

Modelling combined effects of temperature and pH on the heat resistance of spores of Bacillus cereus

Stéphane Gaillard, Ivan Leguérinel, Pierre Mafart

To cite this version:

Stéphane Gaillard, Ivan Leguérinel, Pierre Mafart. Modelling combined effects of temperature and pH on the heat resistance of spores of Bacillus cereus. Food Microbiology, 1998, pp.625-630. hal-00653497

HAL Id: hal-00653497 <https://hal.univ-brest.fr/hal-00653497>

Submitted on 20 Dec 2011

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.

Abstract

 The development of relationships between the pH of a heating medium and the thermal resistance of contaminant microorganisms is important and of a public health significance. A number of mathematical models have been presented in recent years, including that of Mafart and Leguérinel (1998) . However, in this This model , the effect of possible interactions between temperature and pH on D-values was not assessed. The consequences of ignoring interaction terms in models has been assessed and a comparison with Mafart's model that includes an interaction term showed that interaction terms can be neglected and that Mafart's model can be used in thermal process calculations. It appears possible 19 to adopt a standard value of z_{pH} , for example the 3.6 value and the conventional concept of biological destruction value L(T) (ratio of the sterilization value and the exposure time at a fixed heating temperature) may then be extended to 22 L(T,pH) (the same ratio at a fixed temperature with a fixed pH of the heating menstruum).

Keywords:

-
-

Temperature, pH, Heat resistance, Spores, *Bacillus cereus*.

Introduction

 Interactions between heating temperature and pH are frequently observed. In most cases, increasing temperature reduces the effect of pH. This type of interaction can be seen in the data of Xezones and Hutchings (1965) for *Clostridium botulinum*. This same behaviour is observed for *Bacillus stearothermophilus* (Rodrigo et al., 1993; Fernandez et al., 1994; Lopez et al., 1996). Rodrigo et al.(1993) observed that pH did not have any effect on the DT values of spores of *Clostridium sporogenes* when they were heated at high temperature. These results do not agree with those of Cameron et al.(1980) from which it can be shown that increasing temperature enhances the effect of pH. Acidifying the heating medium generally results in a decreased effect of temperature: an increase of the z value is observed when the pH of the medium decreases. This behaviour was showed for *Bacillus stearothermophilus* (Fernandez et al., 1994; Lopez et al., 1996), *Bacillus subtilis* (Condon and Sala, 1992), *Bacillus licheniformis* (Palop et al., 1992) and *Clostridium sporogenes* (Silla Santos et al.1992). However, Cameron et al.(1980) pointed out an opposite effect for *Clostridium sporogenes*(a decrease of the z value with decreasing pH), that disagrees with results of Silla Santos et al.(1992) .

 The mathematical relationship between the pH of the heating medium and the thermal resistance of microorganisms has been poorly documented. Jordan and Jacobs (1948) found a linear relationship between pH and the logarithm of the DT values for *E. coli*. Davey et al.(1978) developed the first model for predicting the combined effects of both process temperature and pH on thermal resistance of bacteria. This empirical model , which was developed from data of Xezones and Hutching (1965) for inactivation of *Clostridium botulinum,* shows a satisfactory goodness of fit and is relatively parsimonious (4 parameters for 2 factors). However it can be shown that Davey's model is still over parameterized (see appendix 1). The four empirical coefficients of the model have no biological significance. As an alternative, Mafart and Leguérinel (1998) proposed a new model with only three parameters, each having a physicochemical significance:

$$
\log D_{(T,pH)} = \log D_{(T^*,pH^*)} - \frac{T - T^*}{z_T} - \frac{(pH - pH^*)^2}{z_{pH}^2} \tag{1}
$$

65 Where T^* is the reference temperature (for example 121.1^oC), pH^{*} is the pH of 66 maximal heat resistance of spores (generally pH 7), z_T is the conventionnal 67 thermal z-value, z_{pH} is the distance of pH from pH* which leads to a ten fold 68 reduction of D value. Lastly, $D_{(T^*, pH^*)}$ is the D value at T^{*} and pH^{*}.

