

Supplementary material

Supplementary methods

Respirometry

The rearing tanks were custom-designed to measure metabolic rate as O₂ uptake by automated stop-flow respirometry (Steffensen, 1989), as previously described in (McKenzie et al., 2012; McKenzie, Pedersen, & Jokumsen, 2007). Briefly, each tank was fitted with a central vertical PVC pipe that was perforated around the base. It housed a submersible pump that drew in water from the perforations and delivered it out through a flexible tube fixed to the outer wall of the tank, so constantly mixing the tank water. For 45 min of every hour, fresh aerated water was pumped from a large biofiltered reservoir (Vol. approx. 100 l) into the central PVC pipe of each tank, to maintain dissolved O₂ levels close to air saturation in the water holding the sardines; the water returned to the reservoir through a standpipe overflow. The pump in the reservoir was controlled by an electrical timer, and was turned off for 15 min of each hour, at which point the water level settled at the overflow to provide a constant volume, but the water continued to be mixed by the pump in the central pipe. Each tank was fitted with an O₂ optode (Pre-Sens sturdy dipping probe, www.presen.de) attached to an O₂ meter (Pre-Sens OXY-10 mini), which used the manufacturers software to record the linear declines in O₂ saturation in each tank, due to consumption by the sardines. Water O₂ saturation never fell below 70% during the 15 min of closed cycle respirometry and was rapidly restored when the tanks received a flow of aerated water from the reservoir. The fact that this flow entered the central pipe meant that the sardines were not aware of the hourly cyclical changes in flow regime.

Oxygen uptake by the fish (MO₂) was then calculated on the stored files using R software and a custom script. The O₂ saturation (in %) was transformed into O₂ concentration based upon established values of O₂ solubility as a function of temperature and salinity. Temperature was monitored continuously by a probe linked to the O₂ meter, salinity was measured once a day every morning. The slopes of decreasing oxygen concentration over time were estimated through a linear model using an automated R script (see Fig. S2); the first and last minute of the measurements were removed before estimating the slopes. Only slopes with an R² > 0.8 were retained, and measurements collected during fish handling or any intervention on the tanks were removed. The MO₂ was calculated in mg kg⁻¹ h⁻¹, from the decline in water O₂ concentration and considering the total volume of water and the total biomass of the fish (McKenzie et al., 2007; Steffensen, 1989). The hourly measures of MO₂ were averaged to provide a measure of metabolic rate for the entire day. Standard metabolic rates represent metabolic costs of maintenance and were estimated as the 10%-quantile of daily measurements per tank for days in which more than 10 measurements were available. The surface of the tank was open to the atmosphere but surface exchange was so limited between air and water that no corrections were applied (McKenzie et al., 2007). A tank respirometer was run in parallel in the system, but without any sardines, to measure background oxygen consumption by the biofiltered water. This did not represent more than 5 % of fish MO₂, therefore no corrections were applied.

39 **References**

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41 McKenzie, D. J., Höglund, E., Dupont-Prinet, A., Larsen, B. K., Skov, P. V., Pedersen, P. B., & Jokumsen,
42 A. (2012). Effects of stocking density and sustained aerobic exercise on growth, energetics and
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45 McKenzie, D. J., Pedersen, P. B., & Jokumsen, A. (2007). Aspects of respiratory physiology and
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47 condition factor. *Aquaculture*, 263(1–4), 280–294. doi: 10.1016/j.aquaculture.2006.10.022

48 Steffensen, J. F. (1989). Some errors in respirometry of aquatic breathers: How to avoid and correct
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52 **Supplementary tables**

53 *Table S1. ANOVA table for the linear mixed model investigating the effects of the number of fasting days,*
54 *treatment and their interaction on individual body condition with individual ID as a random factor.*

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56 *Table S2. Comparison of candidate GLMMs (binomial) to explain one-week survival of sardines. DF stands for*
57 *degree of freedom. X^2 values and associated p-values are provided for tests between successive models. The*
58 *best model (lowest AIC) is indicated in bold.*

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60 *Table S3. ANOVA table for the segmented regression model investigating changes in specific body mass loss*
61 *across time relative to death.*

62

63 *Table S4. ANOVA table for the segmented regression models investigating changes in specific body mass loss*
64 *according to body condition (based on all data).*

65

66 *Table S5. ANOVA table for the segmented regression models investigating changes in daily respiration rates*
67 *according to body condition.*

68

69 *Table S6. ANOVA table for the segmented regression models investigating changes in daily respiration rates*
70 *according to body condition using transformed data of respiration (monotonous positive BoxCox*

71 *transformation: $Resp_{transf} = \left(\frac{1}{\lambda}\right) * Resp^\lambda$*

72

73 Table S1.

74

| <i>Predictors</i> | <i>Mean Sum Sq</i> | <i>Num DF</i> | <i>F</i> | <i>p</i> |
|------------------------------------------------------|--------------------|---------------|----------|------------------|
| Fasting days | 1.05 | 1 | 2822 | <0.001 |
| Treatment | 0.04 | 2 | 101 | <0.001 |
| Fasting days * Treatment | 0.01 | 2 | 38 | <0.001 |
| Random Effects | | | | |
| σ^2 | 0.0019 | | | |
| $\tau_{00 \text{ ID}}$ | 0.0004 | | | |
| ICC | 0.84 | | | |
| N _{ID} | 53 | | | |
| Observations | 289 | | | |
| Marginal R ² / Conditional R ² | 0.83 / 0.97 | | | |

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77 Table S2.

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| Models | DF | AIC | Deviance | χ^2 | p-value |
|--------------------------------|-----------|--------------|-----------------|----------------------------|-------------------|
| ~ Condition * Treatment | 7 | 267.4 | 253 | 1.10 | 0.578 |
| ~ Condition + Treatment | 5 | 264.5 | 254 | 18.89 | < 0.001 |
| ~ Condition | 3 | 279.4 | 273 | 75.89 | < 0.001 |
| ~ 1 | 2 | 353.2 | 349 | | |

79

80 Table S3.

| <i>Treatment</i> | <i>Predictors</i> | <i>Mean Sum Sq</i> | <i>Num DF</i> | <i>F</i> | <i>p</i> |
|---------------------------------|-----------------------------|--------------------|---------------|----------|-------------------|
| All treatments pooled | Days before death ≥ 10 | 0.028 | 1 | 494 | < 0.001 |
| | 2 < Days before death < 10 | 0.031 | 1 | 563 | < 0.001 |
| | Days before death ≤ 2 | 0.014 | 1 | 243 | < 0.001 |
| | Residuals | 0.110 | 1968 | | |
| | Observations | 1974 | | | |
| | Adjusted R ² | 0.40 | | | |
| Poor initial conditions | Days before death ≥ 10 | 0.013 | 1 | 144 | < 0.001 |
| | 2 < Days before death < 10 | 0.011 | 1 | 124 | < 0.001 |
| | Days before death ≤ 2 | 0.007 | 1 | 75 | < 0.001 |
| | Residuals | 0.000 | 479 | | |
| | Observations | 485 | | | |
| | Adjusted R ² | 0.41 | | | |
| Intermediate initial conditions | Days before death ≥ 16 | 0.013 | 1 | 303 | < 0.001 |
| | 2 < Days before death < 16 | 0.012 | 1 | 264 | < 0.001 |
| | Days before death ≤ 2 | 0.009 | 1 | 195 | < 0.001 |
| | Residuals | 0.000 | 1231 | | |
| | Observations | 1237 | | | |
| | Adjusted R ² | 0.38 | | | |
| Good initial conditions | Days before death ≥ 9 | 0.001 | 1 | 28 | < 0.001 |
| | 2 < Days before death < 9 | 0.003 | 1 | 64 | < 0.001 |
| | Days before death ≤ 2 | 0.001 | 1 | 27 | < 0.001 |
| | Residuals | 0.000 | 246 | | |
| | Observations | 252 | | | |
| | Adjusted R ² | 0.31 | | | |

