

## Cell-based measurements to assess physiological status of *Pseudo-nitzschia multiseries*, a toxic diatom

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1 Cell-based measurements to assess physiological status of *Pseudo-nitzschia multiseriata*, a  
2 toxic diatom

3

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## 11 **Abstract**

12 Diatoms of the genus *Pseudo-nitzschia* are potentially toxic microalgae, which blooms can  
13 trigger Amnesic Shellfish Poisoning. The purpose of this study was to test and adapt different  
14 probes and procedures to assess the physiological status of *Pseudo-nitzschia multiseriis* at the  
15 cell-level, using flow cytometry. To perform these analyses, probes and procedures were first  
16 optimized for concentration and incubation time. The percentage of dead *Pseudo-nitzschia*  
17 cells, the metabolic activity of live cells and their intracellular lipid content were then  
18 measured following a complete growth cycle. Additionally, chlorophyll autofluorescence and  
19 efficiency of photosynthesis (quantum yield) were also monitored. The concentration and  
20 viability of bacteria present in the medium were also assessed. Domoic acid (DA) was  
21 quantified as well. Just before the exponential phase, cells exhibited a high metabolic activity,  
22 but a low DA content. DA content per cell became most important at the beginning of the  
23 exponential phase, when lipid storage was high, which provided a metabolic energy source,  
24 and when they were surrounded with a high number of bacteria (high bacteria/*P. multiseriis*  
25 ratio). These physiological measurements tended to decrease during exponential phase and  
26 until stationary phase, where *P. multiseriis* cells did not content any DA nor stored any lipids  
27 and started to die.

28

29 Keywords: flow cytometry; cell physiology; domoic acid; *Pseudo-nitzschia multiseriis*;  
30 bacteria; fluorescent probes

## 31 1. Introduction

32 *Pseudo-nitzschia* is a potentially toxic diatom genus with a worldwide distribution.  
33 Some species are able to produce domoic acid (DA), an amnesic shellfish toxin leading to  
34 food poisoning (Sierra-Beltrán et al., 1998) with a few cases of mortality to humans (Wright  
35 et al., 1989), plus hundreds of sea bird (Sierra-Beltrán et al., 1997, Work et al., 1993) or  
36 marine mammal mortalities (Scholin et al., 2000, Fire et al., 2009, de la Riva et al., 2009).  
37 These poisonings often occurred following a bloom of *Pseudo-nitzschia* spp. The reasons  
38 why these blooms occurred are poorly known. Some studies tried to create models to predict  
39 their occurrence (Anderson et al., 2009, Lane et al., 2009), but the determinism of each  
40 bloom seems different. Although factors enhancing or decreasing *Pseudo-nitzschia* cell  
41 toxicity have been intensively studied, they still remain unclear. The study of *Pseudo-*  
42 *nitzschia* spp. physiology may help to understand why a bloom appears and becomes toxic.  
43 Tools to assess the physiological status of microalgae are still fairly scarce. Photosynthetic  
44 capacities of *Pseudo-nitzschia* spp. have been studied under different conditions (Ilyash et al.,  
45 2007, El-Sabaawi and Harrison, 2006) but do not provide enough information to assess cell  
46 physiological status. Besides photosynthetic parameters and chlorophyll content, others  
47 parameters have sometimes been studied in diatoms, e.g. silicification (Leblanc et al., 2005,  
48 Kroger and Poulsen, 2008) or carbohydrate levels (De Philippis et al., 2002, Magaletti et al.,  
49 2004), but these are also insufficient to characterize the physiological processes occurring  
50 inside the cell. It is therefore important to develop and then simultaneously measure several  
51 different physiological parameters that may help to better understand the factors or status  
52 associated with toxin production.

53

54 Assessment of cell physiology using fluorescent probes is a well-known subject in  
55 medicine (Greenspan et al., 1985, Knot et al., 2005). Among the numerous fluorescent probes

56 available to assess cell physiology, some can be adapted to cultures of unicellular organisms.  
57 They allow measurements of different physiological parameters such as metabolic activity  
58 (with fluorescein diacetate, FDA), intracellular lipid content (Nile Red and BODIPY), total  
59 DNA (SYBR Green) or mortality (SYTOX Green). Some of these probes have been used in  
60 microalgal studies for several years but are often limited to microscopic observations or  
61 spectrofluorimetric methods (Dempster and Sommerfeld, 1998, Okochi et al., 1999). The  
62 latter allow measurement of an entire population, but differences between cells can not be  
63 observed. Microscopic observations allow cell-by-cell analyses but are time consuming and  
64 fluorescence quantification is difficult. On the other hand, flow cytometry (FCM) allows a  
65 rapid analysis of the morphological and fluorescence characteristics of unicellular organisms  
66 or individual cells. Even though FCM has a long history of routine use in medical analyses,  
67 the first experiments to use FCM on microalgae were run only about thirty years ago (Olson  
68 et al., 1983, Yentsch et al., 1983) and the approach still remains only a minor component for  
69 measuring the physiology of phytoplankton. Some probes have already been tested on  
70 microalgae using FCM, such as FDA (Dorsey et al., 1989, Brookes et al., 2000, Jochem,  
71 1999), SYTOX Green (Veldhuis et al., 1997) or SYBR Green (Marie et al., 1997). Each of  
72 these probes provides new insights for understanding how cells react under different  
73 conditions, e.g. dark adaptation (Jochem, 1999), but they have never been applied  
74 simultaneously to assess physiological status in a more comprehensive manner.

75

76 This study aims to assess the physiological status of *Pseudo-nitzschia multiseriis*  
77 using a set of cell-based measurements. To reach this objective, different measurements were  
78 developed and adapted to this species (i) to better understand its physiology under culture  
79 conditions and (ii) to seek the relationship between the production of DA and cell  
80 physiological status. The morpho-functional characteristics of *P. multiseriis* cells were

81 assessed by FCM, using different fluorescent probes (FDA, BODIPY 493/503, Nile Red,  
82 SYTOX Green, SYBR Green and propidium iodide) and the measurement of chlorophyll  
83 autofluorescence. Quantum yield (QY), which is a measurement of the efficiency of  
84 photosynthesis, was measured using a pulse amplitude modulated (PAM) fluorometer.  
85 Dissolved and particulate DA were measured on each culture using an ELISA assay. DA is a  
86 secondary metabolite, thus supposed to be produced when cells have more energy than  
87 necessary for the primary metabolism. Thus primary metabolism was assessed using FDA  
88 and esterases activity. The availability of energy was assessed by measuring storage lipids, as  
89 extra-energy is stored by microalgae under lipid form. The concentration of bacteria may  
90 influence DA production by *P. multiseriis*, as they are known to enhance DA production  
91 (Bates et al., 1995). Chlorophyll and QY measurements allow knowing if the culture is  
92 healthy and were completed by the measure of dead cells percentage.

## 93 2. Materials and methods

### 94 2.1. Cultures

95 Strain CCAP 1061/32 of *Pseudo-nitzschia multiseriata* (isolated in 2007 in England)  
96 was used for the experiments,. Cultures (n=6) were grown in sterilized f/2 medium (Guillard  
97 and Hargraves, 1993) at 15.6°C ( $\pm 0.2^\circ\text{C}$ ), and  $131 \pm 16 \mu\text{mol photons m}^{-2} \text{ s}^{-1}$  (light:dark  
98 photoperiod of 12:12 h). Seawater used for f/2 medium was first filtered at 0.22 $\mu\text{m}$ , to  
99 eliminate any remaining bacteria (which was confirmed by flow cytometric measurements, as  
100 described below) and then autoclaved. Cultures were xenic and grown without antibiotics.  
101 Before each sampling, cultures were homogenized by gentle manual stirring. Almost all the  
102 cells were present as single cells in our cultures; sometimes cells were forming 2 cells chains.  
103 For the flow cytometry analysis, they were all considered as single cells.

104

### 105 2.2. Physiological measurements

106 Measurements were made with a FACScalibur flow cytometer (BD Biosciences, San  
107 Jose, CA USA), using an argon blue laser (488 nm). Three fluorescence signals can be  
108 detected by the flow cytometer: FL1 (green, 530 nm), FL2 (orange, 585 nm) and FL3 (red,  
109 670 nm). Red fluorescence is linearly linked to the chlorophyll content of the cells and was  
110 used as a discriminating characteristic to detect the microalgae (Fig. 1). Bacteria were  
111 detected on the FL1 channel (Fig. 2), with different settings to those used for microalgae  
112 analysis. Cell counts were estimated from the flow-rate measurement of the flow cytometer  
113 (Marie et al., 1999) as all samples were run for 45 s. The flow rate from the FCM was  
114 controlled every two days. Forward Scatter (FSC, light scattered less than 10 degrees) and  
115 Side Scatter (SSC, light scattered at a 90 degree angle) were also measured. FSC is  
116 commonly related to cell size and SSC to cell complexity. The same instrument settings were  
117 used for the entire duration of the experiment to allow comparison between days.