 This model was derived from the same data used by Davey et al., 1978 (i. e. Xezones and Hutching, 1965) and from two other sets of data related to *Clostridium sporogenes* (Cameron et al., 1980) and to *Bacillus stearothermophilus* (Lopez et al.,1996).

 Neither Davey's nor Mafart's models took into account interactions between temperature and pH on thermal resistance of spores. This paper aims to check from our own data related to *Bacillus cereus* that the lack of term accounting for interactions in the Mafart's model can be justified.

 The standard z-value used to compute most heating process is 10°C because it nearly corresponds to the value for *Clostridium botulinum* spores and 79 is closed to z_T -value of most other types of bacterial spores. Another aim of this 80 paper is to check that the z_{pH} value of *Bacillus cereus* is in the same range than that of others studied types of spores. If this paper, confirmed by further works, 82 shows that most of z_{pH} values are close to that of *Clostridium botulinum*, it 83 would be possible to adopt a standard z_{pH} for calculations of heat treatements.

Materials and methods

Microorganism and spore production

 The strain of *Bacillus cereus* (CNRZ 110) was obtained from the Institut National de Recherche Agronomie (France). Spores were kept in distilled water at 4°C.

 Cells were precultivated at 37°C during 24 h in Brain Heart Infusion (Difco). The preculture was used to inoculate nutritive agar plates (Biokar 92 Diagnostics BK021) added with MnSO₄ 40mg 1^{-1} and CaCl₂ 100 mgl⁻¹ on the surface aera. Plates were incubated at 37°C for 5 days. Spores were then collected by scraping the surface of the agar and suspended in sterile distilled water and washed three times by centrifugation (10000xg for 15 min) (Bioblock Scientific, model Sigma 3K30). The pellet was then resuspended in 5 ml distilled water and 5 ml ethanol. The obtained suspension was then kept at 4°C during 12 hours in order to eliminate vegetative non sporulated bacteria, and washed again three times by centrifugation. The final suspension (about 10^{10}) 100 spores ml^{-1}) was at last distributed in sterile Eppendorfs microtubes and kept at $4^{\circ}C$.

Thermal treatment of spore suspension

 $D_{(T,pH)}$ values in citrate-phosphate buffers adjusted respectively to 4.1, 4.5, 5.5, 6.5 and 6.9 were determined for temperatures of 86.6°C, 89°C, 95°C, 101°C and 103.4°C with one replicate at each temperature and pH combination. The whole set of data involves a complete 3x3 factorial design from 89°C to 101°C and from pH4.5 to pH 6.5 and in order to extend the range of validity of the model, the four following combinations were added: pH 5.5 at 86.6°C and 110 103.4 °C, pH 4.1 and 6.9 at 95 °C. First, 30µl of spore suspension was diluted in 3 ml buffer. Capillary tubes of 25

 µl (vitrex) were filled with 10µl of sample and submited to a thermal treatment in a thermostated oil bath. After heating, the tubes were cooled in water/ice bath, washed in a solution of soap and rinsed with sterile distilled water. Finally, ends were flamed with ethanol. The capillary tubes were broken at both ends and their contents poured into a tube containing 9 ml sterile tryptone salt broth (Biokar Diagnostics) by rincing with 1 ml tryptone salt broth contained in a syringe equipped with a needle.

Viable spore count

 The viable spores were counted by duplicate plating in nutritive agar (10g tryptone, 5g meat extract, 5g sodium chloride, 15 g agar for 1000ml water)(Biokar Diagnostic) and incubating at 30°C for 48h.

Data analysis

126 DT values were based on the reciprocal of slopes obtained when the log number of survivors was plotted against time. Multiple linear regressions used to fit the model were carried out with the STAT-ITCF software (Institut Technique du Fourrage France).

