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Maxi Castrillejo, Peter J. Statham, Gary R. Fones, H el ene Planquette, Farah Idrus, et al.. Dissolved trace metals (Ni, Zn, Co, Cd, Pb, Al, and Mn) around the Crozet Islands, Southern Ocean. *Journal of Geophysical Research. Oceans*, Wiley-Blackwell, 2013, 118, pp.5188-5201. 10.1002/JGRC.20359 . hal-00939263

HAL Id: hal-00939263

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Dissolved trace metals (Ni, Zn, Co, Cd, Pb, Al, and Mn) around the Crozet Islands, Southern Ocean

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Received 13 December 2012; revised 23 July 2013; accepted 9 August 2013; published 9 October 2013.

[1] A phytoplankton bloom shown to be naturally iron (Fe) induced occurs north of the Crozet Islands (Southern Ocean) every year, providing an ideal opportunity to study dissolved trace metal distributions within an island system located in a high nutrient low chlorophyll (HNLC) region. We present water column profiles of dissolved nickel (Ni), zinc (Zn), cobalt (Co), cadmium (Cd), lead (Pb), aluminium (Al), and manganese (Mn) obtained as part of the NERC CROZEX program during austral summer (2004–2005). Two stations (M3 and M1) were sampled downstream (north) of Crozet in the bloom area and near the islands, along with a control station (M2) in the HNLC zone upstream (south) of the islands. The general range found was for Ni, 4.64–6.31 nM; Zn, 1.59–7.75 nM; Co, 24–49 pM; Cd, 135–673 pM; Pb, 6–22 pM; Al, 0.13–2.15 nM; and Mn, 0.07–0.64 nM. Vertical profiles indicate little island influence to the south with values in the range of other trace metal deprived regions of the Southern Ocean. Significant removal of Ni and Cd was observed in the bloom and Zn was moderately correlated with reactive silicate (Si) indicating diatom control over the internal cycling of this metal. Higher concentrations of Zn and Cd were observed near the islands. Pb, Al, and Mn distributions also suggest small but significant atmospheric dust supply particularly in the northern region.

Citation: Castrillejo, M., P. J. Statham, G. R. Fones, H. Planquette, F. Idrus, and K. Roberts (2013), Dissolved trace metals (Ni, Zn, Co, Cd, Pb, Al, and Mn) around the Crozet Islands, Southern Ocean, *J. Geophys. Res. Oceans*, 118, 5188–5201, doi:10.1002/jgrc.20359.

1. Introduction

[2] Artificial Fe enrichment experiments (summarized by *Boyd et al.* [2007] and *de Baar et al.* [2005]) confirmed Fe as the prime limiting micronutrient in HNLC regions. A range of other bioactive trace metals are also vital for biological productivity as they are often involved in enzymatic activity or become part of proteins [*Morel and Price*, 2003]. Some trace metals may be colimiting [*Morel et al.*, 1994] when at low concentrations or toxic at elevated concentrations [*Mann et al.*, 2002] and the bioavailability of trace elements such as Zn, Co, and Cd may be influenced

by complexing ligands [e.g., Zn: *Bruland*, 1989; *Ellwood and van den Berg*, 2000; Co: *Ellwood and van den Berg*, 2001; *Saito et al.*, 2004; Cd: *Bruland*, 1992].

[3] The Southern Ocean is the largest HNLC region and plays a key role in climate regulation via the ocean carbon pump [*Anderson et al.*, 2009]. It is typically characterized by biolimiting concentrations of Fe, deep mixed layers in winter, with moderate seasonal warming and stratification. Therefore, this region can be very sensitive to climate change due to an increased stratification [*Sarmiento et al.*, 1998] which may affect the biological pump and the water mass formation and circulation [*Marinov et al.*, 2006; *De Vries and Primeau*, 2011]. However, studies of trace metals other than Fe are limited and only few of them cover the deep waters of the Southern Ocean [*Aparicio-Gonzalez et al.*, 2012]. Data for the Indian sector of the Southern Ocean including the metals investigated here are particularly scarce. *van Beusekom et al.* [1997] provided the first water column and sedimentary distribution of Al and silicic acid along the Enderby and Crozet Basins. *Bucciarelli et al.* [2001] found coastal and offshore waters enriched in dissolved Mn and Fe with sedimentary and lithogenic origin in the wake of Kerguelen Islands, whereas *Bown et al.* [2012] suggested that lateral advection and dissolution of Co from basaltic sediments explain the high dissolved Co concentrations above the Kerguelen Plateau.

[4] The interdisciplinary CROZet natural iron bloom and EXPORT experiment (CROZEX) [*Pollard et al.*, 2007b] confirmed that Fe released from the Crozet Islands and Plateau

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2169-9275/13/10.1002/jgrc.20359

[Planquette *et al.*, 2007] supports an annual bloom and leads to an increase of the C export [Pollard *et al.*, 2009]. Large-scale circulation constrains the bloom to the north of the Islands [Pollard *et al.*, 2007a] providing an ideal location to compare the trace metal behavior of the naturally induced bloom with HNLC conditions south of the archipelago. This is the first reporting of trace metal concentrations in this area to date with the exception of dissolved Fe [Planquette *et al.*, 2007] and particulate Fe and Al [Planquette *et al.*, 2009].

[5] This study provides vertical distributions of dissolved Ni, Zn, Co, Cd, Pb, Al, and Mn, constituting a substantial addition to the trace metal biogeochemistry knowledge around Crozet Islands and more generally, in a Southern Ocean naturally induced bloom. Results are discussed in terms of sources, sinks, and biogeochemical cycles in order to test the hypotheses that (1) trace element concentrations south of the archipelago are representative of other HNLC zones within the Southern Ocean and (2) bioactive trace metals show biological uptake in the bloom area.

2. Methods

2.1. Study Area, the Phytoplankton Bloom, and Sampling Sites

[6] Crozet (46°S, 52°E) is located in the western Indian Sector of the Southern Ocean, 2500 km southeast of South Africa. It comprises a shallow plateau, two main islands, “Île de la Possession” and “Île de l’Est,” as well as three smaller ones 100 km further west (Figure 1).

[7] Lying within the Polar Frontal Zone (PFZ) [Pollard *et al.*, 2002], Crozet is bounded west and north by an “S”-shaped Sub-Antarctic Front (SAF), the northern branch of the Antarctic Circumpolar Current (ACC) and south by the Polar Front (PF) [Pollard *et al.*, 2007a], a permanent feature mainly controlled by the topography of Del Caño Rise and Crozet Plateau [Pollard and Read, 2001]. The accumulation of Fe and development of the bloom to the north is facilitated by the long residence time (~62 days) of water in the SAF semienclosed area and the weak northward surface flow passing over the Crozet Plateau [Pollard *et al.*, 2007a]. Therefore, south of Crozet can be referred to as “upstream,” while north of Crozet is “downstream” of the islands. A complete description of the circulation around Crozet is given by Pollard *et al.* [2007a].

[8] The bloom starts north of the plateau then moves south toward the islands. The beginning and initial location of the bloom are controlled by the gradual increase of light during austral spring and summer, whereas its development is dominated by physical conditions and micro and macro-nutrient availability [Venables and Moore, 2010; Venables *et al.*, 2007]. Venables *et al.* [2007] showed using merged Sea WIFS/MODIS Chl-*a* images that the main bloom began north of the plateau in early-mid September 2004 and followed an exponential-like growth up to mean chlorophyll concentrations of $\sim 4 \mu\text{g Chl-}a \text{ L}^{-1}$ before collapsing by early November 2004.

[9] The samples analyzed here were taken during the first cruise (D285) of the CROZEX project on board of *RRS Discovery*, from 3 November to 10 December 2004, right after the main bloom declined [Venables *et al.*, 2007].