118

119           *Bacteria.* Quantification of free-living bacteria in the *P. multiseriis* culture and the  
120 percentage of dead bacteria in the culture were assessed by adding SYBR Green I (Molecular  
121 probes, Invitrogen, Eugene, Oregon, USA) at a final concentration of 1/10000 of the  
122 commercial solution, and propidium iodide (PI, Sigma, St. Louis, MO, USA) at 10  $\mu\text{g ml}^{-1}$  to  
123 each sample. During analyses, aggregates of bacteria were taken into account with correction  
124 according to aggregate size (Fig. 2). Bacterial counts were estimated as described for *Pseudo-*  
125 *nitzschia* cells, using FL1 as a discriminating characteristic (due to SYBR Green fluorescence  
126 staining).

127

128           *Mortality.* To assess *Pseudo-nitzschia* cell mortality, we used a cell membrane-  
129 impermeable dye, SYTOX Green (Molecular probes, Invitrogen, Eugene, Oregon, USA)  
130 prepared at a working solution of 5  $\mu\text{M}$ . A mix of live/dead cells was prepared to confirm  
131 that SYTOX Green stained only dead cells (Veldhuis et al., 2001) and to calibrate the  
132 measurement. Cells from a dead culture (killed by heating for 15 min at 100°C) were mixed  
133 with those from a live culture to give a range of 0 to 100% dead cells (increments of 10%)  
134 and stained with SYTOX Green at 0.1  $\mu\text{M}$  (final concentration) for 30 min. The percentage  
135 of measured dead cells (those stained with SYTOX Green) was then compared to the  
136 theoretical percentage of dead cells present in the mixture.

137

138           *Metabolic activity.* To assess metabolic activity, esterase activity was measured using  
139 fluorescein diacetate (FDA, Molecular probes, Invitrogen, Eugene, Oregon, USA). FDA is a  
140 probe that is cleaved by esterases inside the cells, resulting in fluorescein accumulation over  
141 time (Jochem, 1999). A 5  $\text{mg ml}^{-1}$  stock solution of FDA was prepared by diluting the  
142 commercial powder in DMSO. A fresh 300  $\mu\text{M}$  working solution was prepared before each



143 experiment by adding stock solution directly into distilled water cooled on ice. The working  
144 solution was kept in darkness and on ice during the experiment and was agitated to prevent  
145 the formation of aggregates.

146

147 *Lipids.* To assess the intracellular lipid content, two probes were tested on *P.*  
148 *multiseriis*. A 10 mM stock solution of BODIPY 493/503 (4,4-difluoro-1,3,5,7,8-  
149 pentamethyl-4-bora-3a,4a-diaza-s-indacene, Molecular probes, Invitrogen, Eugene, Oregon,  
150 USA) was made by diluting the commercial powder in DMSO. A 1 mM working solution  
151 was then prepared by a 10-fold dilution of the stock solution in distilled water. A 1 mg ml<sup>-1</sup>  
152 stock solution of Nile Red (NR, Sigma, St. Louis, MO, USA) was prepared by diluting 100-  
153 fold the commercial powder in acetone and then 10-fold in distilled water to obtain a working  
154 solution of 0.1 mg ml<sup>-1</sup>.

155

156 Each of these measurements had to be optimized for *P. multiseriis*. Thus, final probe  
157 concentrations and incubation times were chosen following two rules: (i) the concentration  
158 had to be as low as possible to avoid toxic effects of the probe itself (and of the DMSO  
159 contained in stock solutions of the probes) and (ii) the staining had to be homogeneous (all of  
160 the cells had to be stained or only the dead cells for SYTOX Green), relatively stable over  
161 time and reproducible between analytical replicates. For each probe, fluorescence  
162 measurements were performed every 5 min during 1 h on the FL1 (or FL2 for NR) channel of  
163 the flow cytometer. Concentrations of 0.5, 1.0, 2.5 and 5.0 µg ml<sup>-1</sup> were tested for NR; 1.0,  
164 2.5, 5.0 and 10 µM for BODIPY; 0.75, 1.50 and 3.00 µM for FDA and 0.025, 0.05, 0.1 and  
165 0.2 µM for SYTOX Green.

166

167 Quantum yield (QY), a measurement of the efficiency of photosynthesis, was  
168 measured using an AquaPen-C AP-C 100 (Photo Systems Instruments, Czech Republic) pulse  
169 amplitude modulated (PAM) fluorometer.  $QY = (F_m - F_0) / F_m$ , where  $F_0$  and  $F_m$  are the  
170 minimum and maximum fluorescence of cells, respectively, after 30 min of dark adaptation.  
171 To ensure that there was no background fluorescence, *P. multiseriis* supernatant and f/2  
172 medium were used as blanks.

173

174 Domoic acid (DA) was quantified using the ASP ELISA kit (Biosense Laboratories,  
175 Bergen, Norway), according to the manufacturer's protocol. Cultures (cells and supernatant)  
176 were sonicated and filtered at 0.22  $\mu\text{m}$  to measure total DA. Supernatant (culture filtered at  
177 0.22  $\mu\text{m}$ ) was used to measure dissolved DA. Intracellular DA was measured by subtracting  
178 dissolved DA to total DA.

179

### 180 2.3. Monitoring the physiology of *P. multiseriis* over a growth cycle

181 Six *P. multiseriis* cultures of the same strain were sampled every day from day 4 to  
182 21. The following were assessed on each sampling day: *P. multiseriis* morphology,  
183 concentration and mortality, bacterial concentration and mortality, total and dissolved DA  
184 concentrations, quantum yield, chlorophyll fluorescence, intracellular lipid content and  
185 metabolic activity. Growth rate was measured during exponential phase following the  
186 formula:  $\mu \text{ (d}^{-1}\text{)} = \ln(N1/N0) / \Delta t$  (in days). Fluorescence measurements were performed using  
187 the optimal concentrations obtained in the previous experiments: 0.1  $\mu\text{M}$  SYTOX Green, 3  
188  $\mu\text{M}$  FDA, 1  $\mu\text{g ml}^{-1}$  Nile Red and 10  $\mu\text{M}$  BODIPY. FDA measurements were performed  
189 precisely after 6 min of incubation, and SYTOX Green and BODIPY measurements after 30  
190 min. Bacteria were stained with SYBR Green and PI for 15 minutes.

191

192 2.4. *Statistics*

193           Results were analyzed statistically with simple regressions, One-Way ANOVA with  
194 time as the main factor, as well as with principal component analysis (PCA) followed by a  
195 factorial plan. For all statistical results, a probability of  $p < 0.05$  was considered significant.  
196 Statistical analyses were performed using StatGraphics Plus (Manugistics, Inc, Rockville,  
197 MD, USA).

198

### 199 3. Results

200

#### 201 3.1. Optimization of probe concentration

202 *Mortality.* A 0.1  $\mu\text{M}$  final concentration of SYTOX Green allowed a good distinction  
203 between dead and live cells (Fig. 1). Incubation time was optimal at 30 min. The best  
204 correlation ( $y=0.95x$ ,  $R^2 = 0.99$ ,  $P<0.01$ ) between the measured and theoretical percentages of  
205 dead cells in the mixtures of dead/live *P. multiseriis* was established at a SYTOX Green  
206 concentration of 0.1  $\mu\text{M}$ , which was therefore applied for the further analyses.

207

208 *Bacteria.* Free-living bacteria are able to form aggregates that can be distinguished  
209 after SYBR Green staining (Fig. 2A). Each aggregates exhibited a fluorescence that was  
210 equivalent to the fluorescence of one bacteria x number of bacteria in the aggregate. The  
211 number of total free-living bacteria can thus be deduced and measurements of FSC and SSC  
212 can be done for each aggregate size (Fig. 2B).

213

214 *Metabolic activity.* Concentrations of FDA lower than 3.0  $\mu\text{M}$  exhibited a low  
215 fluorescence, indicating that there was little accumulation of the probe (data not shown). At  
216 3.0  $\mu\text{M}$ , fluorescein accumulated within the cells (Fig. 3I), in a linear manner during the 15  
217 first min ( $y=35.9x+133.1$ ,  $R^2 = 0.9964$ ,  $p<0.001$ ), and then reached a plateau (Fig. 4). For the  
218 further analyses, fluorescein accumulation was measured after 6 min of staining within the  
219 linear part of the curve.

220

221 *Lipids.* A final concentration of 10  $\mu\text{M}$  BODIPY 493/503 and 1  $\mu\text{g ml}^{-1}$  of Nile Red  
222 (NR) allowed the best staining of all cells (one distinct population of cells, not a diffuse cloud

223 of cells, can be seen on the cytograms, data not shown). After 30 min, all cells were well  
224 stained and the fluorescence was stable (Fig. 3B, C, E, F).

225

### 226 3.2. Monitoring of morpho-functional characteristics during *P. multiseri* growth

227 The exponential growth phase of *P. multiseri* started after a 7-d lag phase, giving a  
228 growth rate of  $0.24 \pm 0.01 \text{ d}^{-1}$ , and then the stationary phase was reached after day 17 (Fig. 5).  
229 A maximal concentration of  $\sim 8 \times 10^4 \text{ cells ml}^{-1}$  was observed at days 17 and 19, after which  
230 the cell concentration rapidly declined.