 The goodness of fit of the model was evaluated by using the per cent 132 variance accounted for (Snedecor and Cochran, 1969) which is given by:

$$
R^{2} = 1 - \frac{(1 - r^{2})(n - 1)}{(n - N - 1)}
$$

where n is the number of observations, N the number of terms and r^2 is the multiple regression coefficient.

141 **Results**

142

 Raw data of the experimental design are shown in Table 1. As the thermal resistance of *Bacillus cereus* is relatively low, we adopted the standard 145 temperature $T^*=100\degree C$ instead of 121.1°C. On the other hand, a preliminary experiment showed that the pH of maximal thermal resistance related to the studied strain of *Bacillus cereus* was close to 7.5 (data not shown). The model 148 was then fitted with pH^{*}=7.5. Fitted parameters according to equation 1 were:

149

150 $D_{(T^*, pH^*)} = 60$ seconds

$$
z_r = 9.15^{\circ}C
$$

$$
z_{\mu} = 3.70
$$

153 with a per cent variation accounted for $R^2 = 97.7\%$.

154 Families of graphs of $D_{(T,pH)}$ as a function of temperature at constant values of 155 pH and as a funtion of pH at constant values of temperature are shown in 156 Figures 1 and 2 respectively. Decreasing pH reduced the effect of temperature: 157 at pH 6.5, the z_T value was 7.97°C whilst it becomes 9.27°C and 10.3°C at 158 pH5.5 and 4.5 respectively. At 95 $^{\circ}$ C and 101 $^{\circ}$ C, the z_{pH} value remains constant 159 and equal to 3.87. On the other hand, a decrease of the z_{pH} value is observed at 160 89 $^{\circ}$ C (z_{pH} =3.06). It is then clear that the heat resistance of spores depends not 161 only on temperature and pH of the medium, but also on interactions between 162 these two factors. In order to take into account interactions, the model (1) was 163 modified by adding an interaction term :

164

$$
\log D_{(T,pH)} = \log D_{(T^*,pH^*)} + C_1(T - T^*) + C_2(pH - pH^*)^2 + C_3(T - T^*)(pH - pH^*)^2
$$

166 167 (2)

168 Where C_1 , C_2 and C_3 are empirical parameters without known physiological 169 significance. Fitted parameters according to this new model were:

170 $D_{(T^*, pH^*)} = 83$ seconds 171 $C_1 = -0.2900^{\circ}C^{-1}$ $C_2^{\prime} = -0.0281$ $C_3 = 0.1807$ °C-1 with a per cent variation accounted for $R2 = 98.5\%$. 175 176 The new interaction term was highly significant $(P<0.01\%)$ The comparaison between goodness of fit of both models (equations 1 and 2) are illustrated by Figures 3 and 4.

4. Discussion

 Variation of the thermal resistance of *Bacillus cereus* spores as a function of temperature and pH is similar to those for many bacterial spores. It is confirmed 184 that the pH of the heating medium has a prominent effect on the $D_{(T,pH)}$ values. For example, at 95°C a decrease of D-value of 5.7 times can be observed when the pH decrease from 6.9 to 4.1. As for of most bacterial spores, an increase of the temperature results in a decrease in the effect of the pH while a decrease of the pH medium results in a decrease of the effect of the temperature. This 189 behaviour means that some interactions on $D_{(T,pH)}$ -values exist between $D_{(T, pH)}$ -values exist between temperature and pH.This is confirmed by the fact that the interactions term of equation (2) is highly significant. However, adding one more parameters to our model in order to account for interactions results in a poor improvement of its goodness of fit : (with a R²-value of 0.985 instead of 0.977). Moreover, in 194 Equation 2, only one parameter ($D_{(T^*, pH^*)}$) keeps a biological significance: because of the occurence of the interaction term, it is not possible to reparameterize C₁ and C₂ into z_T and z_{pH} respectively. Fernandez et al.(1996) proposed two models for combined effects of temperature and pH on the thermal resistance of *Bacillus stearothermophilus* and *Clostridium sporogenes*: one a polynomial quadratic model including an interaction term (6 parameters), the other a simple linear model without interaction terms (3 parameters). From their data it may be concluded that the slight improvement of goodness of fit obtained by the quadratic model with respect to the linear model does not justify the choice of the 6 parameters model rather than that of 3 parameters model. The linear model of Fernandez et al. is of the form:

 $\log K = a + bT + cpH$ (3)

 where K is the death rate of spores and a, b and c empirical coefficients. It can be easly shown (see Appendix 2) that equation(3) can be reparameterized to give:

208 give:
\n
$$
\log D = \log D^* - \frac{1}{z_T} (T - T^*) - \frac{1}{z_{pH}} (pH - pH^*) \quad (4)
$$

 Disregarding pH terms, both models (1) and (4) are reduced to Bigelow's equation. However equation (1) where the pH term is squared differs from the 213 simple linear model (equation (4)) and is the only one to take the sigmoïdal 214 pattern of D values curves versus pH into account.

215 The z_{pH} value (3.7) related to *Bacillus cereus* have the same magnitude 216 that the observed values for *Clostridium botulinum* (3.56-3.61), *Clostridium* 217 *sporogenes* (3.33-4.29) and *Bacillus stearothermophilus* (2.94-3.97) (Mafart 218 and Leguérinel, 1998). Like conventional z_T values of spores close to 10^oC and 219 keep generally the range of $8^{\circ}C$ to $12^{\circ}C$, it appears that z_{pH} values are, in most 220 situations, included in a 3 to 4 range. If this observation is confirmed, it may be 221 possible to adopt a standard value of z_{pH} (for example the 3.6 value of 222 *Clostridium botulinum*) in order to include the pH in thermal process 223 calculations. The sterilization value could be defined as a heat treatment 224 equivalent to an exposure timeof 1 min at 121.1°C and at pH 7. The 225 conventional concept of biological destruction value L(T) could then be 226 extended to $L(T,pH)$ with

$$
L(T, pH) = 10^{\frac{T-T^*}{z_T} + \left(\frac{pH - pH^*}{z_{pH}}\right)^2}
$$
(5)

228

229 This function corresponds to the exposure time, at a T temperature and at a 230 given pH, which would be necessary in order to obtain one sterilization unit.

 Cameron M., Leonard S. and Barret E. (1980) Effect of moderately acidic pH on heat resistance of *Clostridium sporogenes* spores in phosphate buffer and in buffered pea puree. Appl. *Environ. Microbiol*. **39**, 943-949. Condon S. and Sala F. J. (1992) Heat resistance of *Bacillus subtilis* in buffer and foods of different pH. *J. FoodProtect*. **55**, 605-608. Davey K. R., Lin S. H. and Wood D. G. (1978) The effect of pH on continuous high temperature / short time sterilization of liquid. *American Institute of Chemical Eng. J*. **24**, 537-540. Fernandez P. S., Olio M. J., Rodrigo F. and Martinez A. (1996) Mathematical model for the combined effect of temperature and pH on the thermal resistance of *Bacillus stearothermophilus* and *Clostridium sporogenes* spores. Int. J. Food Microbiol. **32**, 225-233. Fernandez P. S., Olio M. J., Sanchez T. and Martinez A. (1994) Thermal resistance of *Bacillus stearothermophilus* spores heated in acidified mushroom extract. *J. Food Protect* **57**, 37-41. Jordan R. C. and Jacobs S. E. (1948) Studies in dynamics of desinfection. XIV. The variation of the concentration exponent for hydrogen and hydroxyl ions with the mortality level, using standard cultures of *Bact. coli* at 51°C. *J. Hyg. Cambridge* **46**, 289-295. Lopez M., Gonzales I., Condon S. and Bernardo A. (1996) Effect of pH heating medium on the thermal resistance of *Bacillus stearothermophilus* spores. Int. *J. Food Microbiol*. **28**, 405-410. Mafart P. and Leguérinel I. (1998) Modelling combined effects of temperature and pH on the heat resistance of spores by a linear-Bigelow equation. *J. Food. Sci*. In press. Palop A., Raso J., Pagan R., Condon S. and Sala F. J. (1996) Influence of pH on heat resistance of *Bacillus licheniformis* in buffer and homogenised foods. Int. *J. Food Microbiol*. **29**, 1-10.

