspecific competition among higher plants XI). *Journal of the Institute of Polythecnics, Osaka City University, Series D, 14*, 107–129.
- Zabarte-Maeztu, I., Matheson, F. E., Manley-Harris, M., Davies-Colley, R. J., & Hawes, I. (2021). Interaction of substrate muddiness and low irradiance on seagrass: A mesocosm study of Zostera muelleri. *Aquatic Botany*, *175*. doi: 10.1016/j.aquabot.2021.103435
- Zimmerman, R., Hill, V., Jinuntuya, M., Celebi, B., Ruble, D., Smith, M., Cedeno, T., & Swingle, W. (2017). Experimental impacts of climate warming and ocean carbonation on eelgrass
 Zostera marina. *Marine Ecology Progress Series*, *566*, 1–15. doi: 10.3354/meps12051



Figure 1. Localities of the eight *Zostera marina* meadows (black stars) monitored by the RÉseau BENThique (REBENT) in Brittany in 2007 and from 2009 to 2012. Each site was sampled for *Zostera marina* phenotypic traits as well as for drifting algae biomass (assessed with two 0.05 m² quadrats). The biomass of associated epiphytes was quantified from 10 additional shoots haphazardly collected within the meadow. A core was also sampled for sediment characterisation. This survey was performed within each site at three points located 200 m apart, as shown for the Glénan meadow.



Figure 2. A. Spatio-temporal map of the membership of each sampling unit to the groups defined by K-mean clustering performed on the standardized *Zostera marina* traits. The two groups are represented with different point shapes and intensities of grey. **B.** Estimated probability density function of key *Z. marina* traits differentiating these two groups with the vertical dashed-lines representing the mean of the variables for each group. Tick marks across the bottom of each plot indicate observed data points. See Table 1 for abbreviations and units and Figure S4 for the distribution of all the traits measured.



Figure 3. Relationship between the mean leaf biomass per shoot and the shoot density of *Zostera marina* in the 8 sites monitored during the 5 years of the study.



Figure 4. Venn diagram illustrating the results of variation partitioning of the standardized traits of the Zostera marina meadows with respect to four sets of explanatory variables: substrate conditions, exposure regime, hydrological and climatic conditions, and tidal cycles (see Table 1 for full description of each set). Only the variables selected by the stepwise procedure were used for this analysis. The redundancy analysis corresponding to this model is presented in Fig. 5. Variation is expressed as a percentage (%) of the total variation of Z. marina traits based on adjusted R². Empty fractions are negative values and may be considered as zeros. Residuals are unexplained variations.



Figure 5. Redundancy analysis of the standardized *Zostera marina* traits against the selected environmental variables (same model than in Fig. 4). The response variables and explanatory variables selected by the stepwise procedure are summarised in Table 1 along with their abbreviations and units. Scaling type 2. The first two axes represent 27.85% and 8.11% of the total variation respectively. **A.** Linear constraints scores for all samples (one point of one site at a given year) with the 95% confidence dispersion ellipses of each site. Within-site dispersions represent the temporal variability during the 5 years of the study, and trait variation among the three points sampled per year, that is explained by the model. **B.** Response variables *i.e. Zostera marina* traits. **C.** Explanatory variables *i.e.* the environmental variables selected.



Figure 6. Proposed generalisation of the phenotypic variation of *Zostera marina* across any environmental gradient based on the 'stress-gradient hypothesis' (SGH). The links between phenotypic types and reproductive strategies (bottom panel) warrants further research. Indeed, observed rates of clonality did not match expectations, but sample size was low (indicated between parenthesis), especially for Type 1 meadows (only one meadow with genotypic data).

Table 1. List of all the variables, with their abbreviations and units, used in this study. Only explanatory variables keptafter the removal of collinear variables, identified using Variance Inflation Factor, are presented here. Stepwise selectionprocedure was performed within each five sets of explanatory variables (associated flora, substrate, hydrological andclimatic conditions, exposure regime and tidal cycles) and selected variables are shown in the 5th column.‡Epiphytes data were not included as explanatory variables in the modelling of *Zostera marina* traits due to missing datafor an entire year but they were kept as they served interpretation.