231

232 Bacteria within the *P. multiseri* culture started their exponential growth phase on  
233 day 7 and were still growing steadily until the end of the experiment (Table 1), exhibiting a  
234 growth rate of  $0.07 \pm 0.01 \text{ d}^{-1}$ . The bacteria/*P. multiseri* cell ratio decreased during the  
235 exponential phase of *P. multiseri*, remained stable between days 14 to 20, and then  
236 increased again on the last day of the experiment, when *P. multiseri* numbers declined (Fig.  
237 5, Table 1). Proportions of bacteria in aggregates of one, two or more cells did not change  
238 with growth phases. The percentage of dead bacteria decreased between days 4 and 12 (from  
239  $5.8\% \pm 0.6$  to  $2.0\% \pm 0.2$ ) and then remained stable between 1.9 and 2.5% until day 21 (Table  
240 1). Values of FSC and SSC for the bacterial community (free-living bacteria that were not  
241 forming aggregates) decreased steadily during the course of *P. multiseri* culture (Table 1).

242

243 The percentage of dead *P. multiseri* cells averaged 28% throughout the entire  
244 experiment (Table 1) and decreased from 30.3% on day 12 to 18.9% at day 16, after which it  
245 increased to 43.8% on day 20 (stationary phase). FSC values for *P. multiseri* continuously  
246 decreased during the experiment, almost linearly with culture age ( $R^2=0.76$ ,  $p<0.01$ ). SSC

247 values decreased until day 13 (mid-exponential phase) and became stable between day 13 and  
248 the end of the experiment (Table 1).

249

250 Total DA in the *P. multiseriis* culture, expressed as  $\text{pg ml}^{-1}$ , increased steadily from  
251 day 7 ( $200 \pm 21 \text{ pg ml}^{-1}$ ) until day 14 ( $798 \pm 164 \text{ pg ml}^{-1}$ ) during exponential growth. Total DA  
252 in the culture then decreased sharply, reaching a concentration below  $100 \text{ pg ml}^{-1}$  on day 21.  
253 Total DA content was highest on days 13 and 14, during the mid-exponential phase, and  
254 decreased steadily after day 14, when it reached late-exponential phase and stationary phase  
255 (Fig. 5). The amount of dissolved DA was low and remained constant throughout the culture,  
256 from  $41.3 (\pm 2.9) \text{ pg ml}^{-1}$  on day 6 to  $103.0 (\pm 7.3) \text{ pg ml}^{-1}$  on day 12, representing 11 to 40 %  
257 of total DA.

258

259 FL3 values (related to chlorophyll content) were measured on live cells, discriminated  
260 from dead cells using SYTOX Green staining. FL3 values sharply decreased from day 4 to 6,  
261 remained stable between days 6 ( $590 \pm 12$ ) and 9 ( $593 \pm 10$ ), and then slightly decreased from  
262 day 9 to 20 ( $506 \pm 12$ ), stationary phase, Fig. 6). Quantum yield (QY) values increased  
263 between days 5 ( $0.46 \pm 0.01$ ) and 8 ( $0.59 \pm 0.01$ ), became relatively stable until day 14  
264 ( $0.62 \pm 0.00$ ), and then decreased in mid-exponential phase after day 14 (Fig. 6). Day 11  
265 exhibited a significant decrease of both FL3 and QY. Supernatant and media did not exhibit  
266 QY values or were below the detection threshold of the fluorometer.

267

268 The metabolic activity of the *P. multiseriis* cells, as measured with the FDA assay  
269 after 6 min of incubation, increased rapidly from day 6 to 7 and then just as rapidly decreased  
270 after day 7, to values just below the initial level, on day 9 (Fig. 7). The percentage of stained  
271 cells after 6 min of incubation increased between day 8 ( $72.5\% \pm 1.6$ ) and 16 ( $87.9\% \pm 1.0$ )

272 and decreased on day 20 ( $74.7\% \pm 5.4$ ). The percentages of live cells, as measured with the  
273 SYTOX Green and FDA assays, were significantly correlated, even though the correlation  
274 remained quite weak ( $R^2=0.67$   $p<0.001$ , Fig. 7).

275

276 The amount of intracellular lipids, interpreted from BODIPY fluorescence, increased  
277 between days 4 and 6, decreased from days 6 to 14 (during the exponential phase), stayed  
278 stable until day 16, and finally increased during the stationary phase (Fig. 8). NR  
279 fluorescence, the traditional indicator of lipid content, decreased between days 9 and 20, with  
280 one higher fluorescence value on day 11 (Fig. 8).

281

282 PCA showed that DA content of cells (total DA) had coordinates really close to those  
283 of Nile Red, SSC and bacteria/*Pseudo-nitzschia* ratio, knowing that components 1 and 2  
284 explained 74 % of the variability (Fig. 9). FSC and BODIPY uptake were also closely  
285 correlated with these previous parameters, indicating that increased DA production was  
286 associated with a higher intracellular lipid content. A factorial plan (Fig. 10) was developed  
287 from the previous PCA, plotting the age of the *P. multiseriis* culture, in exponential and  
288 stationary phases from day 9 to 20. Days follow a consistent trend, from high component 1  
289 and low component 2 (i.e. high lipid concentration, high DA content, high cell/bacteria ratio,  
290 low esterases activity, ....) , towards lower component 1 and higher component 2 (i.e. high  
291 esterases activity, low DA content and low lipid concentration, ...). Day 20 was the only days  
292 which did not follow this trend, on the extremities of the factorial plan (extremely low  
293 components 1 and 2, i.e. driven mainly by the high *P. multiseriis* mortality and high number  
294 of bacteria).

#### 295 4. Discussion

296 The first aim of this study was to test and optimize several methods and probes to  
297 assess *Pseudo-nitzschia* physiological status. The percentage of cell mortality in the cultures  
298 was determined using SYTOX Green, which only penetrates cells that have lost their  
299 membrane integrity, and are thus considered as dead cells (Veldhuis et al., 1997). A final  
300 concentration of 0.1  $\mu\text{M}$  was optimal for staining *P. multiseriis* dead cells and is in good  
301 agreement with those found in the literature for other phytoplankton species (Veldhuis et al.,  
302 2001, Binet and Stauber, 2006, Ribalet et al., 2007, Miller-Morey and Van Dolah, 2004,  
303 Lawrence et al., 2006).

304

305 Fluorescein diacetate (FDA) has previously been used to measure metabolic activity  
306 (Jochem, 1999, Regel et al., 2002, Brookes et al., 2000) as well as viability of microalgae  
307 (Lawrence et al., 2006, Dorsey et al., 1989, Jansen and Bathmann, 2007). It penetrates the  
308 cells passively and once within the cell is hydrolyzed by non-specific esterases into  
309 fluorescein and two acetate molecules. The more metabolically active the cells are, the more  
310 esterases they produce, resulting in a greater amount of fluorescein accumulation within the  
311 cells. The probe will not be cleaved within dead cells, as esterases are inactive. Moreover, if  
312 the probe is hydrolyzed by any remaining esterases, the fluorescein will leak out of the cells,  
313 as the membranes are permeable. Thus, unstained cells are considered as dead cells. In the  
314 literature, measurement of fluorescein released from FDA inside the cells most often occurs  
315 between 5 and 20 min of incubation (Jochem, 1999, Regel et al., 2002, Dorsey et al., 1989,  
316 Jamers et al., 2009). FDA was only accumulated linearly during the first 15 to 20 min, as  
317 previously observed by Gilbert et al. (1992). Accordingly, based on our results and supported  
318 by the above publications, measurements were performed after 6 min of incubation. A final  
319 concentration of 3  $\mu\text{M}$  was optimal for this assay and is consistent with some publications



320 (Dorsey et al., 1989, Gilbert et al., 1992) but lower than others (Regel et al., 2002, Jamers et  
321 al., 2009). Higher concentrations of FDA were not tested, as 3  $\mu\text{M}$  provided satisfactory  
322 staining and higher concentrations of FDA and DMSO may become toxic to the cells.

323

324 BODIPY 493/503 and Nile Red (NR) were tested to localize and quantify intracellular  
325 lipids in *P. multiseriis* cells. NR has been used traditionally to stain lipids of microalgae  
326 (Cooksey et al., 1987), whereas this is the first time that BODIPY 493/503 has been used to  
327 study microalgal lipids. NR fluorescence of microalgal lipids, measured by FCM, has been  
328 shown to be linearly correlated to the lipid content of cells (de la Jara et al., 2003). Lipids of  
329 *P. multiseriis*, revealed by BODIPY and NR, were observed to form vacuoles inside the cells  
330 (Fig. 3), and NR gave a lower fluorescence intensity than BODIPY. These vacuoles are likely  
331 to contain reserve lipids, as BODIPY and NR are reported to stain neutral lipids (Gocze and  
332 Freeman, 1994). Such vacuoles have previously been described within microalgae (Eltgroth  
333 et al., 2005, Liu and Lin, 2001, Remias et al., 2009, Cooper et al., 2010), although lipid-  
334 staining BODIPY and NR did not reveal a specific distribution of these vesicles. Both  
335 BODIPY and NR were used to quantify intracellular lipid contents by FCM, in the green  
336 (FL1) and orange (FL2) channels respectively. In the present study, NR was used at a final  
337 concentration of 1  $\mu\text{g ml}^{-1}$ , which is the same as used in previous studies on microalgae  
338 (Chen et al., 2009, Chen et al., 2010, Liu et al., 2008, Huang et al., 2009, McGinnis et al.,  
339 1997). BODIPY was used at a final concentration of 10  $\mu\text{M}$ . This concentration allowed the  
340 detection of subtle variations in the intracellular lipid content of *P. multiseriis* grown, for  
341 example, in culture media with or without nitrate (data not shown), whereas lower  
342 concentrations did not. Higher concentrations were not tested, as 10  $\mu\text{M}$  provided satisfying  
343 staining and higher concentrations of BODIPY and DMSO may become toxic to the cells.  
344 The concentration used was 100 times higher than the one used for fungus (Saito et al., 2004)

345 but in agreement with those on human muscle (Wolins et al., 2001) and lower than the one  
346 used on amoeba (Kosta et al., 2004).