286 Appendix 1: Reparameterization of Davey's model

287 Davey et al.(1978) described variation of death rates versus temperature 288 and pH by the following equation:

$$
289 \quad LnK = C_0 + \frac{C_1}{T} + C_2 pH + C_3 pH^2 \quad (1)
$$

290 where K is the death rate constant, T is the absolute temperature and C_0 , 291 C_1 , C_2 and C_3 are empirical coefficients. Let pH* be the pH value of maximal 292 thermal resistance of spores (in most cases $pH^*=7$)

293 In the situation of maximal thermal resistance,

$$
d(LnK) = C_2 + 2C_3 pH = 0
$$
 (2)

295

$$
pH^* = \frac{-C_2}{2C_3} \tag{3}
$$

so that Davey's model can be rewritten with only three parameters:
\n
$$
Ln K = C_0 + \frac{C_1}{T} + C_2 pH \left(1 - \frac{1}{2} \frac{pH}{pH} \right)
$$
\n(4)

299 For example, in the case of the heat treatment of *Clostridium botulinum* 300 spores in spaghetti tomato sauce, Davey's coefficient values were $C_2 = -2.3967$; 301 C₃=0.1965. In macaroni creole, C₂=- 2.6170 and C₃= 0.1871. Corresponding 302 pH* values are then 7.07 and 6.99 respectively. 303

304 Appendix 2: Reparameterization of Fernandez's linear model

$$
305
$$
 The following model was proposed:
306 $log K = a + bT + cpH$ (1)

10

D

with *K Ln* $=$ 307

$$
308 \t\t so equation (1) is equivalent to\n309 \t\t $\log D = \log(Ln10) - a - bT - cpH$ (2)
$$

$$
310 \quad \text{or} \quad
$$

310 or
\n311
$$
\log D = \log D^* - \frac{1}{z_T} (T - T^*) - \frac{1}{z_{pH}} (pH - pH^*)
$$
 (3)

312 By comparison between equation (2) and (3) new parameters D^* , z_T and z_{pH} can be identified versus Fernandez's parameters:
 $\log D^* = \log(Ln10) - a - bT^* - cpH^*$

$$
_{314}\quad \log D^{*}=\log(Ln10)-a-bT*-cpH^{*}
$$

$$
z_T = \frac{1}{b} \quad (5)
$$

$$
z_{pH} = \frac{-1}{c} \tag{6}
$$

317 For example, for *Bacillus stearothermophilus* heated in mushroom extract 318 acidified with glucono- δ -lactone, a=-12.0495; b=0.1156; c=0.2990. With 319 $T^*=121.1^{\circ}C$ and pH^{*}=7, new parameters are

- 320 D*=3.20 minutes
- 321 $z_T = 8.65$ °C
- 322 $z_{pH} = 3.34$
- 323
- 324
- 325

326 Table 1

328

-
-
-
-

figure 1

340

figure2

Three param eters m odel

358

- Legends of figures
-
- Figure 1 :
- Log D value of *Bacillus cereus* spores versus (T-T*).
- Key : \Box , log D at pH 6.5 ; Δ , log D at pH 5.5 ; \degree O, log D at 4.5.
-
- Figure 2 :
- Log D value of *Bacillus cereus* spores versus (pH-pH*)
- 376 Key : \Box , log D at 89° C; Δ , log D at 95° C; \degree O, log D at 101 $^{\circ}$ C.
-
- Figure 3 :
- Observed log D value compared to log D value calculated according to the three
- parameters model
-
- Figure 4 :
- Observed log D value compared to log D value calculated according to the four
- parameters model
-
-
- Legends of Table
-
- Table 1
- D values (minutes) of experimental design.

399 400