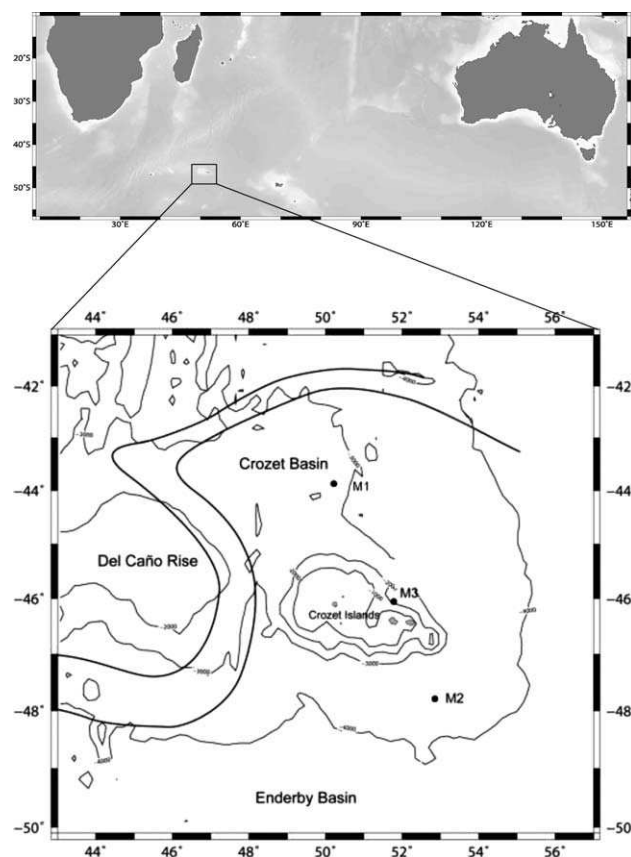


Figure 1. Bathymetry of the survey area between 40 and 50°S and 43 and 57°E. Black filled circles mark sites M1, M2, and M3 sampled during leg 1 (D285) of CROZEX experiment (courtesy of Raymond Pollard, NOCS). The sub-Antarctic Front (SAF) is indicated by the thin black lines. Detailed information about sampling sites and circulation can be found in Pollard *et al.* [2007a, 2007b].

The three stations included in this study were located at sites M1 (bloom area), M3 (island vicinity), and M2 (HNLC area), respectively (Figure 1). Detailed information about sampling sites is given by Pollard *et al.* [2007b].

2.2. Sampling and Cleaning Procedures

2.2.1. Inorganic Nutrients

[10] In summary, samples were collected using a trace-metal clean Titanium conductivity-temperature-depth (CTD) rosette, directly transferred from Niskin bottles to polystyrene coulter counter vials and stored at 4°C for 12 h prior to analysis. A Skalar San^{plus} autoanalyzer was used to determine nitrate + nitrite (hereinafter nitrate), phosphate, and reactive silica (hereinafter Si) following Sanders and Jickells [2000]. Details on analysis and data quality for the inorganic nutrients can be found in Sanders *et al.* [2007]. Since no nutrient data for station M2 were available, we use data from nearest stations (<80 km, stations 500 and 506, Sanders *et al.* [2007]) that were sampled 6 h before and 17 h after M2 using a stainless CTD rosette.

2.2.2. Trace Metals

[11] Prior to the cruise, low-density polyethylene (LDPE) storage bottles (500 and 1000 mL) were soaked for 3 days each in $\sim 2\%$ (v/v) aqueous Decon detergent and,

50% (v/v) hydrochloric (HCl), and 50% (v/v) nitric (HNO₃) acids (Fisher Scientific, Trace Metal Grade). Bottles were rinsed with Milli-Q water (>18.2 MΩ cm) between different steps and after the last acid bath. The bottles were filled with Milli-Q water acidified to a pH of ~1.9 with quartz subboiled distilled HCl (Q-HCl) and stored in clean zipper seal polyethylene bags in a class-100 clean-air laboratory at room temperature. Polycarbonate filters were soaked in 10% (v/v) Q-HCl for 14 days, rinsed and stored in Milli-Q water. Seawater was collected on a trace-metal clean Titanium rosette equipped with 10 L Ocean Technical Equipment (OTE) bottles modified for trace metal sampling. Samples were processed on board in a container equipped with a class 100 laminar flow bench. OTE bottles were pressurized with filtered high-purity nitrogen (BOC Gases) before valves were opened. Seawater was directed through an acid-washed 0.2 μm polycarbonate filter (Whatman) housed in an acid-washed Teflon filtration unit. Samples were stored in 1 L and 500 mL clean low-density polyethylene bottles (Nalgene). Bottles were rinsed twice with the seawater sample prior to storage. After collection, samples were acidified with Q-HCl to pH ~1.7 and stored prior to analysis. A full description is given by *Planquette et al.* [2007].

2.3. Trace Metal Analyses

2.3.1. Dissolved Nickel, Zinc, Cobalt, Cadmium, and Lead

[12] The method involves an off-line removal of the salt matrix and preconcentration of metals (Ni, Zn, Co, Cd, and Pb) using the commercial resin Toyopearl AF-Chelate-650M (Tosoh Bioscience) prior to their determination by ICP-MS. This resin has been recently used for multielemental extraction of trace metals in seawater [*Milne et al.*, 2010].

[13] LDPE bottles (Nalgene) and 125 mL Teflon bottles (Fluorinated Ethylene Propylene (FEP), Nalgene) were used for reagent storage and UV radiation of samples, respectively. All containers were cleaned following the cleaning steps described before and stored in double zipper seal polyethylene bags when not being used. Sample manipulation was done following clean trace metal handling techniques in a laminar flow hood.

[14] High-purity nitric, hydrochloric, and acetic acids (ROMIL SpA grade), and Milli-Q water (>18.2 MΩ cm) were used for the preparation of all reagents, including HNO₃ (1 M) eluent and HCl (2 M) column wash solutions. Saturated ammonium acetate solution (19.2 M) was obtained by adding Milli-Q water to solid ammonium acetate that had been formed by bubbling high-purity ammonia gas through cooled concentrated acetic acid and the saturated solution was then diluted with Milli-Q water to give a 2 M buffer solution. An additional ammonium acetate buffer wash solution (20 mM) was made by further dilution of the 2 M reagent. The ammonium acetate (2 M) was adjusted to the appropriate pH using high-purity ammonia solution such that on addition of the buffer solution to the seawater samples (initial pH 1.7) and acidified North Atlantic bulk seawater that was used as an internal reference material, a final pH of ~5.9 was obtained.

[15] An enclosed flow through system was built to reduce external contamination, in which six parallel micro-

columns (laboratory made from polymethyl methacrylate with an internal volume of ~80 μL) were filled with the chelating resin. Sample, wash, and eluting solutions were pushed through the columns with a six channel peristaltic pump (Gilson Mini-Puls) at a speed of 1.6 mL min⁻¹. Pump tubing was made of PVC and connecting tubing was made of FEP (0.8 mm i.d.).

[16] It has been reported that a significant fraction of bioactive trace metals such as Zn, Co, and Cu form chelates with strong metal-binding organic ligands in seawater [*Bruland*, 1989; *Coale and Bruland*, 1988; *Ellwood and van den Berg*, 2001]. Recent studies suggest that ~40% of Co [*Milne et al.*, 2010; *Shelley et al.*, 2010] and ~10% of Cu [*Milne et al.*, 2010] remain strongly bound to organic ligands even after long periods of acidification, and these complexes may make the metals unavailable to chelating resins. To convert the complexed metals to resin available forms, samples were UV irradiated in a purpose built system containing four low power UV lamps (20W each, Ultra-Violet Products Ltd) placed either side of the water samples at a distance of ~2 cm from the FEP bottles. Initial experiments demonstrated that irradiation for 3 h was required to make Co fully available to the Toyopearl resin, subsequently all Crozet samples were then irradiated for this time prior to the metal preconcentration and salt matrix removal using the column system.

[17] After UV irradiation, ammonium acetate solution (20 mM) with a pH ~5.9 was circulated for 1 min through the sample lines and columns to remove any residual acid from the previous elution step. Aliquots (~100 mL) of the acidified samples (pH ~1.7) were buffered to pH ~5.9 and pumped through the column system. A 1 min rinse with ammonium acetate solution (20 mM) was then used to remove the residual salt in the columns. The exact mass of seawater passed through each column was determined gravimetrically. Metals were eluted using 2 mL of HNO₃ (1 M) into 4 mL acid cleaned auto-sampler polypropylene vials. Columns were cleaned for 1 min with HNO₃ (1 M) after sample collection to remove any residual analyte. Before and after every analytical session with the system, columns, tubing, fittings, and connectors of the manifold were cleaned using HCl (2 M) and Milli-Q water for 10 and 5 min, respectively.