347

348 The development of these methods allowed the physiological status of *P. multiseriis*  
349 cells to be monitored over a complete growth cycle. The lag phase of *P. multiseriis* lasted 7  
350 days, which is long compared to other studies on the same species, but not the same strain  
351 (Thessen et al., 2009, Lundholm et al., 2004, Kudela et al., 2003, Kotaki et al., 1999, Bates et  
352 al., 2000). The *P. multiseriis* growth rate ( $0.24 \pm 0.01 \text{ d}^{-1}$ ) was lower than those previously  
353 reported in the literature (Thessen et al., 2009, Lundholm et al., 2004, Kudela et al., 2003,  
354 Kotaki et al., 1999, Bates et al., 2000). This might be explained by the age of the isolate  
355 (isolated in 2007, more than 2 years ago) and the short cell length ( $\sim 20 \mu\text{m}$ ); Amato et al.  
356 (2005) reported a slight decrease in the growth rate of *P. delicatissima* with a decrease in  
357 apical cell length. Culture conditions were the same or close to studies using *Pseudo-*  
358 *nitzschia* cultures (media, irradiance and temperature) and thus could not explain differences  
359 in growth rates.

360

361 FSC and SSC values of *P. multiseriis* decreased during the entire experiment, by 17%  
362 and 22%, respectively. FSC and SSC result from the diffraction of the laser by the cell  
363 surface. Their decrease in *P. multiseriis* may be related to changes in external morphology,  
364 cell size and internal cell complexity. During growth, cells undergo asexual reproduction and  
365 thus decrease in cell length. FSC and SSC values were, however, similar to values measured  
366 over the last year (data not shown) at both the beginning (during the lag phase) and end of  
367 experiments. This indicates that FSC and SSC values changed very little over the last year,  
368 possibly because this strain isolated in 2007 was already quite old. Inoculation of *P.*  
369 *multiseriis* into a new medium resulted in a return to high FSC and SSC values. Because

370 diatoms cannot increase their cell size, the changes in FSC and SSC values are more likely  
371 related to both surface membrane and cytoplasmic modifications than cell size modifications,  
372 thus modifying the diffraction of the laser. This hypothesis is based on the correlation  
373 between SSC and both BODIPY and NR fluorescences ( $R^2 = 0.77$  and  $0.64$ , respectively, at  
374  $p < 0.01$ ). It may possible that when cells had a lot of lipid vesicles within their cytoplasm, this  
375 increased cell complexity was reflected by the changes in FSC and SSC values.

376

377         Bacterial community counts and morphological changes within *Pseudo-nitzschia*  
378 cultures were estimated for the first time by FCM. In this microalgal culture, the growth rate  
379 of the bacteria was  $0.07 \text{ d}^{-1}$ , which remained constant over the course of the experiment; the  
380 bacteria did not reach stationary phase during the 20 days of the experiment. This growth rate  
381 is in the lower range of bacteria grown in adapted culture media, that can grow from  $0.01 \text{ h}^{-1}$   
382 (Kemp et al., 1993) to  $1.5 \text{ h}^{-1}$  (Makino et al., 2003). These differences may be due to the  
383 competition with *P. multiseriis* for some nutrients or the fact that they may not have all the  
384 nutrients they need and that are usually added in agar plates. The highest bacteria/*P.*  
385 *multiseriis* ratios were measured during the lag phase (day 4 to 7) and at the beginning of the  
386 exponential phase (day 7 and 8). Bacteria measured are the free-living bacteria contained in  
387 the medium; however, some bacteria can also be attached directly to *P. multiseriis* cells  
388 (Kaczmarska et al., 2005), these attached bacteria were not taken into account (their signal  
389 was confounded within these of *P. multiseriis*). The decrease in the number of bacteria per *P.*  
390 *multiseriis* cell during the exponential phase of *P. multiseriis* (from 922 to 180) is explained  
391 by a faster growth rate of *P. multiseriis* compared to bacteria. The increase in the bacteria/*P.*  
392 *multiseriis* ratio during the senescent phase of *P. multiseriis* may be a result of bacteria  
393 taking advantage of organic materials released from dead *P. multiseriis* cells (Kaczmarska et  
394 al., 2005). Stewart et al. (1997) found between 7 and 10 bacteria per *P. multiseriis* cell,

395 which is about 20 to 80 times lower than our values. This difference may be explained by (i)  
396 a high residual percentage of dead *P. multiseriis* cells present during the entire experiment, or  
397 (ii) the age of our isolate, which provided sufficient time (two years) for the bacterial  
398 community to adapt to the culture conditions of *P. multiseriis*. Differences found in bacterial  
399 communities over time in culture, for non-toxic *Pseudo-nitzschia pungens* support this  
400 possibility (Sapp et al., 2007), but Wrabel and Rocap (2007) found no shifts in bacterial  
401 assemblages in a *Pseudo-nitzschia* culture over its initial nine months (Wrabel and Rocap,  
402 2007). Nevertheless, shift in the bacterial community may appear after 9 months in culture.  
403 FSC and SSC values of the bacterial community decreased during the experiment. These  
404 values are related to size and complexity of bacterial cells. This may reflect a shift in species  
405 composition of the bacterial community to smaller bacteria or a decrease in bacterial cell size.  
406 Between 1.9 and 5.8% of the bacteria in our cultures were dead, with the highest percentage  
407 at day 4. The percentage of dead bacteria remained quite low (1.9-2.7%) until the end of the  
408 experiment, as they were still in exponential phase.

409

410 Values of FL3 (related to the chlorophyll content) decreased slightly during the entire  
411 experiment, with a greatest decrease between days 4 and 6. The chlorophyll content of *P.*  
412 *multiseriis* decreased only slightly during the exponential phase. Nevertheless, cells with  
413 more chlorophyll may not necessarily have the most efficient photosynthesis. Indeed, QY, a  
414 measure of the efficiency of photosynthesis, was not well correlated to FL3 values, as QY  
415 decreased during the stationary phase when FL3 remained high. QY increased at the  
416 beginning of the exponential phase and remained high during the remaining exponential  
417 phase, with cells having an efficient photosynthesis with a lot of energy produced. Such an  
418 increase of QY during the exponential phase has been shown for other microalgal species,  
419 e.g. *Symbiodinium* sp. (Rodriguez-Roman and Iglesias-Prieto, 2005), and is currently used as

420 a measure of algal culture health. As the QY value is not affected by the percentage of dead  
421 cells in the cultures (Franklin et al., 2009), it can be speculated that at the end of the  
422 stationary phase, live *P. multiseriis* cells still contained high amounts of chlorophyll, but with  
423 a poor photosynthetic efficiency.

424

425         During the entire experiment, the percentage of dead *P. multiseriis* cells was  
426 relatively high, ranging from 19% to 54%. Nevertheless, our cultures reached a maximum  
427 cell concentration of  $8 \times 10^4$  cells ml<sup>-1</sup>, which is consistent with some previous studies  
428 (Mengelt and Prézelin, 2002, Bates and Richard, 1996, Lewis et al., 1993, Kotaki et al., 1999)  
429 but lower than the results of the majority of the studies (Bates and Richard, 1996, Kotaki et  
430 al., 1999, Mengelt and Prézelin, 2002), suggesting that our cultures were not in good health,  
431 which also explains the low growth rate and the high percentage of dead cells. Generally, in  
432 healthy and young cultures of *Pseudo-nitzschia* sp., the percentage of dead cells has been  
433 described under 5% (Mengelt and Prézelin, 2002). The increase in dead cells at the end of the  
434 experiment may be due to the limitations in nutrients and associated with the beginning of the  
435 stationary phase. Such a consistently high percentage of dead cells in the culture may be  
436 explained by the age of the isolate. The percentage of dead cells assessed with FDA was  
437 significantly but not perfectly correlated ( $R^2 = 0.67$ ,  $p < 0.01$ ) to those obtained with SYTOX  
438 Green and appeared slightly lower than when measured with SYTOX Green. Cells can have a  
439 compromised cell membrane, and be considered as dead when assessed with SYTOX Green,  
440 but they may still have active esterases. These false-positive cells (dead but stained with  
441 FDA) have been shown to represent 1.6% of total cells of *Chlamydomonas reinhardtii*  
442 (Jamers et al., 2009). Such differences between SYTOX Green and FDA have also been  
443 previously observed in *Heterosigma akashiwo* (Lawrence et al., 2006). Using these two  
444 probes not only provides the percentage of dead versus live cells but also provides an

445 indication of the way cells are dying. In our cultures, cells most likely died by loss of  
446 membrane integrity prior to inactivation of esterases, which was also observed by Lawrence  
447 et al. (2006) in cultures of *H. akashiwo*. Thus, SYTOX Green and FDA provide useful  
448 information and could both be used in physiological measurements.