82 Table S4.

| <i>Treatment</i> | <i>Predictors</i> | <i>Mean Sum Sq</i> | <i>Num DF</i> | <i>F</i> | <i>p</i> |
|---------------------------------|-------------------------|--------------------|---------------|----------|----------------|
| All treatments pooled | Condition < 0.72 | 170.78 | 1 | 221 | < 0.001 |
| | Condition ≥ 0.72 | 129.23 | 1 | 167 | < 0.001 |
| | Residuals | 0.773 | 1970 | | |
| | Observations | 1974 | | | |
| | Adjusted R ² | 0.16 | | | |
| Poor initial conditions | Condition < 0.56 | 99.32 | 1 | 77 | < 0.001 |
| | Condition ≥ 0.56 | 1.75 | 1 | 1 | 0.245 |
| | Residuals | 1.29 | 481 | | |
| | Observations | 485 | | | |
| | Adjusted R ² | 0.13 | | | |
| Intermediate initial conditions | Condition < 0.68 | 109.22 | 1 | 195 | < 0.001 |
| | Condition ≥ 0.68 | 76.08 | 1 | 136 | < 0.001 |
| | Residuals | 0.560 | 1233 | | |
| | Observations | 1237 | | | |
| | Adjusted R ² | 0.21 | | | |
| Good initial conditions | Condition < 0.69 | 7.52 | 1 | 11 | 0.001 |
| | Condition ≥ 0.69 | 1.66 | 1 | 2 | 0.109 |
| | Residuals | 0.68 | 248 | | |
| | Observations | 252 | | | |
| | Adjusted R ² | 0.04 | | | |

| <i>Treatment</i> | <i>Predictors</i> | <i>Mean Sum Sq</i> | <i>Num DF</i> | <i>F</i> | <i>p</i> |
|---------------------------------|-------------------------|--------------------|---------------|----------|----------------|
| All treatments pooled | Condition < 0.64 | 438,801 | 1 | 34 | < 0.001 |
| | Condition ≥ 0.64 | 828,293 | 1 | 65 | < 0.001 |
| | Residuals | 12,772 | 254 | | |
| | Observations | 258 | | | |
| | Adjusted R ² | 0.27 | | | |
| Poor initial conditions | Condition < 0.63 | 480,375 | 1 | 17 | < 0.001 |
| | Condition ≥ 0.63 | 172,786 | 1 | 6 | 0.016 |
| | Residuals | 27,910 | 57 | | |
| | Observations | 61 | | | |
| | Adjusted R ² | 0.25 | | | |
| Intermediate initial conditions | Condition < 0.65 | 339,509 | 1 | 37 | < 0.001 |
| | Condition ≥ 0.65 | 327,268 | 1 | 36 | < 0.001 |
| | Residuals | 9,159 | 146 | | |
| | Observations | 150 | | | |
| | Adjusted R ² | 0.32 | | | |
| Good initial conditions | Condition < 0.78 | 12290 | 1 | 3 | 0.094 |
| | Condition ≥ 0.78 | 22,069 | 1 | 5 | 0.026 |
| | Residuals | 4,192 | 43 | | |
| | Observations | 47 | | | |
| | Adjusted R ² | 0.10 | | | |

86 Table S6.

87

| <i>Treatment</i> | <i>Predictors</i> | <i>Mean Sum Sq</i> | <i>Num DF</i> | <i>F</i> | <i>p</i> |
|---------------------------------|-------------------------|----------------------|---------------|----------|----------------|
| All treatments pooled | Condition < 0.64 | 0.0003 | 1 | 19 | < 0.001 |
| | Condition ≥ 0.64 | 0.0008 | 1 | 42 | < 0.001 |
| | Residuals | 0.0000 | 254 | | |
| | Observations | 258 | | | |
| | Adjusted R ² | 0.18 | | | |
| | λ | -0.71 | | | |
| Poor initial conditions | Condition < 0.64 | 3.4 10 ⁻⁷ | 1 | 16 | < 0.001 |
| | Condition ≥ 0.64 | 0.5 10 ⁻⁷ | 1 | 2 | 0.119 |
| | Residuals | 0.1 10 ⁻⁷ | 57 | | |
| | Observations | 61 | | | |
| | Adjusted R ² | 0.21 | | | |
| | λ | -1.31 | | | |
| Intermediate initial conditions | Condition < 0.65 | 0.021 | 1 | 28 | < 0.001 |
| | Condition ≥ 0.65 | 0.020 | 1 | 26 | < 0.001 |
| | Residuals | 0.001 | 146 | | |
| | Observations | 150 | | | |
| | Adjusted R ² | 0.25 | | | |
| | λ | -0.38 | | | |
| Good initial conditions | Condition < 0.79 | 0.8 10 ⁻⁶ | 1 | 2 | 0.126 |
| | Condition ≥ 0.79 | 2.3 10 ⁻⁶ | 1 | 7 | 0.013 |
| | Residuals | 0.1 10 ⁻⁶ | 43 | | |
| | Observations | 47 | | | |
| | Adjusted R ² | 0.12 | | | |
| | λ | -0.99 | | | |

88

89 Supplementary figures

90 Fig S1: Body condition at the start of the fasting experiment according to the feeding treatment experienced
91 before. LP and SP stand for large and small particles respectively, while LQ and SQ stand for large and small
92 quantities respectively.

93

94 Figure S2: Dissolved oxygen in tank 2 during two days (2017-07-07 and 2017-07-08) as an example of
95 respiration rate estimation. Cycles, during which oxygen consumption are calculated, are indicated in colour
96 depending on the r -square of the linear regression. On the first day, a period was removed as fish were handled
97 during that time for biometry, tanks cleaned, etc.

98

99 Figure S3: Q-Q plot of linear mixed model residuals of the body condition index over time.

100

101 Figure S4: Slopes of individual body condition loss (d^{-1}) through fasting according to initial feeding condition.

102

103 Figure S5: Number of daily sardine deaths (A) and cumulative mortality (in %) of sardines (B) along the fasting
104 experiment. Days where sardines were handled are shown in black bars, while days with no handling appear as
105 white bars.

106

107 Figure S6: Cumulative mortality of sardines (in %) originating from each of the three initial feeding conditions
108 (as indicated by colours) according to body condition.

109

110 Figure S7: Mean \pm SE specific body mass loss $\left(\frac{dm}{mdt}\right)$ per day along time according to each initial feeding
111 treatment. Colours indicate the initial feeding treatment sardines originated from. As individuals died at
112 different time in the experiment, the number of days has been estimated relative to death. The vertical dashed
113 line shows a rupture in the slope of all three treatments.

114

115 Figure S8: Specific body mass loss $\left(\frac{dm}{mdt}\right)$ expressed as % according to body condition. Colour indicates the
116 treatment sardines originated from. The segmented regressions are indicated by the black line and the 95%
117 confidence intervals with dashed lines. The breakpoint along with its 95% CI is also indicated at the bottom of
118 the figure.