[18] Measurements of metals in concentrates were performed using an Agilent 7500ce Inductively Coupled Plasma-Mass Spectrometer (ICP-MS) that uses an octopole reaction system. The reaction system was run in collision mode using helium gas for the analysis of Co⁵⁹, Ni⁶⁰, Zn⁶⁶, Cd¹¹¹, and Pb²⁰⁸. The use of the octopole reaction system eliminates any polyatomic spectral interference and, therefore, the need for interference corrections. For Pb, the isotopes of Pb were summed using the Agilent ChemStation software. This approach helps minimize any errors caused by variations in the natural isotopic pattern which is often observed. For this work, mixed standards were prepared in the range of 0.001–50 μg L⁻¹ in a 2% HNO₃/0.5% HCl (v/v) solution. For Co⁵⁹, Cd¹¹¹, and Pb²⁰⁸, the calibration range was from 0.001 to 5 μg L⁻¹, while for Ni⁶⁰ and Zn⁶⁶, it was from 0.01 to 50 μg L⁻¹. The calibration standards were run at the start of the day and quality control standards of 0.1 and 1 μg L⁻¹ were run every 20 samples to check for drift, these were then incorporated into the set of

Table 1. Blanks ($n = 5$), Limits of Detection (3σ), and Column Recoveries for Ni, Zn, Co, Cd, and Pb

	Blank	Limit of Detection	Recovery (%)
Ni (nM)	0.01–0.02	0.02–0.04	93–99
Zn (nM)	0.08–0.19	0.12–0.43	89–93
Co (pM)	1–3	2–8	91–97
Cd (pM)	5–6	2–7	74–77
Pb (pM)	2–4	2–8	92–98

standards to produce new calibration curves at the end of the run. An internal standard of $20 \mu\text{g L}^{-1}$ Rh¹⁰³ and Bi²⁰⁹ (stock solution of $400 \mu\text{g L}^{-1}$ Rh¹⁰³ and Bi²⁰⁹ with 20-fold online dilution) was used to determine any variation in the intensity of their signal; Rh¹⁰³ was used to correct the Co⁵⁹, Ni⁶⁰, Zn⁶⁶, and Cd¹¹¹ signal while Bi²⁰⁹ was used for Pb²⁰⁸. The data processing procedure including linear drift correction, blank subtraction, calibrations, and Pb²⁰⁸ correction was all undertaken using the Agilent software ChemStation.

[19] Information relevant to blanks, limit of detection, and metal recovery of the preconcentration and salt matrix removal step is presented in Table 1. The blank was calculated using Milli-Q water which was processed in the same way as samples at the beginning of every analytical day and between each batch of samples. Blank range ($n = 5$) spanned from 0.01 to 0.02 nM; 0.08 to 0.19 nM; 1 to 3 pM; 5 to 6 pM; and 2 to 4 pM for Ni, Zn, Co, Cd, and Pb, respectively. Blanks were generally 1–2 orders of magnitude lower than measured concentrations in the samples, and limits of detection calculated as three times the standard deviation of blank values (3σ) were well below the concentrations found in the samples. A mixed standard (2.5 nM for Ni and Zn; 0.1 nM for Co, Cd, and Pb) was used to check the recovery of metals in each column, typically one standard every six Crozet samples. Recovery was always above 90% except for Cd that showed a recovery of 74%. Thus, hereinafter all presented Cd data are corrected for the recovery. Low recoveries of Cd when hydrochloric acid is used for acidification during storage have been reported elsewhere and the use of nitric acid seems to be more effective at removing organic interferences (P. Croot, personal communication, 2013). Recoveries between columns only varied 3–6% and data were very reproducible. Duplicates were made every three Crozet samples and North Atlantic bulk seawater was used as internal standard to address any bias of the analytical method. Accuracy was determined using Certified International Standard Materials NASS-6

and CASS-5 (National Research Council of Canada) that were processed in the same way as the samples (Table 2) but without UV irradiation to ensure consistency with analyses done in generating the certified values.

2.3.2. Dissolved Aluminium

[20] Dissolved Al was determined using the lumogallion fluorimetric method described by *Hydes and Liss* [1976], that has been used extensively for studies of dissolved Al in the ocean without [*van Beusekom et al.*, 1997] or with a preconcentration step included [*Resing and Measures*, 1994]. High-purity reagents were used throughout including the ammonium acetate buffer (to provide a pH of 5) and ammonia solution, and the fluorescent Al compound was allowed to form overnight at room temperature in FEP bottles. The fluorescence signal was measured using a Perkin Elmer LS55, with excitation and emission wavelengths set at 480 and 590 nm, respectively. Procedural blank was 0.758 nM with a precision of 0.048 nM. The detection limit (calculated as three times the standard deviation of the blank) was 0.143 nM, and averaged relative standard deviation (based on measurements of one sample) was 2.6%. The certified reference materials (CRM) used did not have certified values for Al, and the SAFe samples were not available at the time of analysis. However, the generally good agreement between data here and earlier measurement of Al in nearby waters supports the accuracy of the method.

2.3.3. Dissolved Manganese

[21] Dissolved Mn was determined using a luminol chemiluminescent technique based on the methods described by *Doi et al.* [2004], in which Mn is initially preconcentrated on Toyopearl 650 AF resin, prior to elution and separation from potentially interfering metal ions using an 8-hydroxyquinoline resin and determination using the luminol light-emitting reaction. Samples were individually buffered prior to analysis. Procedural blank was 40 pM with a precision of 20 pM. The detection limit (calculated as three times the standard deviation of the blank) was 60 pM with an average relative standard deviation for the profile samples at M2 at these low concentrations of 11.7%. Accuracy was determined using Certified International Standard Material NASS-5 (National Research Council of Canada) that was processed in the same way as the samples (Table 2).

3. Results and Discussion

3.1. Hydrography

[22] Sites M1 and M3 are bounded to the west and north by the SAF, thus they receive little influence from

Table 2. Trace Metal Values for Certified Reference Materials ($n = 3$)^a

	NASS-6 Certified Value	This Study	CASS-5 Certified Value	This Study
Ni	0.301 ± 0.025	0.286 ± 0.008	0.330 ± 0.023	0.302 ± 0.011
Zn	0.257 ± 0.020	0.200 ± 0.020	0.719 ± 0.068	0.664 ± 0.013
Co	0.015	0.015 ± 0.001	0.095	0.091 ± 0.002
Cd	0.0311 ± 0.0019	0.0350 ± 0.0033	0.0215 ± 0.0018	0.0232 ± 0.0029
Pb	0.006 ± 0.002	0.006 ± 0.001	0.011 ± 0.002	0.010 ± 0.001
Mn	NASS-5 certified value	This study		
	0.919 ± 0.057	0.917 ± 0.055		

^a95% of confidence limits are presented. All concentrations are in $\mu\text{g L}^{-1}$. Due to low recovery ($<90\%$), Cd data presented in this study and in this table have been recovery corrected.

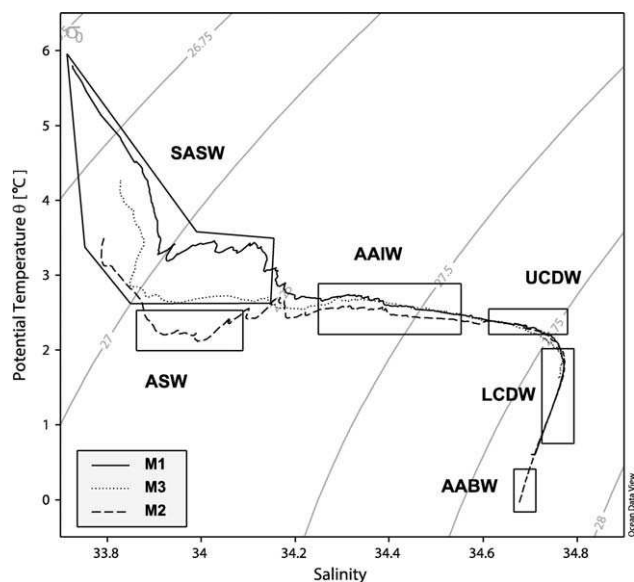


Figure 2. Potential temperature ($^{\circ}\text{C}$) versus salinity for sites M1, M3, and M2. Water mass abbreviations in alphabetical order: AABW: Antarctic Bottom Water; AAIW: Antarctic Intermediate Water; AASW: Antarctic Surface Water; LCDW: Lower Circumpolar Deep Water; SASW: Sub-Antarctic Surface Water; and UCDW: Upper Circumpolar Deep Water. Figure made with Ocean Data View (R. Schlitzer, 2011, Ocean data view, version 4.3.10, <http://odv.awi.de>).

subtropical waters (Figure 1). Nevertheless, there is evidence of eddies penetrating from the SAF [Pollard *et al.*, 2007a; Read *et al.*, 2007] causing the Chl-*a* patchiness observed in the satellite imagery [Venables *et al.*, 2007]. Surface waters ($S < 34.2$) (Figures 2 and 3) had similar physical properties to those found in a nearby station by Read *et al.* [2007] and were related to Sub-Antarctic Surface Water (SASW) (Figure 2). Upper 300 m of the water column at M3 was also identified as SASW by Charette *et al.* [2007] based on ^{226}Ra activities. Attribution of intermediate and deep waters at M3 is rather difficult and Charette *et al.* [2007] suggested a mixture of Antarctic Intermediate Water (AAIW) and underlying Circumpolar Deep Water (CDW). Macronutrients cyclings, ratios, and concentrations at M2 were typical of Southern Ocean HNLC regions [Pollard *et al.*, 2007b; Sanders *et al.*, 2007].