449

450 Lipid-related fluorescence assessed with BODIPY was high during the lag phase,  
451 indicating that the cells contained energy stored as neutral lipids. BODIPY fluorescence  
452 decreased during the entire exponential phase, suggesting that cells were using these stored  
453 lipids to grow, in addition to the energy produced by photosynthesis. Cells stopped growing  
454 at the stationary phase, and energy was once again stored as lipids, as evidenced by the  
455 increase in BODIPY fluorescence. Although no data are available between days 4 and 6, NR  
456 fluorescence decreased during the remainder of the experiment, with the exception of a high  
457 value on day 11. There was a weak correlation between BODIPY and NR fluorescence  
458 during the exponential phase (between days 7 and 18;  $R^2 = 0.65$ ,  $p < 0.01$ ). During the  
459 stationary phase, however, BODIPY fluorescence is higher than that of NR, which confirms  
460 that these two probes may not actually stain the same compounds during that period. This  
461 emphasises the importance of using both lipid probes. These differences may be explained by  
462 the chemical properties of the two probes. BODIPY 493/503 stains intracellular lipids more  
463 effectively than NR, with a higher sensitivity and lower background (Kacmar et al., 2006).  
464 BODIPY 493/503 also stains intracellular lipid droplets more specifically than does NR  
465 (Gocze and Freeman, 1994). NR is an uncharged hydrophobic molecule whose fluorescence  
466 is strongly influenced by the polarity of its environment. As well as lipids, NR interacts with  
467 many, but not all, native proteins (Sackett and Wolff, 1987) and can undergo changes in  
468 fluorescence intensity when it binds to certain proteins (Brown et al., 1995). The fact that NR  
469 binds proteins may explain its lower sensitivity to small variations in lipid content, as

470 measured by BODIPY. This is especially evident during the stationary phase, when  
471 differences in lipid staining were observed between the two probes. Thus, the combined use  
472 of BODIPY and NR probes is of interest as they may reflect different physiological changes.

473

474 The maximum of total DA per cell was observed on days 9 and 10, in early  
475 exponential phase, and it decreased during the remainder of the exponential phase, and the  
476 stationary phase. The same pattern of DA production has been observed for *Pseudo-nitzschia*  
477 *calliantha* (Besiktepe et al., 2008) and *Pseudo-nitzschia pseudodelicatissima* (Pan et al.,  
478 2001), where the maximum DA production was observed during the early exponential phase.  
479 All studies of *P. multiseriis*, however, have found a maximum DA production during the  
480 stationary phase (Kotaki et al., 1999, Bates et al., 2000, Lewis et al., 1993, Osada and  
481 Stewart, 1997). Maybe old cultures of *P. multiseriis* exhibit a shift of DA production from  
482 stationary phase to early exponential phase, which is difficult to prove, as one strain has  
483 never been studied throughout its lifetime in laboratory. Moreover, strains exhibiting DA  
484 production during early exponential phase seem to have a lower DA content per cell  
485 (Besiktepe et al., 2008, Pan et al., 2001). In our study, total cellular DA varied between 0 and  
486  $192 \text{ fg cell}^{-1}$ , which is low compared to previous studies on *P. multiseriis*, where DA attained  
487  $1.2$  to  $45 \text{ pg cell}^{-1}$  (Bates et al., 2000, Thessen et al., 2009). Our values are more consistent  
488 with those of *P. calliantha* (Álvarez et al., 2009) or *P. pseudodelicatissima* (Pan et al., 2001),  
489 which had a maximum toxicity of  $10$  and  $36 \text{ fg cell}^{-1}$ , respectively, but these species have a  
490 smaller cell volume. Our strain of *P. multiseriis* was really short (around  $20 \mu\text{m}$  length here,  
491 whereas cells can be  $100 \mu\text{m}$  long just after sexual reproduction, which may explain the low  
492 values of DA it produced. DA intracellular content started to decrease from day 11 to the end  
493 of the stationary phase. This decrease of DA may also coincide with a physiological stress.  
494 Unfortunately, bacteria were not measured that day. Nevertheless, day 11 exhibited surprising  
495 values of NR, FL3 and QY (i.e. out of the trend). Cells might have undergone a stress, with

496 loss of chlorophyll and thus decreased QY, thus energy was stored under lipid form and DA  
497 production was stopped. Dissolved DA was particularly low but remained constant over time,  
498 with cells excreting 11 to 40 % of their total DA. This low DA release may be due to the age  
499 of the strain, isolated in 2007, and its consequent smaller size.

500

501 DA is a secondary metabolite and is thus believed to be produced when cells have  
502 excess energy that is not used for primary metabolism (Bates, 1998). Meanwhile, extra  
503 energy is stored as lipids when cells are not able to use it for primary metabolism. In this  
504 study, the measure of FDA provided information regarding primary metabolism and QY  
505 (photosynthetic efficiency) was measured to estimate the production of energy. There was no  
506 clear relationship between DA production (total or dissolved) and QY or FDA hydrolysis.  
507 Conversely, a positive correlation was observed between total DA content and NR after PCA  
508 analysis (Fig. 9). Cells of *P. multiseriis* seemed to produce more DA when they had more  
509 lipids, thus more available stored energy, which is in agreement with some studies (Whyte et  
510 al., 1995) but not all (Pan et al., 1996). Indeed, Pan et al. (1996) made the hypothesis that DA  
511 and lipid synthesis shared some precursors as Acetyl-CoA, so when DA is produced, lipids  
512 can not be stored. Bacteria are also known to play a role in DA production, by enhancing DA  
513 production through unknown mechanisms (Bates et al., 1995, Stewart et al., 1997). In this  
514 study, the ratio of bacteria per *P. multiseriis* cell was also weakly correlated to DA content,  
515 as the variable coordinates are quite close in the PCA analysis ( $R^2=0.49$ ,  $p<0.001$ , Fig. 9).  
516 Cells seemed to produce more DA when more bacteria per *P. multiseriis* cell were present in  
517 the culture, possibly indicating that more DA was produced either when competition with  
518 bacteria was greater or if bacteria produced toxin-enhancing compounds. FSC and BOPIDY  
519 uptake were also closely correlated with DA content, SSC, NR and ratio bacteria/*Pseudo-*  
520 *nitzschia*, indicating that increased DA content is associated with a higher intracellular lipid



521 content. This lipid increase can cause an increase in the amount and/or the size of lipid  
522 vacuoles within the cells, which could also explain the increase observed in FSC and SSC of  
523 the *P. multiseriis* cells. A factorial plan (Fig. 10) was developed from the previous PCA,  
524 which plots the incubation time of the *P. multiseriis* culture, from day 9 to 20 (values  
525 included in the previous PCA). The position of the days included on this factorial plan clearly  
526 demonstrates and summarizes our findings: the gradual and continuous shift of the culture  
527 from low algal concentration, high bacteria/algal ratio, large SSC, high lipid and DA content  
528 in early stationary phase, towards increasing concentrations, reaching a maximum at the end  
529 of the exponential phase, to finally showing a high percentage of dead algal cells and bacteria  
530 in late stationary phase.

531

532 FCM has been previously used on microalgae, mainly to cell count or measure of only  
533 one physiological parameter per experiment. Here, we developed a set of physiological  
534 measurements, which provides a more complete description of the physiological status of the  
535 microalgae. This technique has been applied to one species of *Pseudo-nitzschia* but can be  
536 broadened to other microalgal species, whether or not they are toxic or diatoms. Developing  
537 cell-based physiological measurements with FCM will help to further our understanding of  
538 phytoplankton physiology and its responses to environmental changes, both biotic and  
539 abiotic.

540

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545

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786 **Legends to figures**

787 Figure 1. Cytograms of 50/50 dead/live cells of *Pseudo-nitzschia multiseriis* stained with  
 788 SYTOX Green. A. Cytogram of FSC and SSC (morphological parameters, expressed in  
 789 arbitrary units, AU) of *P. multiseriis*. B. Cytogram of FL1 and FL3 fluorescence of *P.*  
 790 *multiseriis*. FL1 is the green fluorescence due to SYTOX Green, FL3 is the red fluorescence  
 791 due to chlorophyll (AU). R1 are unstained cells (considered as live cells, in red) and R2 are  
 792 stained cells (considered as dead cells, in green).

793 Figure 2. Bacteria stained with SYBR Green and propidium iodide. A. Histogram of FL1  
 794 (green) fluorescence of bacteria; 1 to 7 representing aggregates of 1 to 7 or more bacteria.  
 795 FL1 is the green fluorescence due to SYBR Green. B. Cytograms of morphological  
 796 parameters of bacteria (FSC and SSC, expressed in arbitrary units, AU). Each colour  
 797 represents one aggregate size (light green=one bacteria, dark blue=2 bacteria, pink=3  
 798 bacteria, light blue=4 bacteria, yellow=5 bacteria, red=6 bacteria, dark green=7 or more  
 799 bacteria).

800 Figure 3. Photomicrographs of *Pseudo-nitzschia multiseriis* cells in white light (A, D, G),  
 801 epifluorescence light with filter “BP 515/560 / BS 580 / LP 590” (B, E, H), and filter “BP  
 802 450-490 / BS 510 / LP 515” (C, F, I). A, B, C) Cells stained with BODIPY. D, E, F) Cells  
 803 stained with Nile Red. G, H, I) Cells stained with FDA. Scale bar=10  $\mu\text{m}$ .