		Abbreviation	Description	Units	
Response	e variables				
		leaf.biom	Leaf biomass per square meter	g.m⁻²	
		root.biom	Root biomass per square meter	g.m ⁻²	
		leaf.biom/shoot	Mean leaf biomass per shoot	g	
		root.biom/shoot	Mean root biomass per shoot	g	
			Ratio leaf to root biomass	/	
<i>Zostera marina</i> traits		density	Mean shoot density per square meter	shoot.m	1 ⁻²
		sheath.heigth	Mean sheath heigth	mm	
		leaf.length	Mean leaf length	mm	
		leaf.width	Leaf width	mm	
		leaves/shoot	Number of leaves per shoot	/	
		broken	Percent of broken leaves	%	
Explanato	ry variables			_	Selection
		and the state	Ratio between epiphytes and Zostera	,	+
		epiphytes	marina leaf biomass	/	+
Associa	ited flora	green.alg	Biomass of green algae	g.m⁻²	
		brown.alg	Biomass of brown algae	g.m⁻²	
		red.alg	Biomass of red algae	g.m⁻²	
Substrate			Sediment fraction between 63µm and	0	
		sand	1mm	%	\checkmark
		mud	Sediment fraction < 63µm	%	\checkmark
		So	Sorting index	/	
		grain.50	Median grain size	μm	
		OM	Organic matter content	%	
		sal.max	Maximum salintiy	‰	
	Marine	sal.50	Median salinity	‰	\checkmark
		sal.range	Salinity range	‰	
		T°C.sea.max	Maximum sea water temperature	°C	\checkmark
Hvdrology		T°C.sea.50	Median sea water temperature	°C	\checkmark
and		T°C.sea.range	Sea water temperature range	°C	
climate		T°C.air.max	Maximum air temperature	°C	
	Atmospheric	T°C.air.drange	Mean daily range of temperature	°C	
		frost	Number of frost days	davs	\checkmark
		rain	, Cumulative rainfall	, mm	\checkmark
		NAO	North Atlantic Oscillation index	/	
Exposure	Marine	current.50	Median current velocity	m.s ⁻¹	V
		current.min	Minimum current velocity	m.s⁻¹	
	Atmospheric	wind.max	Maximum wind force	m.s⁻¹	
		wind.mean	Mean wind force	m.s⁻¹	\checkmark
		[400	Number of days with a tidal coefficient		
Tidal cycles		coet+100	superior to 100	days	M
		coef.max	Distribution of the tidal coefficients over	/	$\overline{\mathbf{A}}$
		coef.50	the period with minimum, maximum and	, , ,	
		coef.min	median (50) values	/	
				1	

Table 2. Results from MANOVA by RDA analyses testing for spatial and temporal structures with [S] being the spatial variables (Helmert contrasts) coding for the 8 sites monitored, [T] being temporal variables coding for the 5 years of the study and [I] being the interaction terms between sites and years. [I|S+T] represents the pure effect of interaction, once the effect of spatial and temporal contrasts was removed, and likewise for the pure spatial and temporal fractions. Spatial and temporal structures were first quantified and tested on the standardized *Zostera marina* traits (Raw data). A second analysis was then performed on the residuals of the RDA modelling *Z. marina* traits with the selected environmental variables (Fig. 4 and 5) to test for remaining spatial and temporal signals once the explanatory variables effects were removed. Unadjusted R^2 are presented as percentage of total variance and *p*-values were obtained after 9999 permutations with *** < 0.001,** < 0.01 and * < 0.05. Residual unadjusted R^2 were also expressed as percentage of total variance by multiplying the R^2 with the unexplained variance of the RDA model (1- R^2adj = 59.8%)

		Raw data			Residuals		
	df	F	R^2	р	F	R^2	Р
[I S+T]	28	2.3	22.9	***	1.7	18.5	*
[S I+T]	7	17.7	43.2	***	2.7	7.7	***
[T I+S]	4	4.2	5.9	***	0.8	1.3	
[S+T+I]	39	5.3	72.0	***	1.8	27.5	**