119

120 Figure S9: Q-Q plots of residuals of models explaining the specific body mass loss by body condition through
121 fasting considering all data (left), only specific body mass loss lower than 4% (middle) and only specific body
122 mass loss lower than 2% (right).

123

124 *Figure S10: Distribution of specific body mass loss $\left(\frac{dm}{mdt}\right)$ through fasting*

125

126 *Figure S11: Q-Q plots of residuals of models explaining the metabolic rates by body condition through fasting*
127 *considering either raw (left side) or transformed data (right side) from all sardines, sardines from poor initial*
128 *conditions, intermediate initial conditions or good initial conditions.*

129

130 *Figure S12: Mean \pm SE body condition of sardines sampled in the wild before (in blue) or after (in red) 2008 for*
131 *each month of the year.*

132

133 *Figure S13: Body condition of sardines sampled in the wild before or after 2008 depending on maturity stages. n*
134 *indicates the sample size in each category. Boxes sharing common letters are not significantly different from*
135 *each other according to Bonferroni-corrected Wilcoxon tests. Maturity stage 1 corresponds to sexual rest,*
136 *stages 2 to 4 to increasing development of the gonads, 5 to active spawning and 6 to post spawning.*

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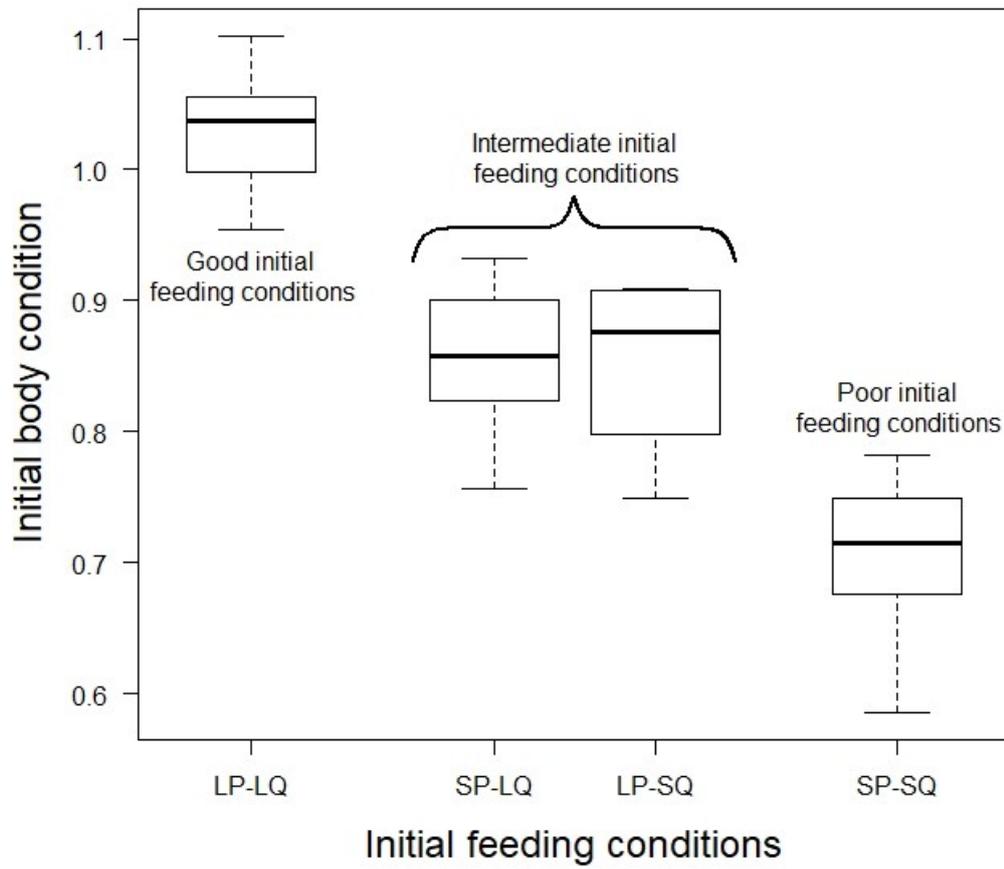
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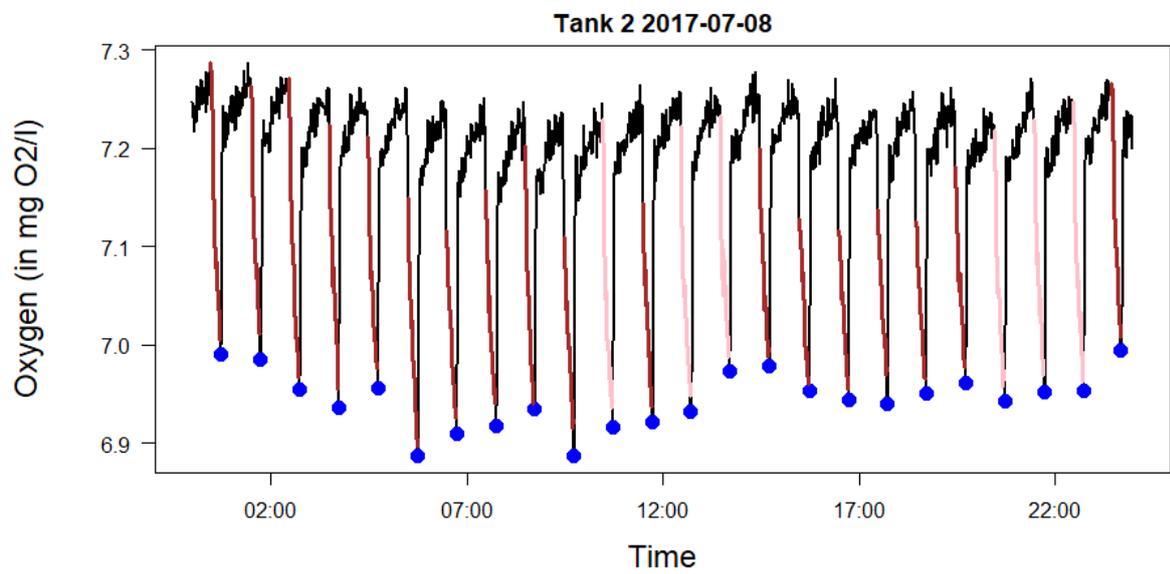
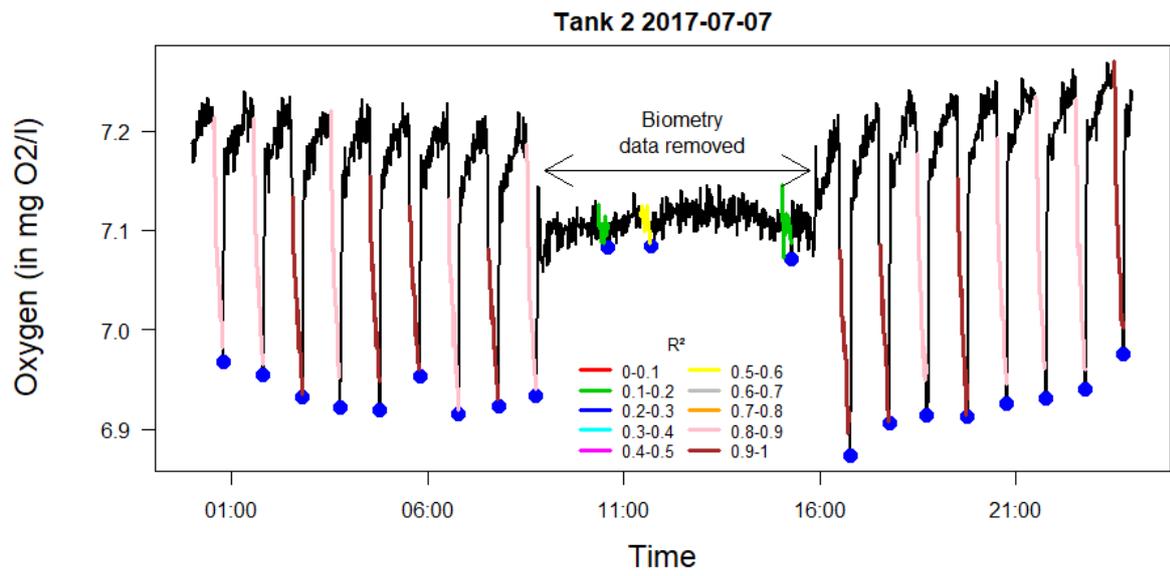