[23] M2 was, therefore, chosen as a control representative of typical HNLC conditions, although Argo floats show that water from the islands or the plateau may occasionally reach this site [Pollard *et al.*, 2007a]. Surface waters (Figures 2 and 3) possibly show the mixture of SASW and Antarctic Surface Water (AASW) as the subsurface minimum in the potential temperature ($\theta < 2.1^{\circ}\text{C}$) is typical for AASW of the Antarctic Zone (AAZ), which suggests that M2 is likely south of the PF. This is consistent with a detailed description of water masses around the Crozet Plateau and Basin reported by Park *et al.* [1993] and more recent findings by Read *et al.* [2007]. Intermediate waters extending from ~ 300 to ~ 800 m were assumed to belong to the relatively fresh AAIW in agreement with the water mass classification

adopted in a nearby station [Charette *et al.*, 2007]. The underlying water mass extending up to 3000–3500 m showed a gradual decrease in temperature and an increase in salinity with depth probably corresponding to Circumpolar Deep Water (CDW). This water mass often distinguished between Upper (UCDW) and Lower (LCDW) branches showed a salinity maximum ($S > 34.77$) at 2000 m likely due to the influence from more saline North Atlantic Deep Water (NADW). Below ~ 3500 m, colder and fresher ($\theta < 0.3$ and $S < 34.71$) water was found and attributed to Antarctic Bottom Water (AABW) as it is documented that AABW passes near Crozet on its way to the Indian Ocean [Mantyla and Reid, 1983; Park *et al.*, 1993].

3.2. Trace Metal Distributions

[24] Depth profiles for each of the trace metals (Ni, Zn, Co, Cd, and Pb) are plotted along with potential temperature for the three studied sites (M1, M2, and M3) in Figure 3. Al and Mn data are only available for M2 (Figure 3). Table 3 provides a general comparison of dissolved trace metal concentrations obtained in this study and previous work in the Southern Ocean. Macronutrients nitrate, phosphate, and Si are plotted in Figure 4. Nutrient-metal correlations are calculated in order to assess surface (upper 250 m, Table 4) biological uptake of bioactive trace metals in the bloom area and their recycling in deep waters. Errors included in the correlations are quoted to one standard deviation.

3.2.1. Nickel

[25] Vertical profiles for dissolved Ni showed surface depletion and regeneration with depth occurring along with macronutrients at all stations (Figures 3 and 4). Ni increased by ~ 1 nM with increasing depth and showed little variation below 1000 m. Dissolved Ni concentrations ranged from 4.6 to 6.3 nM (Figure 3) and are consistent with the recently reported concentrations of 3–6 nmol kg^{-1} for wintertime sub-Antarctic waters south of Australia [Ellwood, 2008], 2.2–6.6 nM for the upper 300 m in the Australian PFZ [Lai *et al.*, 2008] and earlier work in the Southern Ocean (Table 3).

[26] Strong correlation of Ni with nitrate (slope = 0.09, intercept = 3.03, $r^2 = 0.81$; $n = 50$; $p < 0.001$) and Ni with phosphate (slope = 1.23, intercept = 3.11, $r^2 = 0.77$; $n = 50$; $p < 0.001$) and moderate correlations of Ni with Si (slope = 0.01, intercept = 5.16, $r^2 = 0.58$; $n = 50$; $p < 0.001$) were found using the entire data set. Similar strong correlations with phosphate were reported for Australian sub-Antarctic waters [Butler *et al.*, 2013; Ellwood, 2008], whereas Lai *et al.* [2008] found much weaker relationships ($r^2 = 0.50$) in that region.

[27] Dissolved Ni was highly correlated with all macronutrients in the upper 250 m (Table 4) north of the islands (M1) where higher biomass and primary production is found [Seeyave *et al.*, 2007; Venables *et al.*, 2007] confirming surface Ni and macronutrient uptake within the bloom area (Figures 3 and 4, Table 4). The small depletion of Ni (typically 0.5–1 nM) in the bloom area north of Crozet is comparable to findings of Ellwood [2008] and Butler *et al.* [2013], who suggested a low biological requirement of Ni compared to other macronutrients in waters of the sub-Antarctic and Polar Frontal Zones south of Australia. This is corroborated by the positive intercept in excess of 3 nM calculated at M1 between Ni and all macronutrients

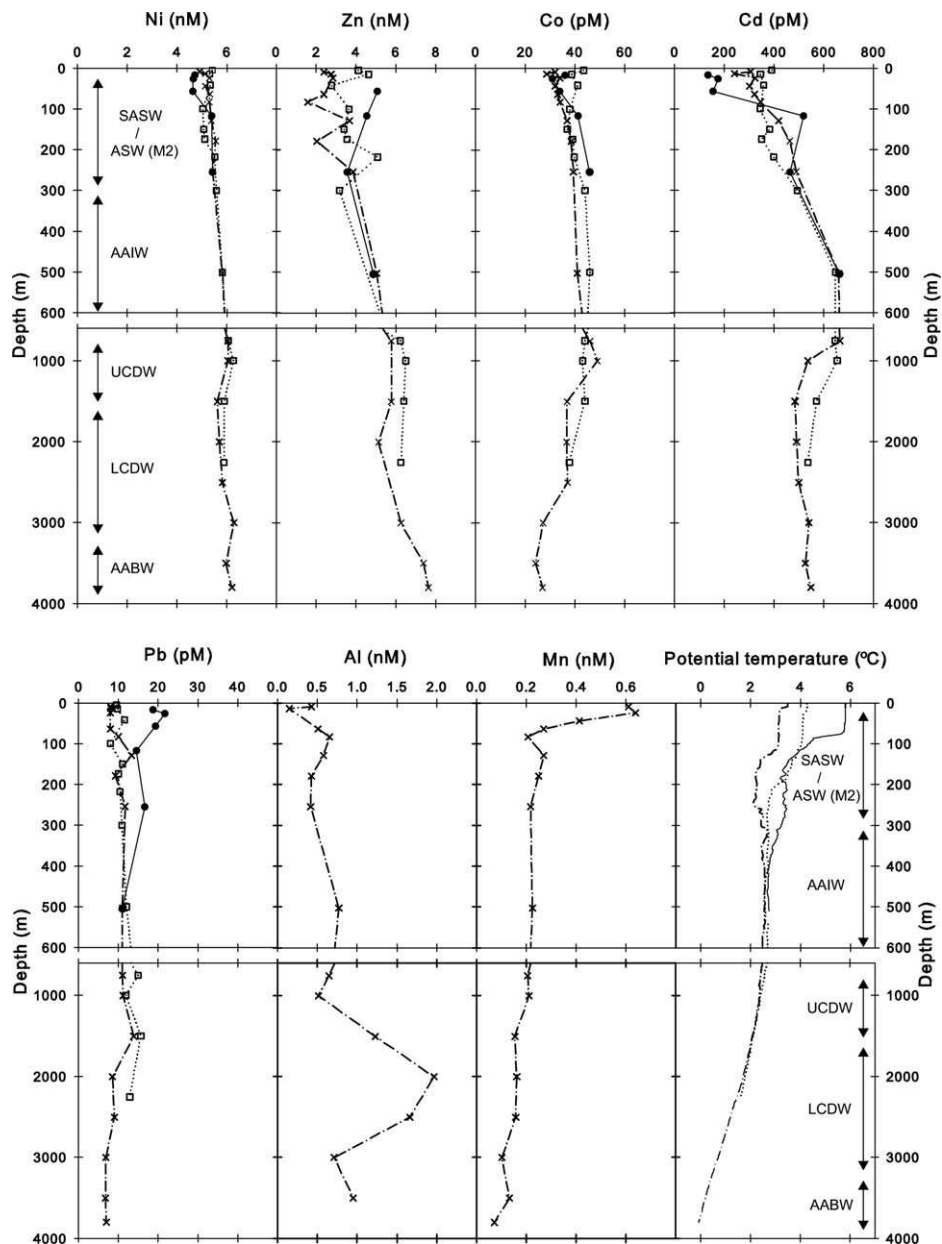


Figure 3. Vertical profiles of dissolved Ni, Zn, Co, Cd, Pb, Al, and Mn along with potential temperature for sites M1 (filled circle, solid line), M3 (empty square, dotted line), and M2 (cross, dash-dot line). Note the different scale on the y axis. Water mass abbreviations in alphabetical order: AABW: Antarctic Bottom Water; AAIW: Antarctic Intermediate Water; AASW: Antarctic Surface Water; LCDW: Lower Circumpolar Deep Water; SASW: Sub-Antarctic Surface Water; and UCDW: Upper Circumpolar Deep Water.