804 Figure 4. Green fluorescence of *Pseudo-nitzschia multiseriis* cells (in arbitrary units, AU)  
 805 stained with 3.0  $\mu\text{M}$  of fluorescein diacetate (FDA) and measured on FL1 detector of a flow  
 806 cytometer (n=3, mean  $\pm$  SD).

807 Figure 5. A- *Pseudo-nitzschia multiseriis* growth curve (y-axis) and bacteria/*P. multiseriis*  
 808 ratio (z-axis, n=6, mean  $\pm$  SE). The exponential growth phase of *P. multiseriis* is framed with  
 809 a black-lined rectangle. B- Concentration of total and dissolved domoic acid (DA) in the  
 810 whole culture (y-axis,  $\text{pg ml}^{-1}$ ) and cellular DA in  $\text{fg cell}^{-1}$  (z-axis, n=6, mean  $\pm$  SE).  
 811 Exponential growth phase of *P. multiseriis* is framed with a black-lined rectangle.



812 Figure 6. Chlorophyll fluorescence (FL3, in arbitrary units, AU, y-axis) and Quantum Yield  
813 (QY, z-axis) of live *Pseudo-nitzschia multiseriis* cells as a function of culture age. FL3 was  
814 measured using flow cytometry on live cells, as determined by SYTOX Green staining (n=6,  
815 mean  $\pm$  SE). The exponential growth phase of *P. multiseriis* is framed with a black-lined  
816 rectangle.

817 Figure 7. Fluorescein diacetate (FDA) uptake (FL1 fluorescence of live cells, y-axis) and  
818 percentage of *Pseudo-nitzschia multiseriis* live cells stained by FDA (z-axis) and detected  
819 using flow cytometer FL1 detector (n=6, mean  $\pm$  SE). Exponential growth phase of *P.*  
820 *multiseriis* is framed with a black lined rectangle. Correlation between the percentages of live  
821 cells measured with the SYTOX Green and FDA assays is indicated in the small graph (in  
822 arbitrary units, AU).

823 Figure 8. Green and orange fluorescences of *Pseudo-nitzschia multiseriis* cells stained with  
824 BODIPY 493/503 and Nile Red (indicators of lipid content) and detected by the FL1 (y-axis)  
825 and FL2 (z-axis) detectors, respectively, on a flow cytometer, in arbitrary units (n=6, mean  $\pm$   
826 SE). Exponential growth phase of *P. multiseriis* is framed with a black lined rectangle.

827 Figure 9. Principal Component Analysis (PCA) plot of all physiological measurements  
828 between days 9 and 20 (D9 to D20) of the *P. multiseriis* culture (n=52).

829 Figure 10. Factorial plan issued from the previous PCA and plotting days of culture of *P.*  
830 *multiseriis*, from day 9 to day 20 (D9 to D20, n=52).

831

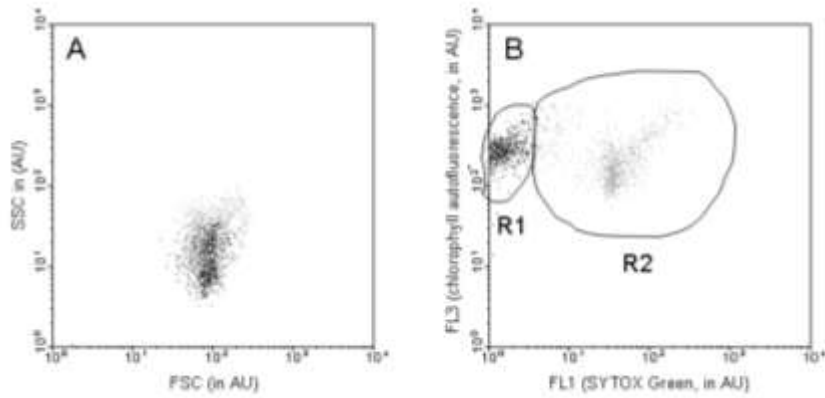
832 Table 1. *Pseudo-nitzschia multiseriis* and associated bacteria concentration (of live cells),  
 833 morphological parameters (FSC and SSC, in arbitrary units), percentage of dead *P.*  
 834 *multiseriis* measured using SYTOX Green and percentage of dead bacteria assessed using  
 835 SYBR Green - propidium iodide double staining (n=6, mean  $\pm$  SE).

Day	<i>P. multiseriis</i>								Bacteria							
	FSC (AU)		SSC (AU)		% dead cells		Concentration (cell ml <sup>-1</sup> )		FSC (AU)		SSC (AU)		% dead cells		Concentration (10 <sup>6</sup> bact ml <sup>-1</sup> )	
	mean	$\pm$ SE	mean	$\pm$ SE	mean	$\pm$ SE	mean	$\pm$ SE	mean	$\pm$ SE	mean	$\pm$ SE	mean	$\pm$ SE	mean	$\pm$ SE
4	196.3	2.6	76.8	0.7	25.0	1.2	4 526	190	226.4	10.8	23.5	0.7	5.8	0.6	3.01	0.07
5	203.5	0.7	76.9	1.4	27.9	1.7	4 574	255	207.3	9.1	23.1	0.5	4.2	0.3	4.14	0.15
6	186.0	1.3	80.0	1.8	22.6	0.9	5 452	214	179.1	5.2	22.9	0.4	4.0	0.3	3.80	0.13
7	197.6	2.7	71.2	1.6	32.7	0.7	3 533	93								
8	183.7	2.9	72.6	1.5	30.6	0.8	6 226	275	177.4	2.8	23.0	0.2	3.5	0.3	5.61	0.04
9	185.2	1.5	72.1	1.0	32.6	2.0	7 711	677	179.1	4.6	24.5	0.3	2.6	0.3	5.33	0.23
10	190.5	2.0	67.5	1.3	28.5	2.7	9 059	1 179	168.9	7.5	24.3	0.4	2.7	0.2	5.15	0.20
11	190.5	1.6	62.9	0.7	54.5	9.4	12 393	1 730								
12	186.3	1.4	63.3	0.9	30.3	3.8	15 863	2 716	127.6	5.0	23.2	0.4	2.0	0.2	6.09	0.21
13	180.4	1.3	59.1	0.8	22.8	3.0	23 837	5 343	186.5	18.7	23.0	1.5	1.9	0.1	6.60	0.12
14	177.4	2.0	58.7	1.1	22.9	2.9	30 726	6 223	171.4	26.7	20.2	0.6	2.2	0.2	6.80	0.21
15	173.5	0.9	59.7	1.3	19.1	0.7	33 796	4 967	193.3	7.7	21.1	0.7	2.2	0.1	7.71	0.23
16	171.1	1.9	60.0	1.3	18.9	1.6	36 148	2 064	225.9	18.1	22.7	1.4	2.2	0.1	8.19	0.16
17	185.4	0.9	57.3	1.2			77 322	3 718								
18	173.9	3.1	60.2	1.3	27.4	4.2	49 222	4 818	99.2	5.9	19.3	0.7	1.9	0.1	8.72	0.55
19	174.9	2.1	56.3	0.7			78 730	10 135								
20	168.8	1.3	60.3	0.3	43.8	5.5	55 889	8 682	177.9	8.6	19.0	0.7	2.0	0.1	11.31	0.97
21	163.7	2.3	59.7	0.5			40 163	2 226	25.1	0.7	17.3	0.4	2.1	0.2	13.30	0.81

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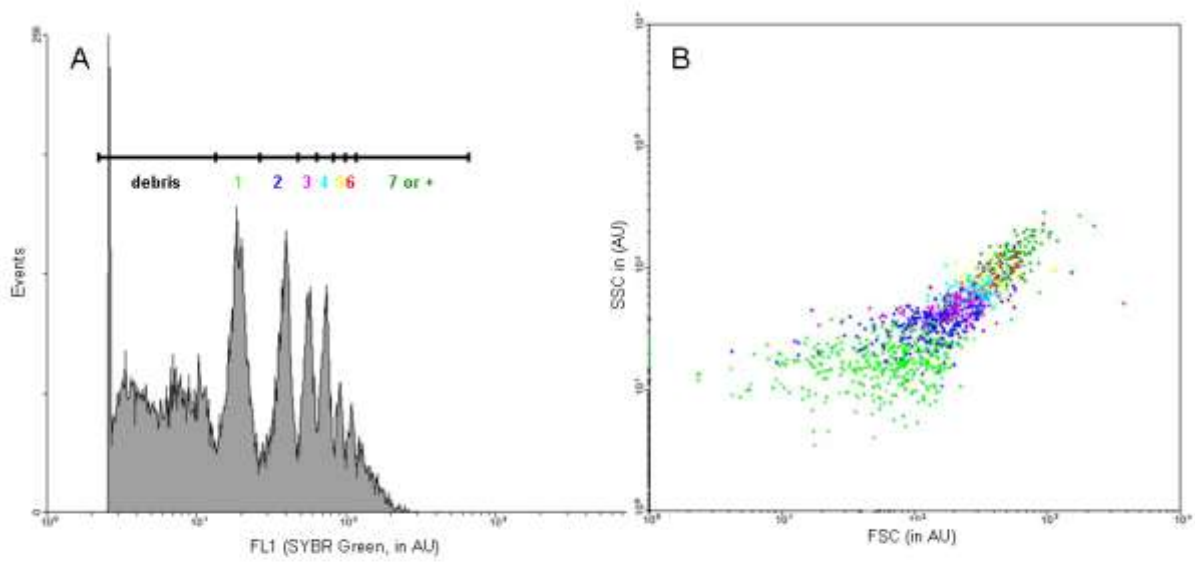
837 **Figures**