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161 Fig. S1

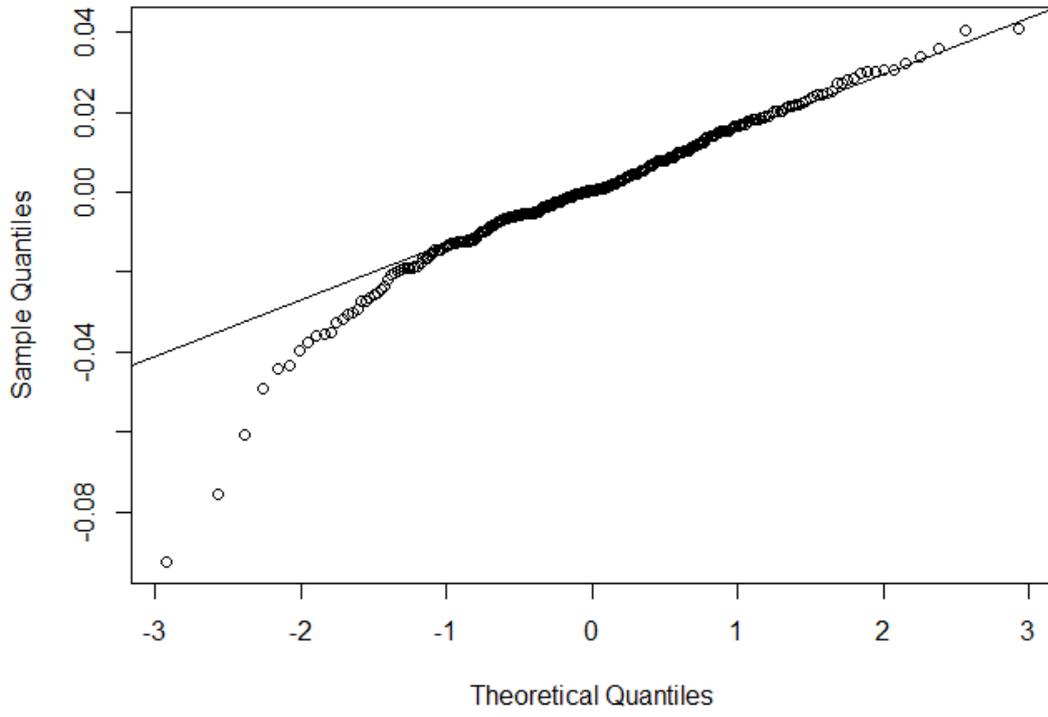
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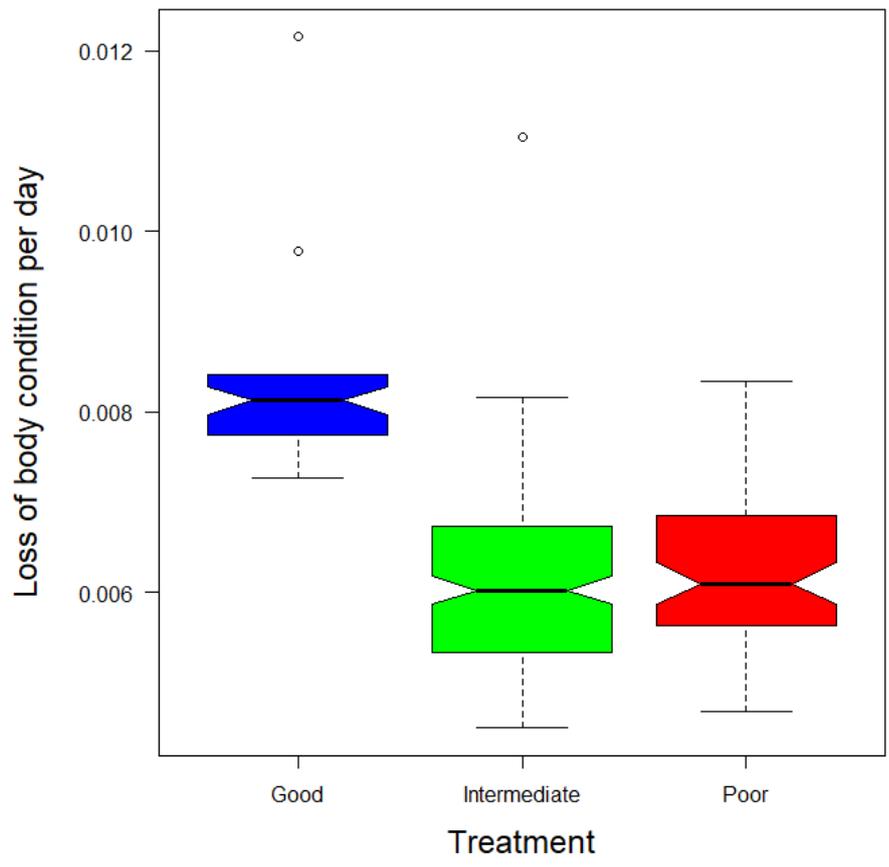
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165 Fig. S2