(Table 4) which indicates that an excess of Ni will persist even in the case of macronutrient depletion.

[28] Depletion of trace metals including dissolved Ni have been reported elsewhere, for example, during a spring diatom bloom in the PFZ [Löscher, 1999] and in an artificial Fe experiment in the HNLC sub-Arctic Pacific [Kinugasa et al., 2005], whereas no apparent Ni uptake was observed during the Southern Ocean Iron Release Experiment (SOIREE) in the Southern Ocean [Frew et al., 2001].

[29] Depletion of Ni and Si at M1 were largest relative to nonbloom sites (Figures 3 and 4), and high correlations

between the two elements in the upper 250 m were found only here (Table 4) suggesting that Ni uptake was linked to the presence of diatoms. At the time of sampling, the bloom was already in decline and the phytoplankton community was dominated by *Phaeocystis antarctica* but diatoms still accounted for 38% of the biomass at M1 [Poulton et al., 2007]. Furthermore, diatoms species *Eucampia antarctica* were dominant in sediment traps north of the plateau and accounted for 72–100% of the particle export [Salter et al., 2007]. The presence of diatoms was also confirmed during short-term incubation experiments on board where

Table 3. Comparison of Reported Ranges for Dissolved Ni, Zn, Co, Cd, Pb, Al, and Mn in the Southern Ocean

Study	Location	Season ^a	Depth ^b	Ni (nM)	Zn (nM)	Co (pM)	Cd (pM)	Pb (pM)	Al (nM)	Mn (nM)
This study	Indian sector, Crozet Islands	Spring	3800	4.64–6.31	1.59–7.75	24–49 ^c	135–673	6–22	0.13–2.15	0.07–0.64
<i>Butler et al.</i> [2013]	Australian sector	Late summer	~1000	2.51–7.31	<0.22–1.79	5–43	~4–900			
<i>Boye et al.</i> [2012]	Southeast Atlantic sector	Summer	~4000				4–895	7–55		~0.2–1.3
<i>Bown et al.</i> [2012]	Indian sector, Kerguelen Plateau	Summer	2500			19–375 ^c				
<i>Middag et al.</i> [2012]	Drake Passage	Autumn	4250						0.09–14.43	0.08–2.64
<i>Abouchami et al.</i> [2011] ^d	Atlantic sector, Zero meridian	late Summer	Surface				36–620			
<i>Bown et al.</i> [2011]	Atlantic sector, Zero meridian	Summer	4000			5–74 ^c				
<i>Croot et al.</i> [2011]	Atlantic sector	Autumn	>5000		0.21–7.00					
<i>Middag et al.</i> [2011a]	Atlantic sector, Zero meridian	Summer	5320						0.10–6.54	
<i>Middag et al.</i> [2011b]	Atlantic sector, Zero meridian	Summer	5320							0.05–3.20
<i>Saito et al.</i> [2010]	Ross Sea	Spring	>1400			40–85 ^c				
		Summer	>1400			20–60 ^c				
<i>Ellwood</i> [2008] ^d	Australian sector, sub-Antarctic zone	Winter	1000	3.00–7.00	0–3.00	10–50	20–650	12–33		
<i>Lai et al.</i> [2008]	Australian sector, 140° meridian	Summer	300	2.15–7.91				6–170		
<i>Coale et al.</i> [2005] ^d	Ross Sea and ACC (170° W)	Spring-summer	2000		0.18–6.54					
<i>Corami et al.</i> [2005] ^d	Ross Sea	Summer	1580		2.2–11.9		340–900	3–127		0.33–4.10
<i>Ellwood et al.</i> [2005]	Atlantic sector, across the PF	Autumn	1000			15–45 ^c				
<i>Abollino et al.</i> [2004]	Ross Sea	Spring	250	5.2–9.4			420–800			0.2–1.9
	Gerlache Inlet	Spring	380	3.5–11.5			100–580			0.5–2.5
<i>Sañudo-Wilhelmy et al.</i> [2002]	Weddell Sea	Summer	Surface	5.10–6.52	1.44–5.99	18–172	170–670	4–40	1.21–5.78	
<i>Frew et al.</i> [2001]	Australian sector, 61° S–140° E	Summer	40	5.7–6.6	1.9–2.5		120–280			
<i>Bucciarelli et al.</i> [2001]	Indian sector, Kerguelen Islands	Autumn	280							0.68–8.54
<i>Grotti et al.</i> [2001]	Gerlache Inlet	Spring	380							0.10–6.60
<i>Fitzwater et al.</i> [2000] ^d	Ross Sea	Summer	375	4.78–6.88	0.24–7.11	5–41	40–800			
<i>Sedwick et al.</i> [2000]	Ross Sea	Spring-summer	300							0.03–0.83
<i>Löscher</i> [1999]	Atlantic sector, 6° W meridian	Spring	4225	3.70–7.55	0.8–10					
<i>Löscher et al.</i> [1998]	Atlantic sector, 6° W meridian	Spring	4839			155–1309				
<i>Sedwick et al.</i> [1997]	Australian sector, sub-Antarctic zone	Summer	350							0.09–2.25
<i>van Beusekom et al.</i> [1997]	Indian sector, Enderby and Crozet Basins	Autumn	>5000						0.10–4.09	
<i>Nolting and de Baar</i> [1994]	Scotia-Weddell Seas	Spring	300	3.07–10.39	1.38–13.91		170–950			
<i>Moran et al.</i> [1992] ^c	Weddell Sea	Summer	4100						1.0–4.9	
<i>van Bennekom et al.</i> [1991] ^c	Scotia-Weddell Seas	Spring-summer	~4000						1.0–3.1	
<i>Westerlund and Öhman</i> [1991a]	Weddell Sea	Summer	3250							0.11–2.07
<i>Westerlund and Öhman</i> [1991b]	Weddell Sea	Summer	4000	5.08–8.23	1.88–8.00	12–58	480–970	~10		
<i>Nolting et al.</i> [1991]	Scotia-Weddell Seas	Spring-summer	3931				290–990			
<i>Martin et al.</i> [1990] ^d	Drake Passage and Gerlache Strait	Summer	1420			0.24–6.34	21–82	260–810		0.08–5.05

^aSeasons are given for the Southern Hemisphere.

^bMaximum sampled depth in meters.

^cUV-digested data for dissolved Co.

^dData per kilogram of seawater.

^eUnfiltered samples.

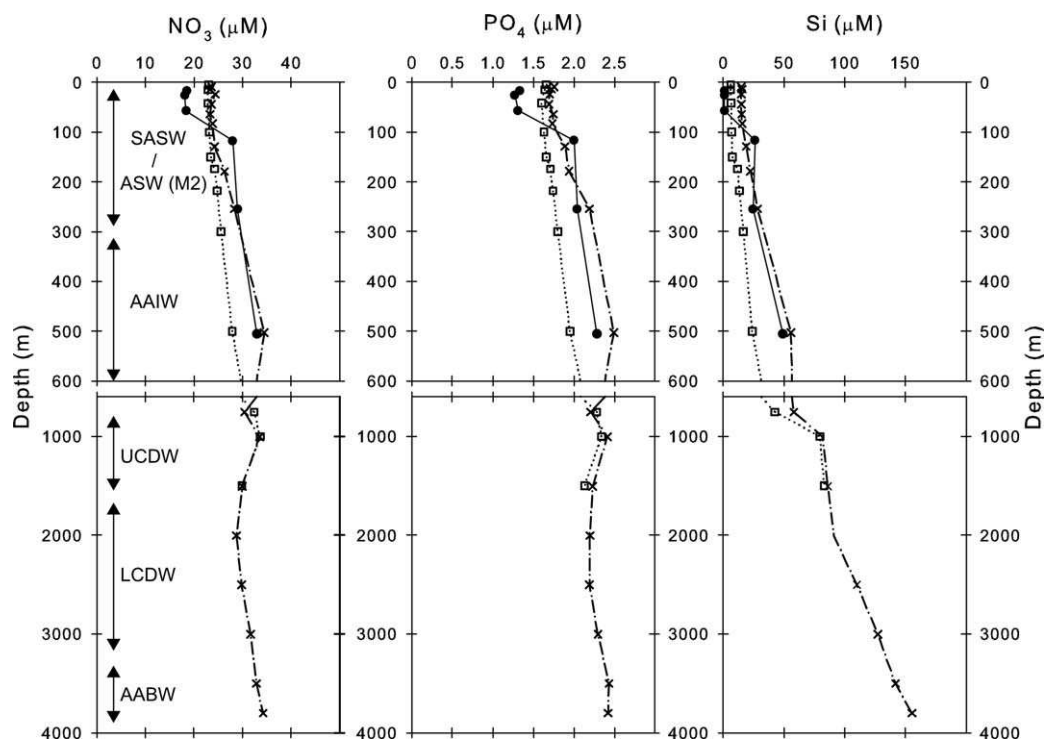


Figure 4. Vertical profiles of nitrate, phosphate, and Si for sites M1 (filled circle, solid line), M3 (empty square, dotted line), and M2 (cross, dash-dot line). Note the different scale on the y axis. Water mass abbreviations in alphabetical order: AABW: Antarctic Bottom Water; AAIW: Antarctic Intermediate Water; AASW: Antarctic Surface Water; LCDW: Lower Circumpolar deep Water; SASW: Sub-Antarctic Surface Water; and UCDW: Upper Circumpolar Deep Water.