838 Fig. 1



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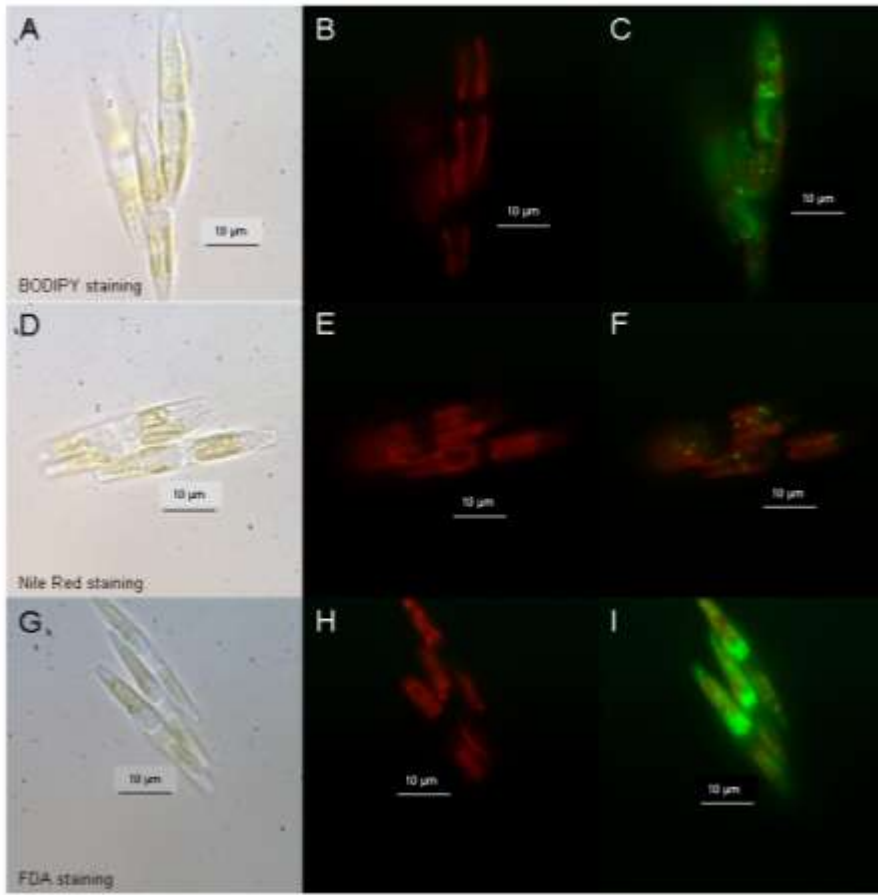
840 Fig. 2



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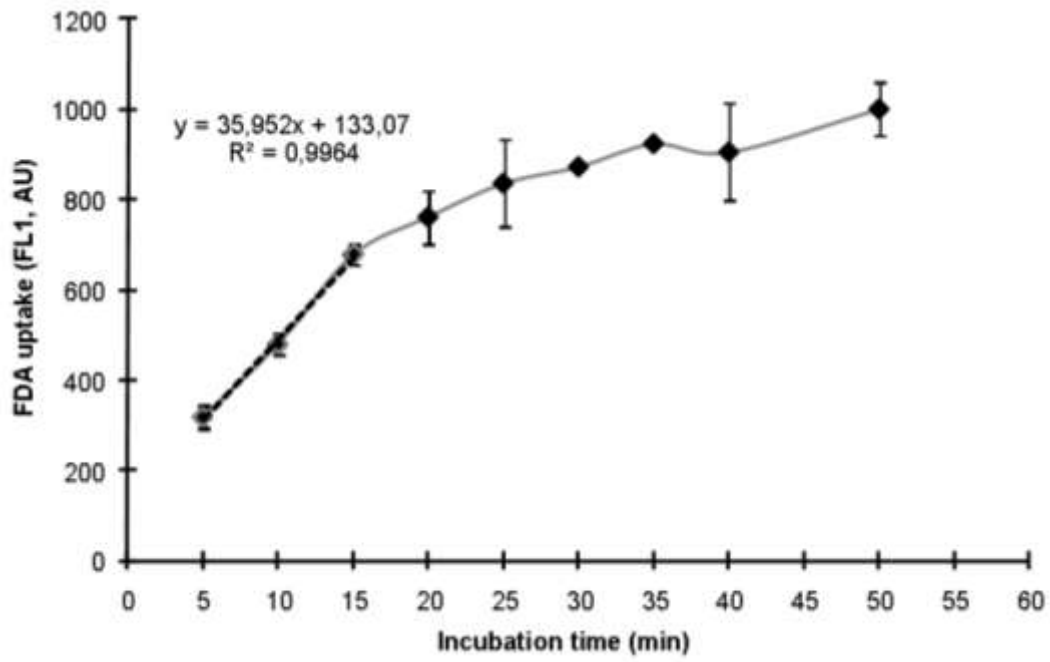
843 Fig. 3



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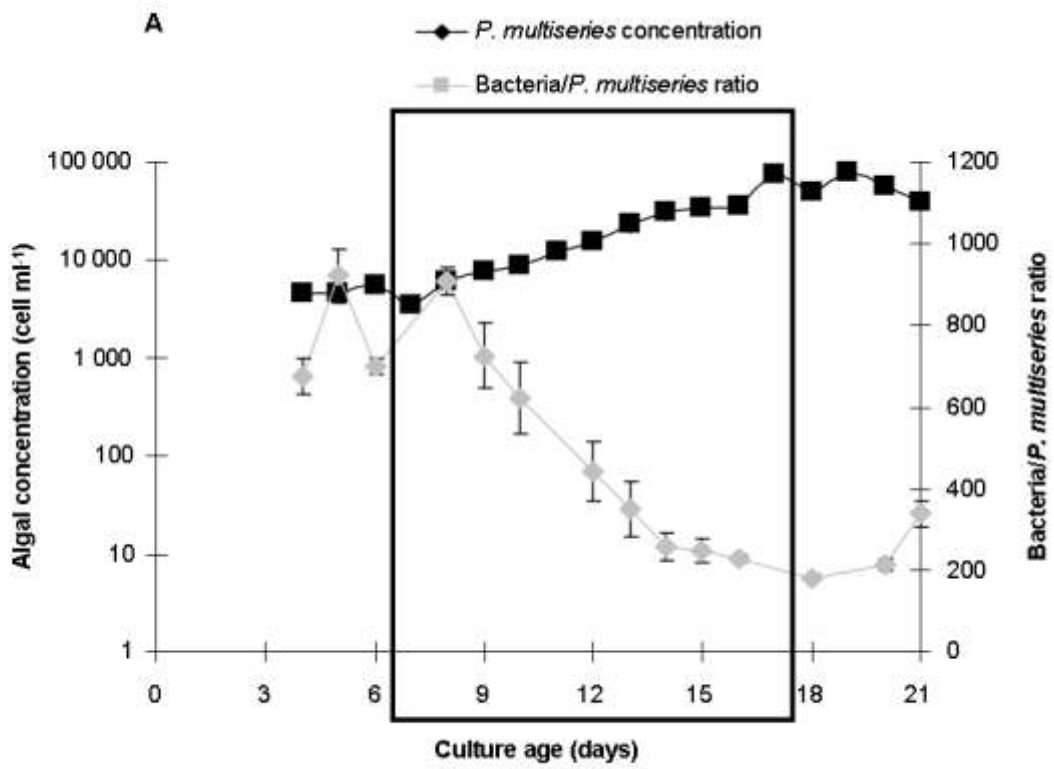
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846 Fig. 4