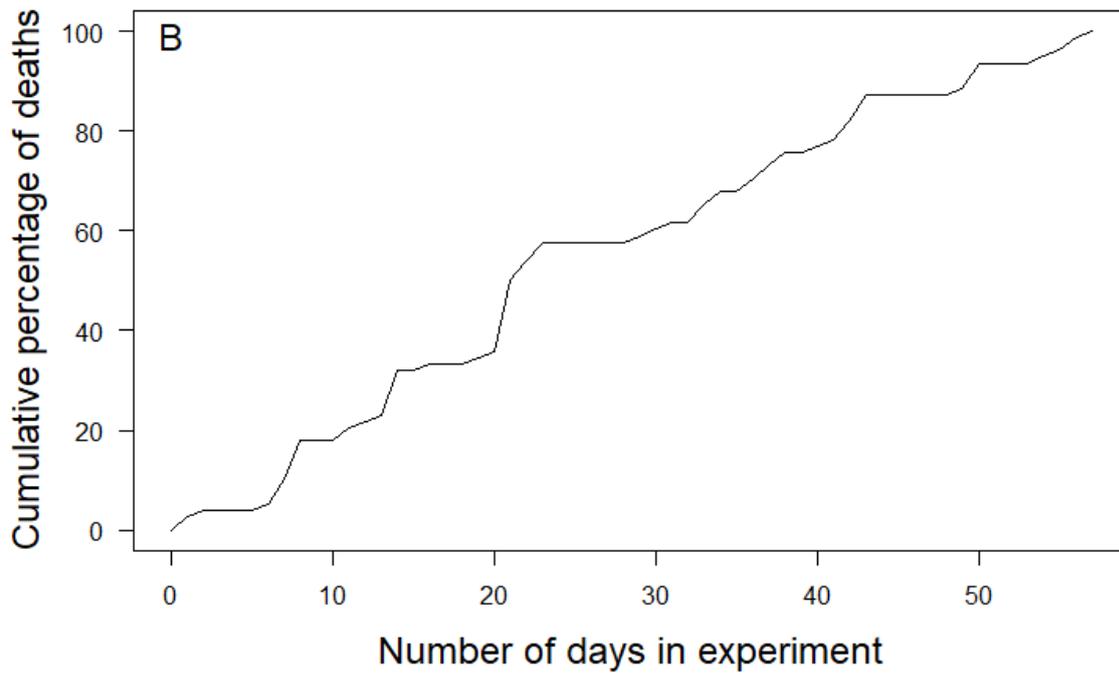
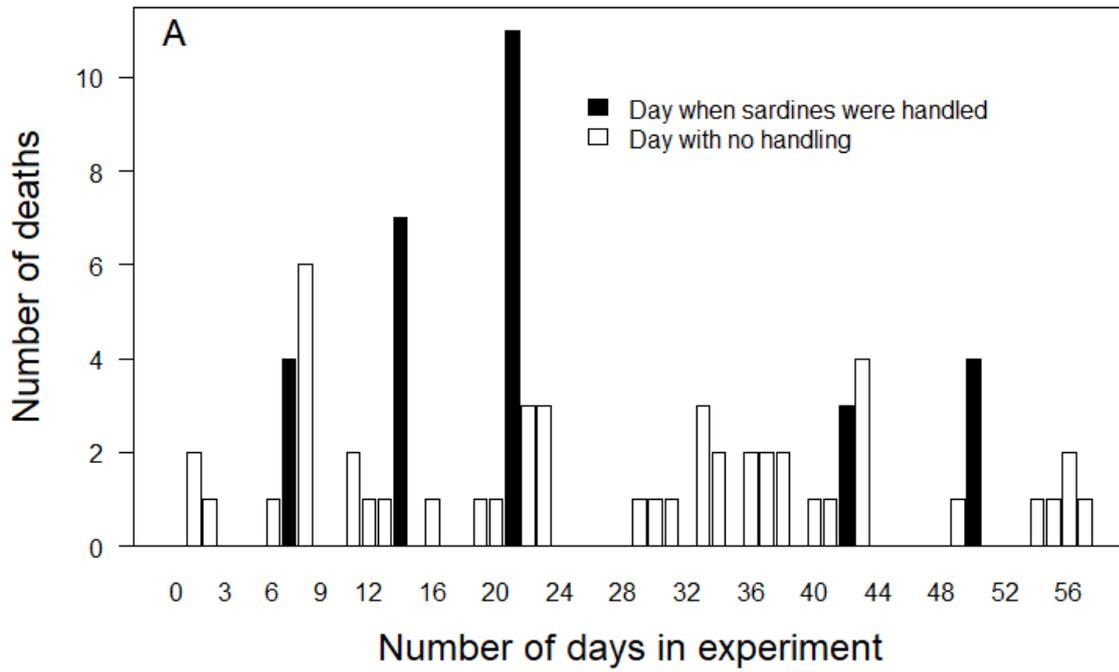


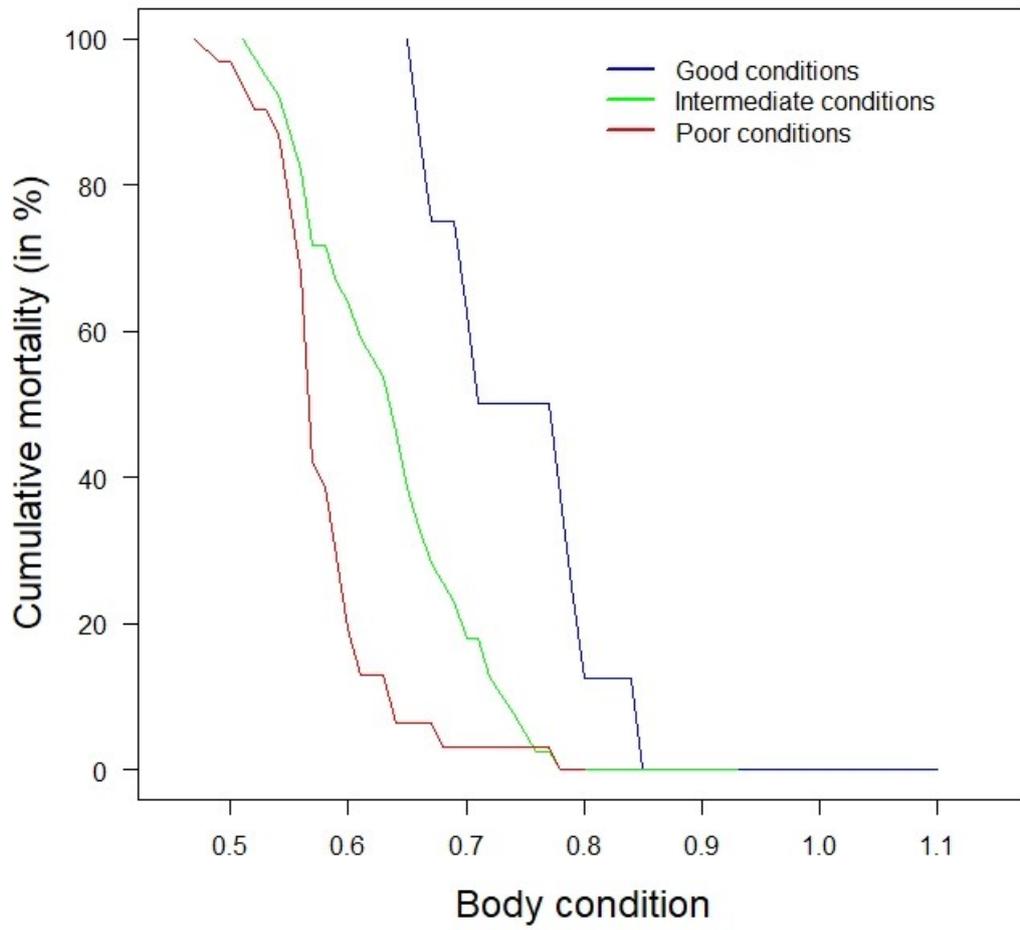
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167 Fig. S3