medium-sized diatoms showed a strong response upon Fe addition suggesting that these organisms were important at the beginning of the bloom [Moore *et al.*, 2007]. Finally, synchrotron X-ray fluorescence analysis of phytoplankton cells has revealed that diatoms have 2 to 5-fold higher Ni quotas than other phytoplankton groups [Twining *et al.*, 2004, 2011, 2012] and that Ni is equally accumulated in the intracellular material and the frustules [Twining *et al.*, 2012], supporting the strong depletion observed at M1.

Table 4. Significant ($p < 0.01$) Correlations Between Dissolved Trace Metals and Inorganic Nutrients in the Upper 250 m of Crozet Waters^a

	Site	n	Slope	\pm	P Value	Intercept	\pm	R^2
Ni:P	M1	5	1.03	0.03	<0.001	3.33	0.05	0.99
Ni:N	M1	5	0.07	0.00	<0.001	3.33	0.06	0.99
Ni:Si	M1	5	0.03	0.00	<0.001	4.64	0.03	0.99
Co:P	M2	15	16	4	<0.01	4	7	0.54
Co:N	M2	15	2	0	<0.01	-6	10	0.57
Co:Si	M2	15	1	0	<0.01	24	2	0.56
Cd:P	M1	5	465	52	<0.01	-449	84	0.96
Cd:N	M1	5	33	4	<0.01	-446	86	0.96
Cd:Si	M1	5	14	1	<0.001	144	12	0.99
Cd:P	M2	15	441	66	<0.001	-447	119	0.77
Cd:N	M2	15	42	7	<0.001	-681	168	0.74
Cd:Si	M2	15	15	2	<0.001	85	36	0.80

^aOne standard deviation of slope and intercept are indicated in columns with \pm . Slopes are in nmol/ μ mol for Ni and pmol/ μ mol for Co and Cd. Intercepts are in nmol/L for Ni and pmol/L for Co and Cd.

Incorporation of Ni into frustules is not yet understood but it is known that Ni is required by diatoms for the assimilation of urea [Oliveira and Antia, 1984; Price and Morel, 1991] and replacement of Fe by Ni in enzyme superoxide dismutase (SOD) appears to be a common adaptation to low Fe concentrations [Cuevelier *et al.*, 2010]. Under these conditions, assimilation of Ni using divalent metal transporters [Agranoff *et al.*, 2005] in a similar way to Fe [Kustka *et al.*, 2007] could also be considered.

3.2.2. Zinc

[30] Vertical profiles showed a nutrient-like behavior of Zn with regeneration occurring together with inorganic nutrients and particularly with Si in surface and intermediate waters (Figures 3, 4 and 5) of all stations. From 1000 to 2000 m Zn was rather invariant (M2 and M3) and increased again along with Si reaching maximum concentrations in deep waters at M2.

[31] Dissolved Zn generally ranged between 2 and 6 nM (Figure 3) although, maximum concentrations as high as 7.75 nM were found at the greatest sampled depth (3750 m, M2) coinciding with a colder and fresher nutrient-rich water mass identified here as AABW (Figure 2). These deep water concentrations were ~ 1.5 nM higher than those reported for the Zero meridian across the PFZ [Croot *et al.*, 2011] but similar to those found in the Madagascar Basin that were also attributed to AABW [Morley *et al.*, 1993]. The range of our dissolved Zn concentrations is slightly higher in magnitude than the 0.5–6 nM range reported for the PFZ in the Atlantic sector (stations S103 & S104 of Croot *et al.* [2011]) but lower than the 0.4–12.1 nM

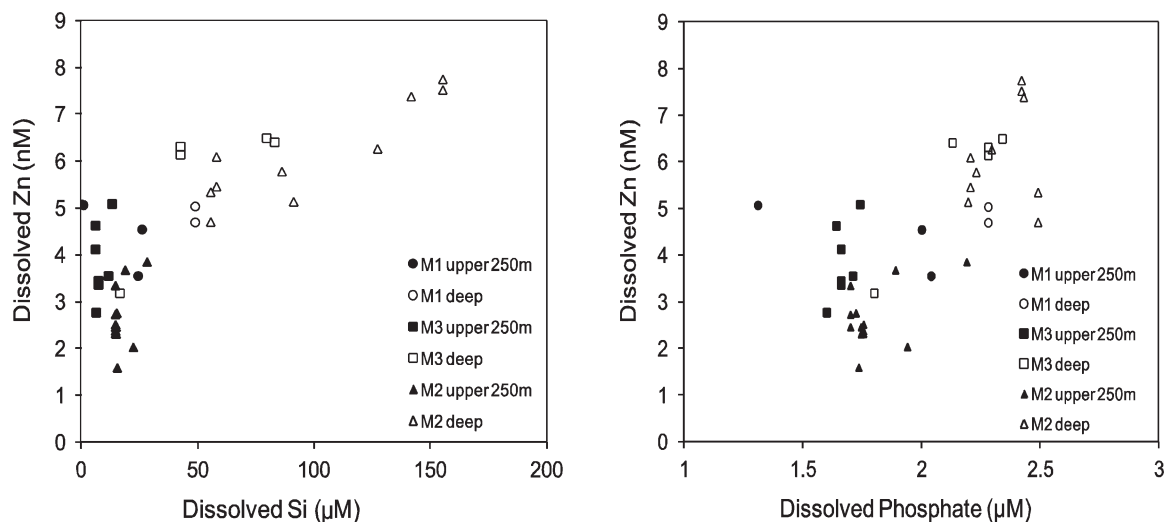


Figure 5. (left) Dissolved Zn (nM) versus Si (μM) and (right) dissolved phosphate (μM) for sites M1 (circle), M3 (square), and M2 (triangle). Filled and empty symbols indicate surface and deep water data, respectively.

for the southwestern Indian Ocean [Morley *et al.*, 1993]. Ellwood [2008] reported lower wintertime concentrations of 0.5–3.5 nmol kg^{-1} for the upper 500 m in the Australian sub-Antarctic zone and recent work by Gosnell *et al.* [2012] found Zn concentrations from 0.02 nM in the surface up to >3.5 nM at about 1300 m in the southern Indian Ocean. Their stations, however, were located further north and had less influence from nutrient and metal-rich Antarctic waters, therefore, higher Zn values around Crozet were expected. Other studies including dissolved Zn in the Southern Ocean generally obtained higher or similar values to this study (Table 3).

[32] At the time of sampling, surface Si reached limiting values of $<1 \mu\text{M}$ at M1 [Poulton *et al.*, 2007] and dissolved Fe concentrations were moderately limiting, with an average of 0.35 nM in the upper 250 m [Planquette *et al.*, 2007]. As mentioned above (section 3.2.1), diatoms were dominating the earlier stages of the bloom [Moore *et al.*, 2007; Poulton *et al.*, 2007; Salter *et al.*, 2007], which probably explains Si and Fe drawdown. Diatom growth li-

mitation by silicic acid in PFZ waters of the Crozet Basin has also been reported by Sedwick *et al.* [2002] and diatom cultures have shown that Zn is required for Si uptake [De La Rocha *et al.*, 2000; Rueter and Morel, 1981]. Unfortunately, no enhanced biological uptake of Zn nor strong correlations with Si were appreciable in surface waters at M1 due to analytical scatter and limited data (some surface samples appeared to be contaminated during sampling or handling and did not prove to be oceanographically consistent. Details can be found in supporting information). The moderate correlation of Zn with Si (Figure 5) when considering the whole data set (slope = 0.032, intercept = 3.02, $r^2 = 0.66$; $n = 40$; $p < 0.001$), however, suggests that likely diatoms significantly influenced the biogeochemical cycling of Zn around Crozet.