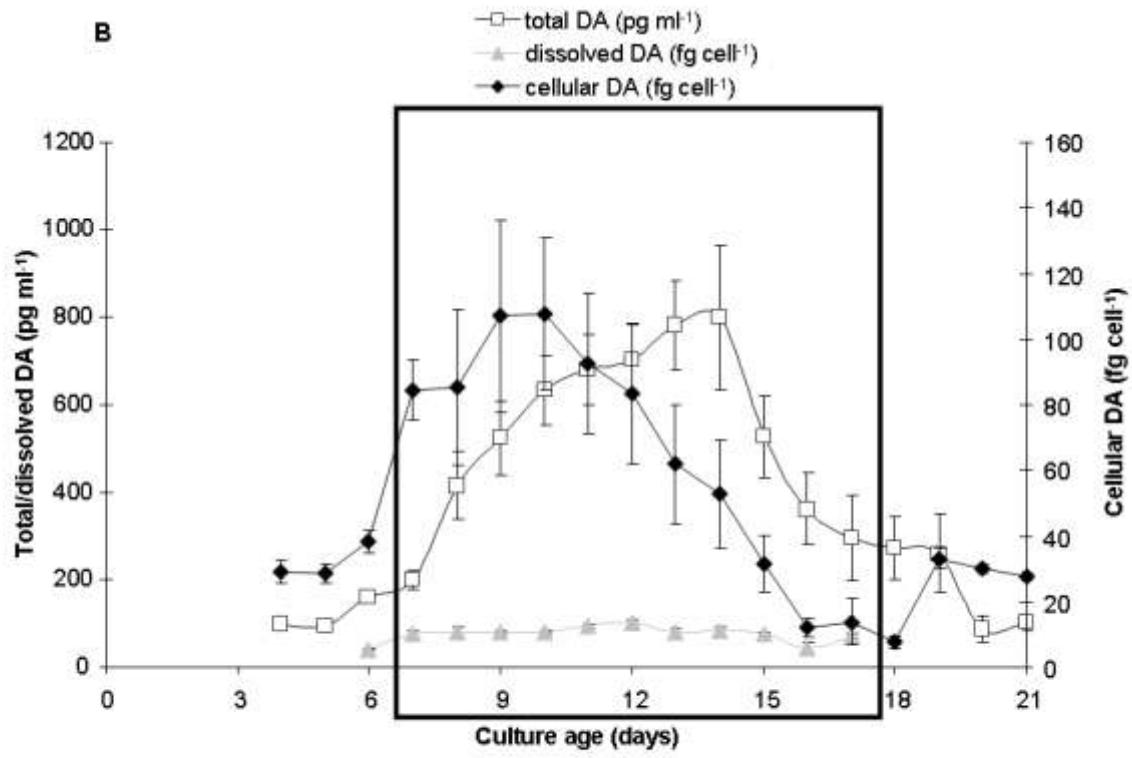


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848 Fig. 5

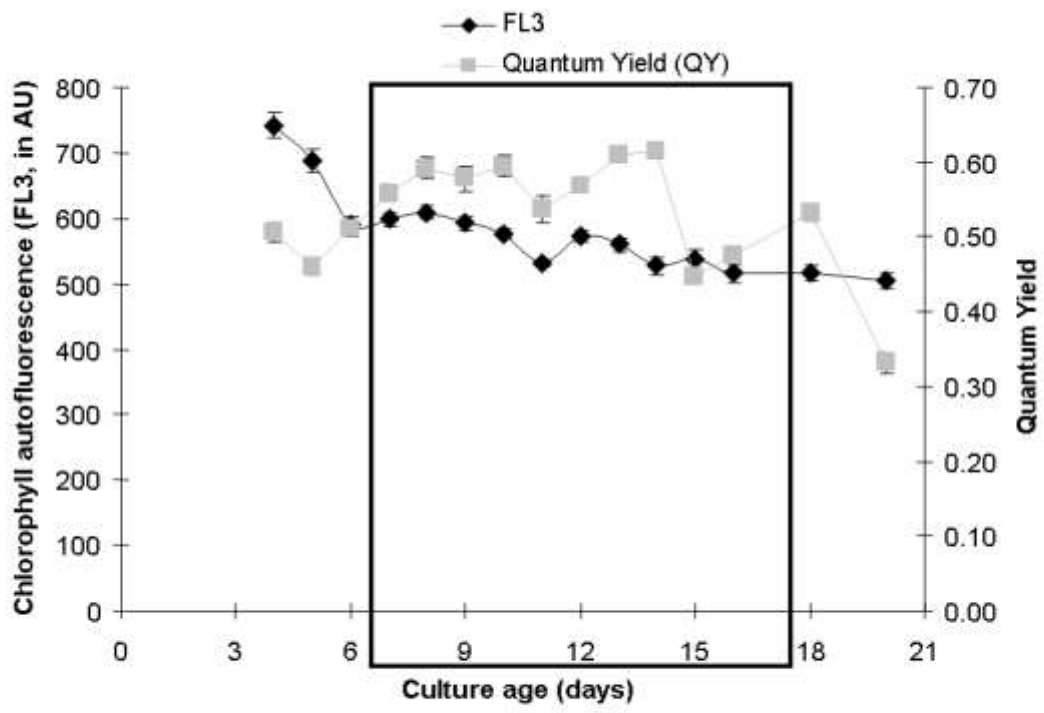


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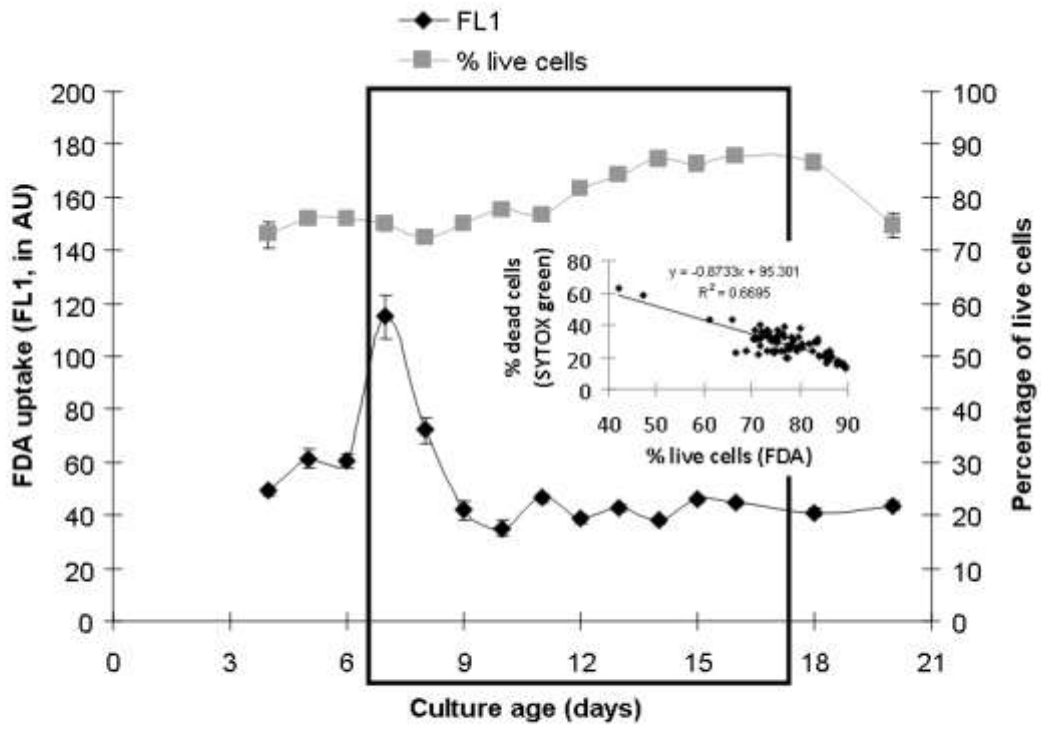
851 Fig. 6



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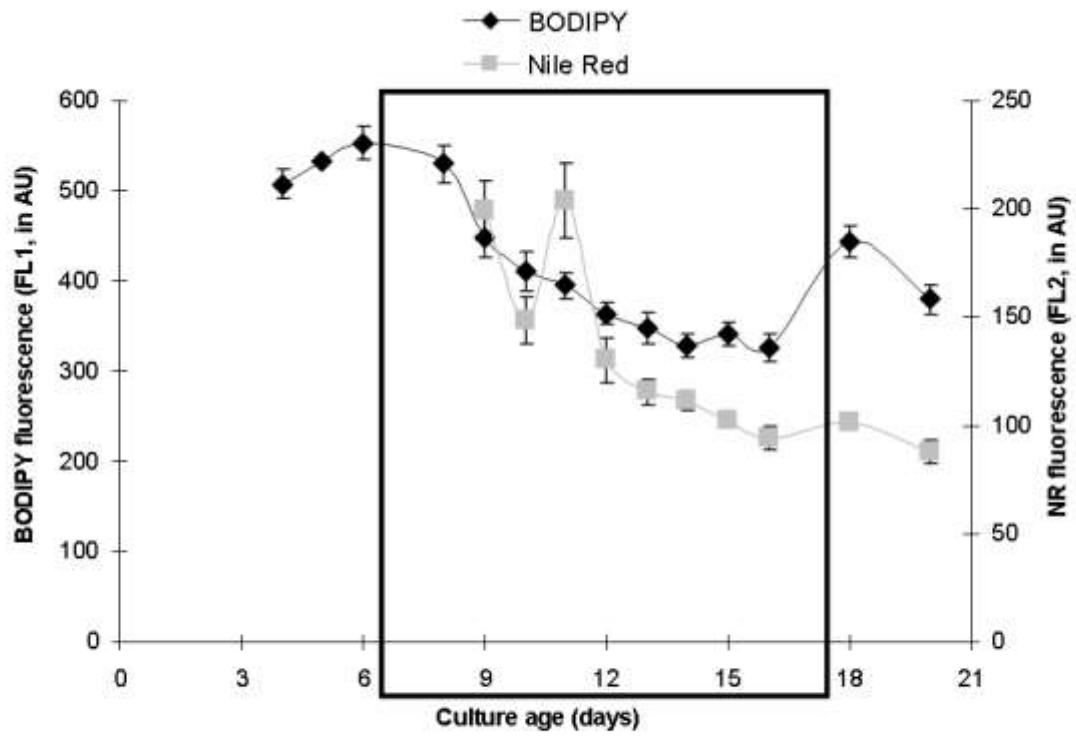
854 Fig. 7



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856 Fig. 8





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858 Fig. 9

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861 Fig. 10

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