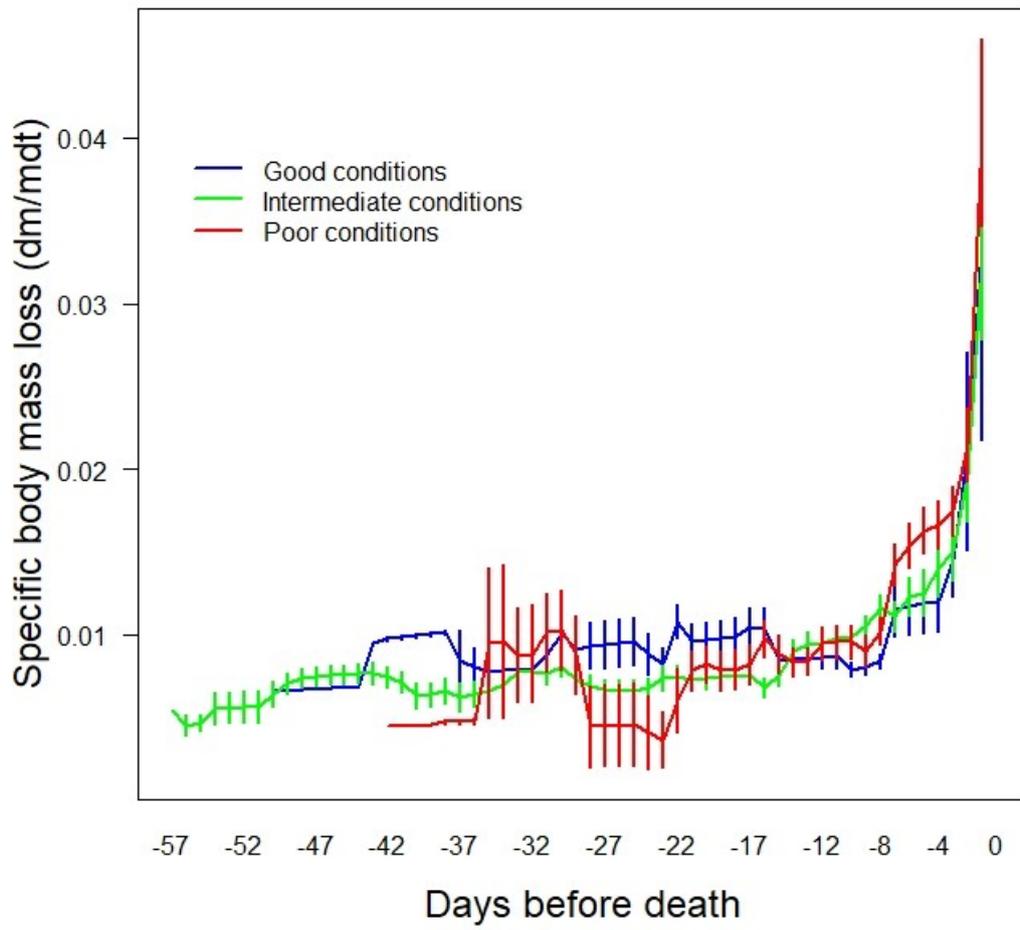
169 Fig. S4





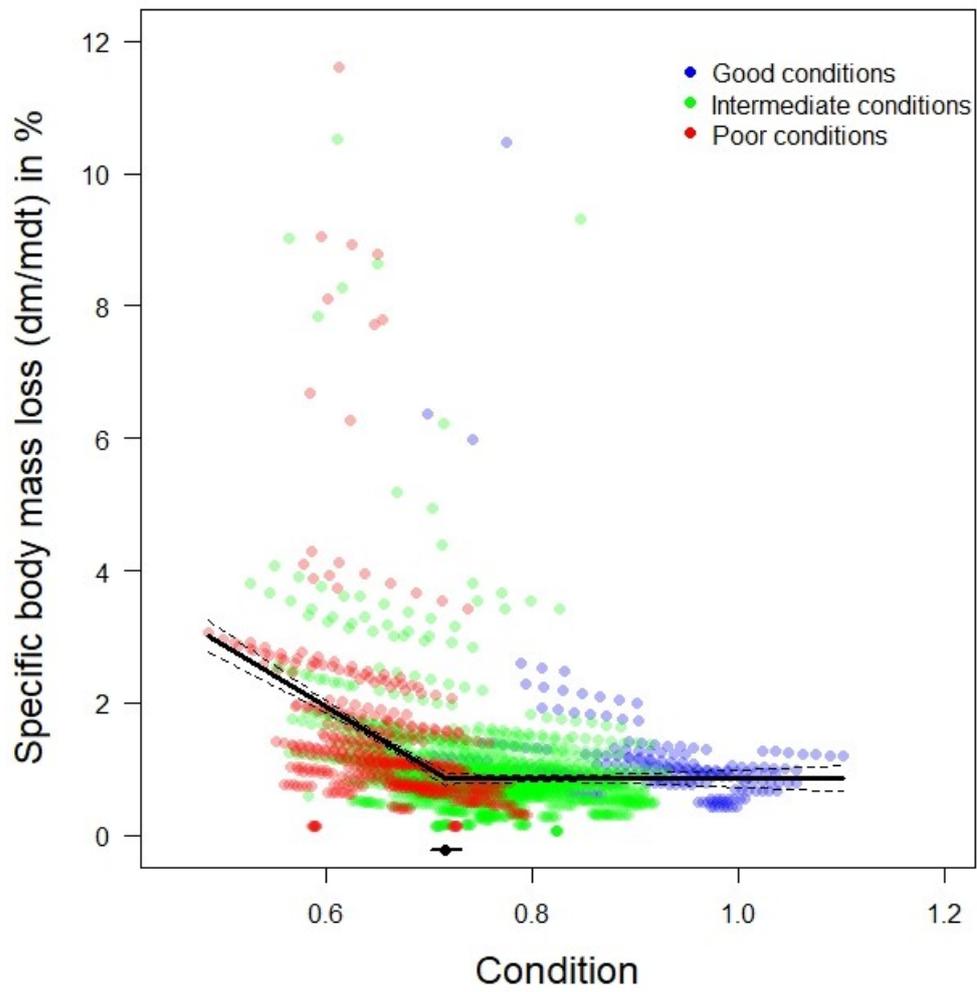
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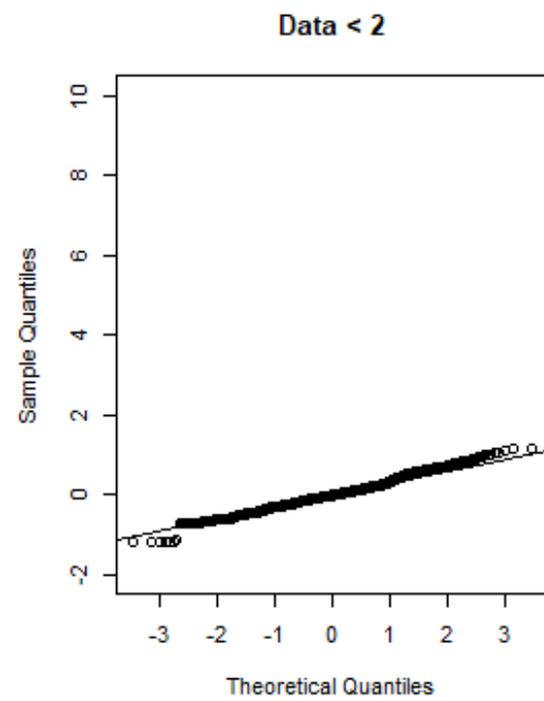
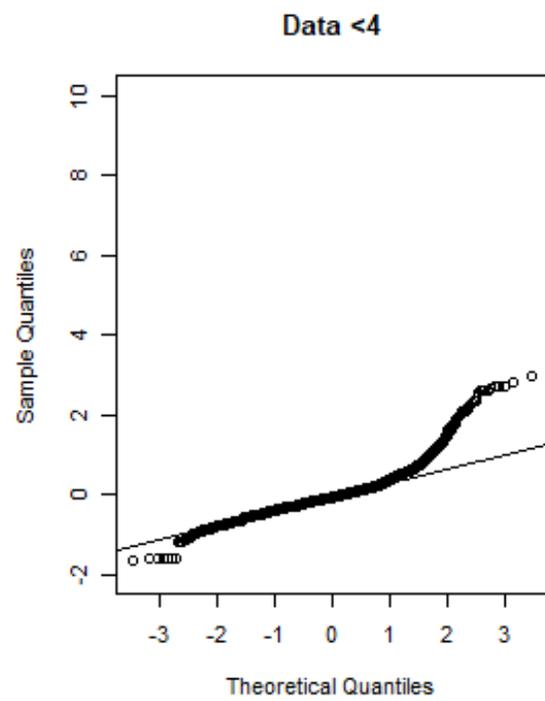
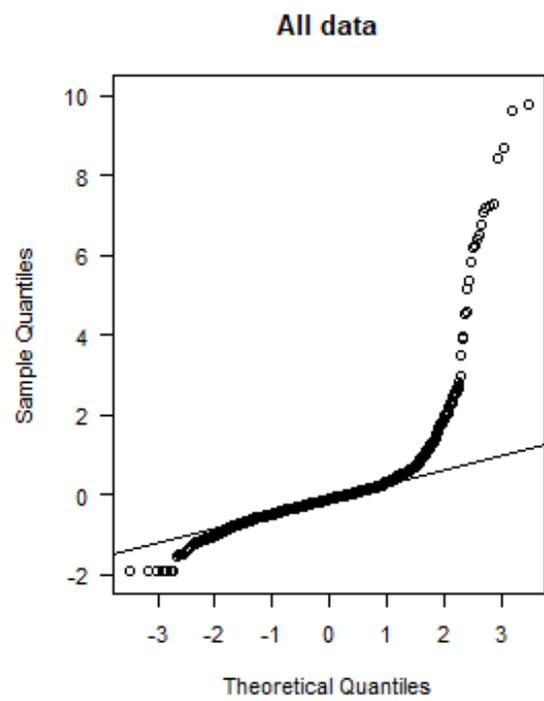
176 Fig. S6