[33] Croot *et al.* [2011] provide a comparison of Zn:Si ratios with particular emphasis on the Southern Ocean. Our low overall ratio of 0.032 $\text{nmol Zn}:\mu\text{mol Si}$ found in Crozet waters was similar to the ratio of 0.033 $\text{nmol Zn}:\mu\text{mol Si}$ reported for productive waters of the ACC [Löscher, 1999]

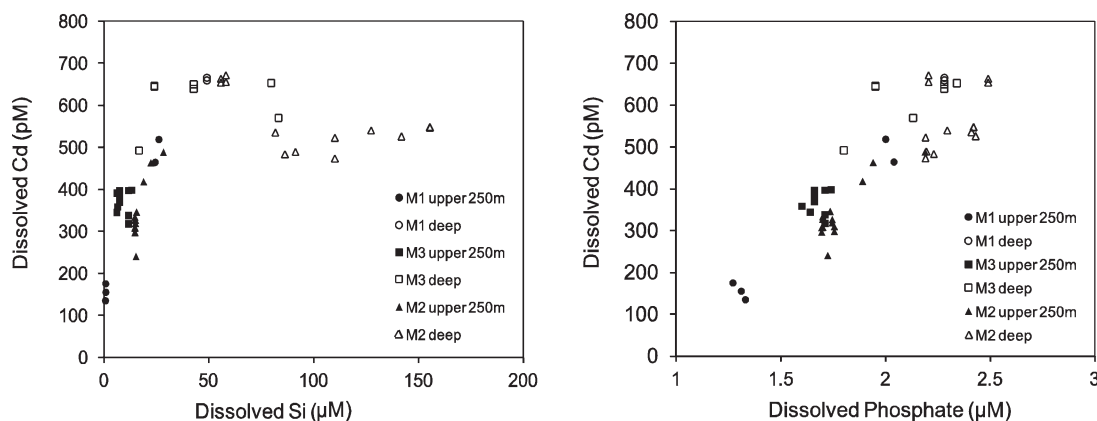


Figure 6. Dissolved Cd (pM) versus (left) Si (μM) and (right) dissolved phosphate (μM) for sites M1 (circle), M3 (square), and M2 (triangle). Filled and empty symbols indicate surface and deep water data, respectively.

but lower than for the southern Indian Ocean [Gosnell *et al.*, 2012], the Zero meridian [Croot *et al.*, 2011] and the Australian sub-Antarctic zone [Ellwood, 2008]. Waters north of Crozet receive a natural supply of dissolved Fe (Fe concentrations of 0.35 nM in the upper 250 m of station M1) that supports high primary production rates during spring and summer [Planquette *et al.*, 2007] which may explain the lower utilization of Zn compared to other Fe deprived HNLC waters in the Southern Ocean.

[34] Furthermore, Zn concentrations at M3 were ~ 0.7 nM higher than at M1 and M2 throughout most of the water column. Although this increase is not clearly related to an island source, sediments from Crozet plateau and islands have been shown to supply dissolved and particulate Fe [Planquette *et al.*, 2009; Planquette *et al.*, 2007], and to a lesser extent, particulate aluminium [Planquette *et al.*, 2009]. Similarly, higher Zn values were reported and associated with advective inputs from coastal and shelf processes in the sub-Arctic northeast Pacific [Lohan *et al.*, 2002]. Horizontal advection is important in the vicinity of Île de la Possession and Île de l'Est, as well as at M3 [Charette *et al.*, 2007] and constitutes the main input of Fe [Planquette *et al.*, 2007], indicating that Crozet could act as source of Zn. The fact that Fe [see Planquette *et al.*, 2007] and Zn have similar variations in the upper 250 m of M3 strengthens this hypothesis. Upwelling of deep waters is highly improbable at M3 due to its proximity to a steep bathymetric change [Pollard *et al.*, 2007a; R. Pollard, personal communication, 2011].

3.2.3. Cobalt

[35] Vertical profiles of Co reflected different behaviors in the three distinct biogeochemical regions around Crozet. Overall, dissolved Co concentrations ranged from 24 to 49 pM (Figure 3). Since the samples were UV irradiated, they are deemed to be true total dissolved values. UV digested concentrations for dissolved Co were reported elsewhere: Saito *et al.* [2010] reported concentrations in the Ross Sea between 20 and 85 pM during spring-summer and Bown *et al.* [2011] reported a range of 5–74 pM during the summertime in the southeast Atlantic sector. In the sub-Antarctic zone of the Australian sector, non-UV irradiated concentrations of dissolved Co ranged from 10 to 50 pmol kg⁻¹ in the wintertime [Ellwood, 2008] and were comparable to the range of 5–43 pM found during late summer by Butler *et al.* [2013]. Our data are in good agreement with these values and in line with other published data for the Southern Ocean (Table 3). Finally, Bown *et al.* [2012] reported concentrations as high as 375 pM around Kerguelen and argued an island source of Co to adjacent waters. Such an island source is not obvious at our offshore sampling sites around Crozet.

[36] In this study, surface Co concentrations were more depleted at M2 (~ 34 pM) than at M1 (38 pM) and M3 (40 pM). However, the magnitude of the depletion compared to the average mesopelagic concentration was greater at M1 than at M2, probably indicating biological uptake. Lower surface Co concentrations related to biological uptake have been previously reported in the sub-Antarctic zone [Bown *et al.*, 2011; Ellwood, 2008] and the Ross Sea [Saito *et al.*, 2010]. Such an uptake is visible at M1, where Co is moderately correlated with all macronutrients in surface (correlation with phosphate: $r^2 = 0.87$, $n = 5$; nitrate: $r^2 = 0.87$, $n = 5$; Si:

$r^2 = 0.80$, $n = 5$; $p < 0.05$). At M2, lower correlations were found (Table 4), which is consistent with this station being a HNLC site. At M3, dissolved Co concentrations were rather invariant throughout the water column and no correlation was noted between dissolved Co and macronutrients, probably due to the strong vertical mixing observed by Charette *et al.* [2007]. Alternatively, dissolved Co might have been strongly scavenged in the euphotic layer by a higher vertical particles export highlighted by Morris *et al.* [2007], essentially at station M1. Finally, the absence or the lack of organic ligands may also have limited the stabilization of Co in the dissolved fraction, thus decreasing its residence time [Bown *et al.*, 2012]. At M2, a strong recycling component in the upper 1000 m can be observed then followed by a decrease with increasing depth (Figure 3). Scavenging processes likely explain most of the distribution of dissolved Co concentrations at intermediate and deep waters, particularly at M2. At this station, intrusions of CDW at about 1000–1500 m, then AABW at 3000–3500 m, may explain the observed distribution. CDW and AABW should have been more exposed than AAIW to the removal by sinking particles during their longer circulation pathways. Removal of dissolved Co linked to adsorption onto particles has been also suggested by Bown *et al.* [2011] to explain the lower Co concentrations found in deep waters of the Antarctic Circumpolar Current. Overall, Co data presented in this study generally agree very well with recent data reported for the Atlantic sector [Bown *et al.*, 2011], where scavenging and mixing of water masses have been shown to exert a major control on the vertical distribution of Co within the PFZ.

3.2.4. Cadmium

[37] Vertical profiles of Cd showed the strongest surface drawdown and shallowest regeneration among the targeted metals of this study (Figure 3). Below 1000 m, Cd concentrations decreased by ~ 100 pM and remained constant before slightly increasing again at greatest depths.

[38] Dissolved Cd concentrations ranged from 135 to 673 pM (Figure 3) which is in line with the recently reported range of 36–620 pM for the Australian sub-Antarctic zone [Ellwood, 2008] and other areas of the Southern Ocean (Table 3).

[39] As with Ni, lowest surface concentrations for Cd were found at M1. At this station, the decrease in the top 250 m of Cd and Co corresponds to an increase in Zn concentration. This might be explained by the fact that Zn, Co, and Cd can all act as a substitute into the active site of the enzyme carbonic anhydrase, depending on the species [Sunda, 2012]. Cd was also well correlated with macronutrients phosphate and Si (Figure 6) and, with nitrate in the upper 250 m of Station M1 (Table 4). In the upper 250 m of Station M3, Cd/phosphate ratio was on average of 0.441 nmol Cd:μmol phosphate. These strong correlations indicate that the depletion of Cd of >100 pM and subsequent recycling are driven by biological processes [Lane and Morel, 2000]. Phytoplankton uptake of Cd has been reported in sub-Antarctic waters south of Australia [Ellwood, 2008], in artificial Fe experiments [Frew *et al.*, 2001; Kinugasa *et al.*, 2005] and is evident in bottle incubations [Nakatsuka *et al.*, 2009].