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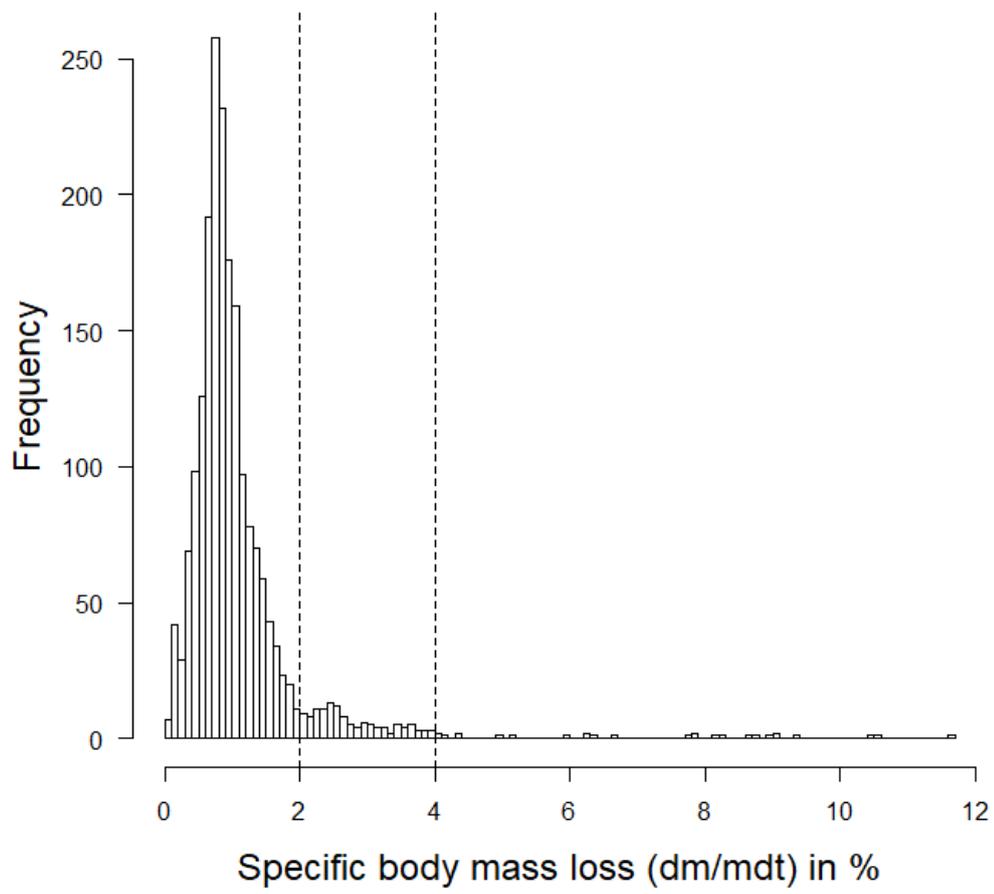
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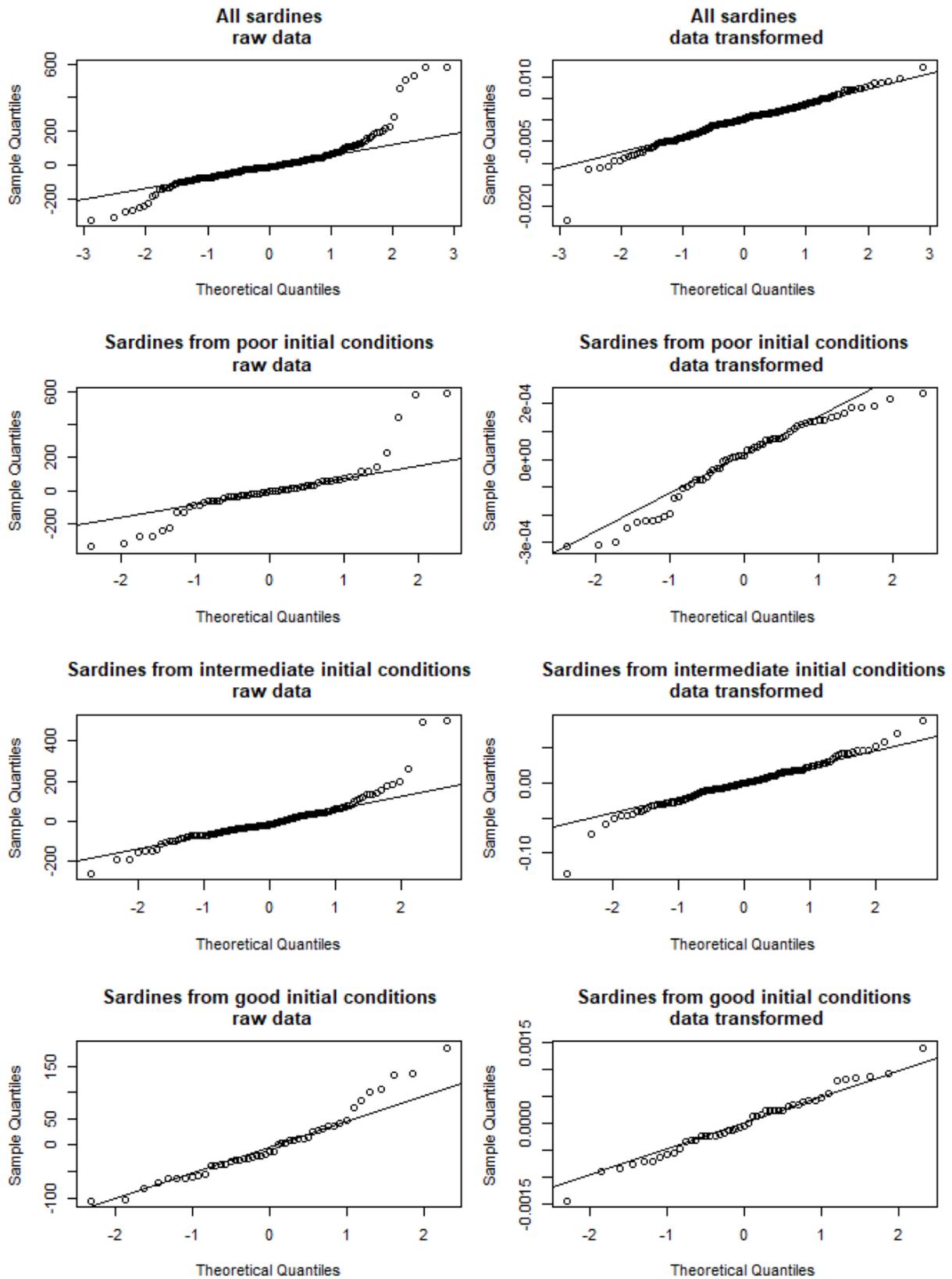