[40] The slope and nonzero intercept found for Cd against phosphate (Table 4) is typical of the Southern Ocean [de Baar *et al.*, 1994] and similar to the one reported

by *Ellwood* [2008] in the sub-Antarctic region. Table 4 and Figure 6 suggest that Cd uptake is enhanced to that of phosphate in HNLC waters with limiting Fe [*Cullen et al.*, 2003]. Stations analyzed here were sampled once the main bloom collapsed [*Venables et al.*, 2007] and some sites such as M3 already showed Fe limiting conditions [*Planquette et al.*, 2007]. Explanation for higher Cd concentrations at M3 compared to M2 at depths of 1000–2500 m is not straightforward and possibly implies a combination of a sedimentary source (based on some small increases in dissolved Cd observed close to the islands) and the general sinking trend of Antarctic waters as they move northward.

[41] The transition from AAIW to CDW may also explain the lower Cd found below 1000 m as it has been suggested that Cd is consumed during the northward transport of the CDW [*Abouchami et al.*, 2011]. Although not as clearly as for Zn, the increase of Cd at greatest depths also seem to be related to the presence of AABW, indicative of the importance of water masses defining the trace metal concentrations around Crozet and particularly in deep waters where biological activity is negligible.

3.2.5. Lead

[42] Dissolved Pb decreased with depth at M1 which is likely due to scavenging processes related to the higher export of particles in the bloom [*Morris et al.*, 2007]. In contrast, concentrations at M3 and M2 were constant throughout most of the water column and only decreased at deeper depths (Figure 3).

[43] Dissolved Pb concentrations ranged from 6 to 22 pM (Figure 3) with lowest concentrations found in deep waters at M2 (Figure 3) probably related to older waters that suffered from scavenging over longer time scales such as AABW, whereas highest concentrations were measured in the surface layer at M1. Our range for dissolved Pb concentrations is consistent with reported values of 11–21 pmol kg⁻¹ [*Ellwood*, 2008] and 9–22 pM for the Australian PFZ (JARE-43 cruise, *Lai et al.* [2008]). *Lai et al.* [2008] also provided values as high as 165 pM associated to episodic dust events during the KH-01-03 cruise. Other studies in the Southern Ocean (Table 3) were also in agreement with these data.

[44] A north-south gradient was observed in surface waters with M1 showing Pb concentrations twice as high as for M3 and M2 (Figure 3). Because concentrations around Crozet were homogeneous throughout the water column or decreased with depth, upwelling could not explain such a surface maximum at M1. Given the anthropogenic nature of Pb and the remoteness of the sampling area, an atmospheric source seems the most likely explanation for such high surface values. Model estimations suggest little atmospheric input to the Southern Ocean [*Jickells et al.*, 2005] and air parcel back trajectories indicated no contact with other land masses than Antarctica at least 5 days prior to the sampling [*Planquette et al.*, 2007]. Nevertheless, it is possible that an air mass which has been in contact before with the African continent reached the Crozet region after the main nutrient drawdown, thus explaining lack of significant surface Pb removal. There is evidence of Pb enriched dust supply along 30°S in the southwestern Indian Ocean that was attributed to southerly and westerly air mass trajectories likely originating from South Africa [*Witt et al.*,

2006]. M3 and M2 were 260 and 475 km southeast of M1, possibly enough to explain such a north-south gradient with lower Pb concentrations in the south. Recently, *Heimbürger et al.* [2013] reported daily deposition fluxes at Crozet Islands for several elements, including Pb (246 pmol m⁻² d⁻¹). Using a K_z of 11 cm² s⁻¹ [*Charette et al.*, 2007] and a vertical Pb gradient of 4.6 pmol m⁻³ m⁻¹, the vertical input of Pb at station M3 would account for 438 pmol m⁻² d⁻¹, which is in a similar order of magnitude than the atmospheric flux calculated by *Heimbürger et al.* [2013].

3.2.6. Aluminium

[45] Dissolved Al concentrations at M2 were low (Figure 3) and ranged from 0.13 to 2.15 nM which is in line with values reported (Table 3) for the Crozet Basin region [*van Beusekom et al.*, 1997] and more recent data for the Southern Ocean [*Middag et al.*, 2011a].

[46] In the upper 1000 m, dissolved Al remained below 0.60 nM. Then Al concentrations increased with depth reaching maximum concentrations of 2.15 nM at about 2000 m that coincided with a maximum in salinity associated to LCDW influenced by NADW. Higher Al concentrations within the LCDW due to mixing with NADW have been reported before in the PFZ of the Atlantic sector [*Middag et al.*, 2011a] and in the S11 profile of *van Beusekom et al.* [1997].

[47] The lower Al concentration at about 3000 m could be either the source concentration of the AABW or, the consequence of removal by scavenging processes which often influence the fate of Al in the water column [*Orians and Bruland*, 1985], whereas the small increase observed at the greatest sampled depth possibly reflects the picking up of Al from contact with sediments. This sedimentary source likely originates from Del Caño Rise rather than from the Crozet Islands since site M2 is shown to have very low particulate Al inputs at least in the upper water column [*Planquette et al.*, 2009]. Dissolved Al and Si were uncoupled and did not show a significant correlation ($r^2 = 0.36$; $p < 0.001$) throughout the water column and in benthic waters, where Al showed a relative small increase compared to Si, which is not what was observed by *Middag et al.* [2009]. This might be due to the fact that this station did not have enhanced biological activity, being in a HNLC zone.

3.2.7. Manganese

[48] Dissolved Mn concentrations to the south of the Crozet Islands (Site M2, Figure 3) were very low and varied from 0.64 nM in the surface to ~70 pM at the deepest point which is consistent with other trace metal work in the Southern Ocean (Table 3). This station showed a typical oceanic profile, with high surface concentrations followed by scavenging with depth [*Landing and Bruland*, 1980]. Elevated surface concentrations probably show the combination of aeolian inputs [*Landing and Bruland*, 1980], also seen for Al, but practically unnoticeable for Pb (at M2) and Fe [*Planquette et al.*, 2007] and, the photo reduction of Mn oxides [*Sunda and Huntsman*, 1988, 1994; *Sunda et al.*, 1983]. The low Mn concentrations below 100 m were likely linked to the sinking and scavenging of Mn oxides [*Statham et al.*, 1998; *Sunda and Huntsman*, 1988; *Sunda et al.*, 1983], particularly in older waters masses such as CDW and AABW that have been influenced by particle

flux and Mn oxidation over longer time scales. *Middag et al.* [2011b], report a similar general pattern with a surface maximum of around $\sim 0.4 \text{ nmol kg}^{-1}$ rapidly decreasing to values below 0.15 nM in the PFZ.

4. Conclusions

[49] Dissolved Ni, Zn, Co, Cd, Pb, Al, and Mn distributions were affected by biological uptake, scavenging processes, the Crozet Islands, and possible atmospheric inputs. These processes led to different biogeochemical regimes. The HNLC control zone to the south showed no apparent island influence and trace metal concentrations in the range of other areas of the Southern Ocean. Mixing of water masses of different age appear to influence the vertical distribution of the metals to the south and near Crozet Islands, particularly at intermediate and deep layers where biological activity is not significant. In the bloom area, significant surface depletion was found for Cd and to a less extent for Ni indicating the importance of biological processes such as the active uptake in their biogeochemical cycling. Concentrations of Ni in the bloom and concentrations of Zn around Crozet appear to be related to the cycling of Si and diatoms. Even though Crozet is truly remote, dissolved Pb, Al, and Mn results suggest that possible atmospheric inputs in surface waters could have led to small increases in trace metal concentrations especially to the north. Zn and Cd concentrations were significantly higher near the islands and could be that Crozet is acting as a source in a similar manner to Fe.

[50] **Acknowledgments.** We would like to thank the captain and crew of the *RRS Discovery* for their brilliant work at sea. Thanks as well to the scientific crew that provided the inorganic nutrient data. This cruise was the first of the CROZEX project which was a component of the British BICEP (Biophysical Interactions and Control of Export Production) program supported by NERC (Natural Environmental Research Council). We finally acknowledge Peter Croot and the anonymous reviewers for their very constructive comments on this manuscript.

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