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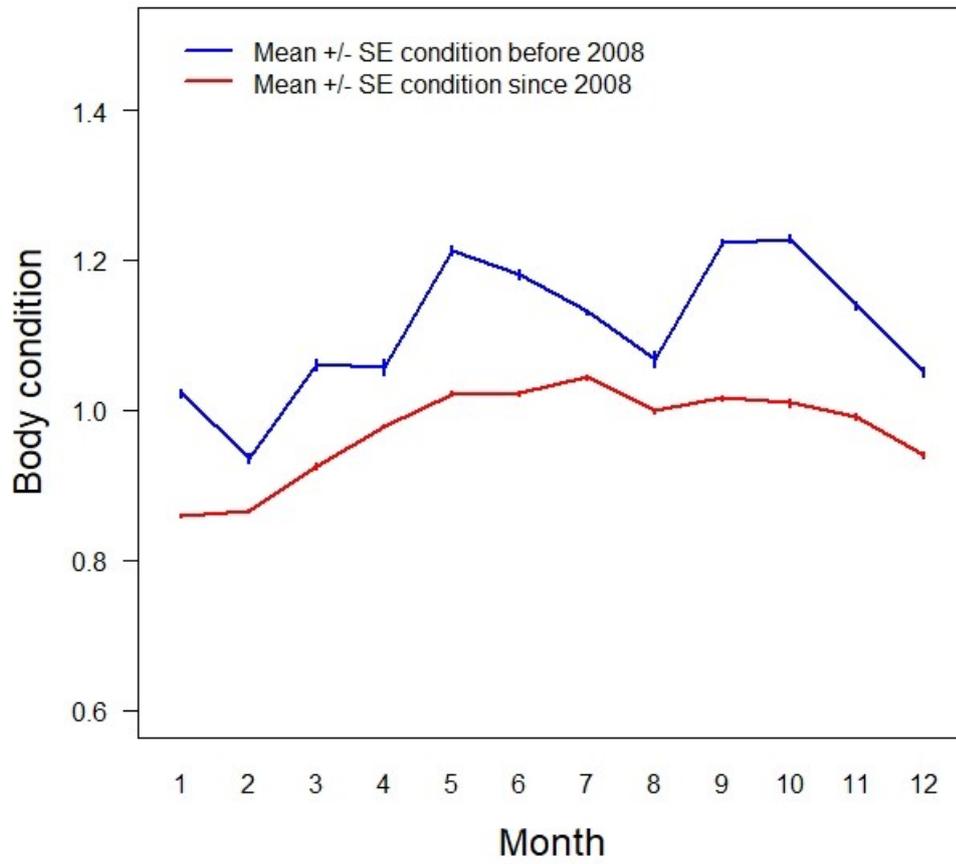


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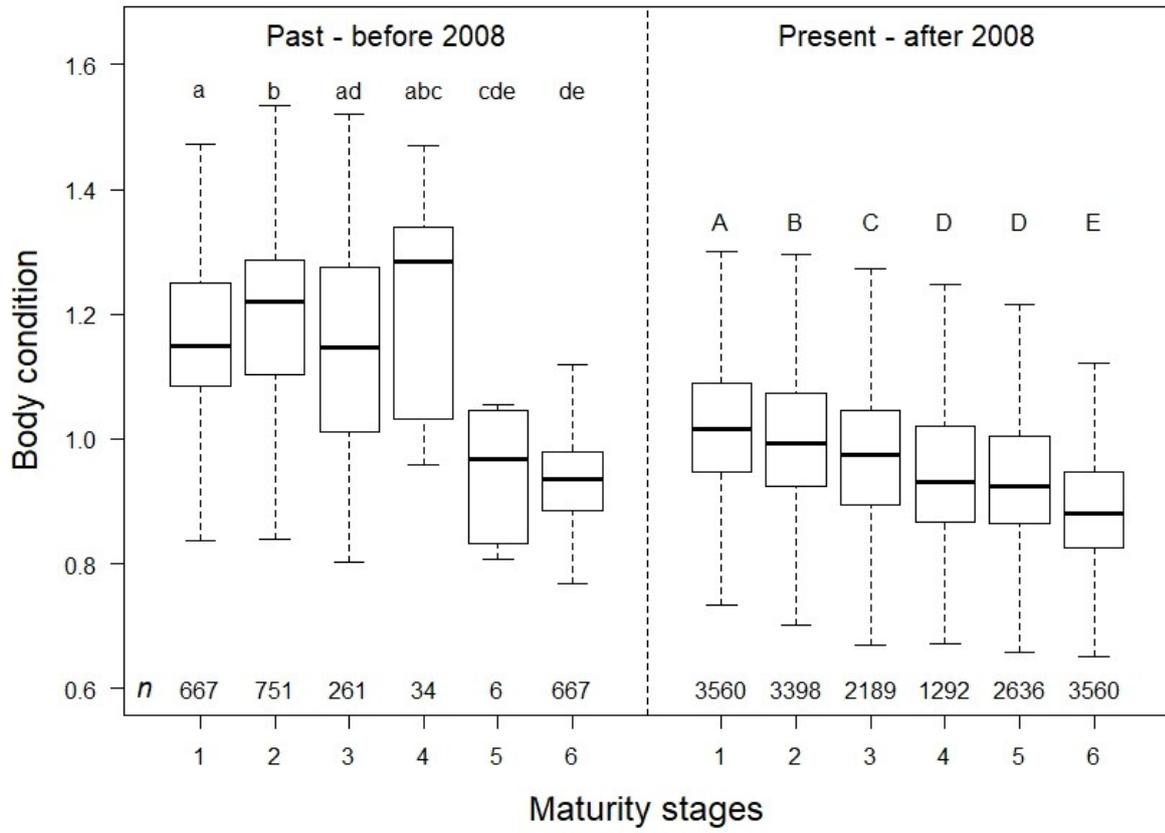
186 Fig. S11

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189 Fig. S12



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191 Fig. S13