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Recent population expansion and connectivity in the hydrothermal shrimp *Rimicaris exoculata* along the Mid-Atlantic Ridge

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Running head: Genetic diversity of a hydrothermal vent shrimp

**Abstract**
Aim: Deep-sea hydrothermal vents are unstable habitats that are both spatially and temporally fragmented. In vent species, a ‘short-term insurance’ hypothesis would lead us to expect mostly self-recruitment, limiting the loss of larvae in the deep ocean or water column and increasing genetic differentiation over the time elapsed since colonization. Alternatively, a ‘long-term insurance’ hypothesis would support the prediction of selection for large scale dispersal, to ensure long-term persistence in these ephemeral habitats. The main goal of this study was to infer the spatial and temporal distribution of genetic diversity of the shrimp *Rimicaris exoculata*, which forms high density local populations on hydrothermal vents along the Mid-Atlantic ridge.

Location: Deep-sea hydrothermal vents along the Mid-Atlantic Ridge.

Methods: We used sequences of mitochondrial cytochrome c oxidase subunit I (COI, 710 bp) to assess the spatio-temporal distribution of genetic diversity across five hydrothermal fields from 36°N to 4°S.

Results: In contrast to previous results from pioneer studies, very high haplotype diversity was observed in vents across the entire region (i.e. 0.69 to 0.82), indicating current large effective population size and low drift. The star-like shape of the network of haplotypes, the lack of spatial genetic structure and the significance of tests reflecting demographic effects, together with the fitting of a population expansion model, all support a recent population expansion.

Main conclusions: Our results suggest a very recent common history of *R. exoculata* populations/demes along the Mid-Atlantic Ridge, derived after a common bottleneck or founder event and followed by a concomitant demographic expansion. This study therefore suggests large effective population size and/or high dispersal capacity, as well as a possible recent (re)colonization of the Mid-Atlantic hydrothermal vents by *R. exoculata*.

Keywords: Bottleneck, deep sea biogeography, dispersal, genetic diversity, hydrothermal vents, marine biogeography, Mid-Atlantic Ridge, *Rimicaris exoculata*, spatio-temporal distribution.
Introduction

Dispersal plays a central role in both the dynamics and evolution of species. Effective dispersal (i.e., gene flow) attenuates genetic differences among populations and increases the genetic variability within populations (Billiard & Lenormand, 2005). Dispersal is a risky strategy as immigrants may never reach a suitable habitat, or face higher mortality and lower reproductive success in the receiver population (Ims & Andreassen, 2000; Garant et al., 2007). Yet, dispersal can also offer clear advantages, particularly in fragmented ephemeral habitats, as it allows areas to be explored for more favourable habitats and extinction to be avoided by escaping locally deteriorating physical or biotic conditions, such as competition (Hamilton & May, 1977; Olivieri et al., 1995). By enabling populations to randomly spread their progeny more evenly among different sites dispersal may indeed represent a favourable bet-hedging strategy in temporally variable or ephemeral ecosystems (Ronce & Olivieri, 2004).

Deep-sea hydrothermal vents are an extreme example of a spatially and temporally unstable environment. Environments of this type are scattered worldwide along the global mid-ocean ridge system, and are sometimes thousands of kilometres apart (Tunnicliffe & Fowler, 1996; Van Dover, 2000). They are extreme and unique habitats, with communities composed of specialized animals, dependent on chemoautotrophic microorganisms that exploit the reduced compounds (mainly H$_2$S and CH$_4$) abundant in vent fluids (Desbruyères et al., 2000). Each hydrothermal vent has a characteristic profile that may differ in depth, temperature and/or chemical composition (Van Dover, 2000), adding abiotic heterogeneity to the spatial component of habitat fragmentation that potentially influences the patchy community distribution and limited effective dispersal. Besides the distance separating successive hydrothermal vents, communities may also become genetically isolated across transform faults, which act as potential barriers between populations and limit effective dispersal.
among sites (Plouviez et al., 2009). This hypothesis is supported by the high levels of endemism found across biogeographic provinces (Van Dover et al., 2002; Fabri et al., 2006; Bachraty et al., 2009; Moalic et al. submitted) as well as by the rather high genetic differentiation found across some neighbouring sites (e.g. Jollivet et al., 1999; Won et al., 2003; Hurtado et al., 2004; Matabos et al., 2008; Young et al., 2008; Plouviez et al., 2009), which indeed suggest a limited effective dispersal over large distances. On the other hand, depending on the sea floor geology and magmatic and tectonic events, individual hydrothermal vents have a relatively short lifespan of 1 to 100 years for fast spreading ridges such as the East Pacific Rise (Hessler et al., 1988; Lalou et al., 1993), and up to 300 years in more stable, slow spreading ridges such as the Mid-Atlantic Ridge (Brazelton et al., 2010). These common temporal fluctuations in vent activity and changes in the geochemical profile of their habitat prevent long-term local population persistence (Tunnicliffe, 1991; Jollivet, 1996). Those species that escape local extinction due to habitat disappearance and which are capable of (re)colonizing newly active sites may also have the capacity for long-distance effective migration. Species associated with hydrothermal vents should therefore also have good colonization abilities with high rates of dispersal, growth and fecundity (Vrijenhoek, 1997; Tyler & Young, 1999). Indeed, in 1991 the formation of new hydrothermal vents in the East Pacific Rise was studied and, astonishingly, microbe-derived material was found in active areas of the vent immediately following their formation. Within the first year, mobile vent and non-vent fauna such as amphipods, copepods, crabs and non-vent shrimps then proliferated (Shank et al., 1998a). These vastly differing sources of influence lead to contradictory expectations about strategy, which have yet to be tested. On one hand, low effective dispersal and high self recruitment may be favoured across limited distances due to the patchy nature of vent distribution and abiotic characteristics. On the other hand, larger scale dispersal potential may be selected for in the long term, allowing species persistence despite instability of vent activity. Finally, temporal instability (Desbruyères et al., 1998), although
apparently less pronounced in the Mid-Atlantic Ridge (MAR) than in the East Pacific Rise (EPR) (Copley et al., 2007 on one site), may also result in short-term changes in genetic composition.

Along the Mid-Atlantic Ridge (MAR), vent communities are dominated by shrimps, mussels, anemones and crabs (Tunnicliffe, 1991; Van Dover, 1995). The first described and the most abundant species is the shrimp *Rimicaris exoculata* Williams & Rona, 1986. This species is often present in high-density swarms (> 1500 individuals/m²) around high-temperature sulphide chimneys at deeper hydrothermal vent sites (Segonzac et al., 1993; Copley et al., 1997). The species has highly modified eyes that form a single dorsal organ, which has been suggested to detect black-body radiation associated with high temperature vents (Van Dover et al., 1989; Van Dover, 1995). This shrimp does not seem to be a scavenger or a predator because its gut is filled with minerals, mainly iron oxides and sulphurs, and its nutrition is hypothesized to be derived from several epibiotic bacterial sources, mainly those located in the gill chambers and gut (see Durand et al., 2010 and references therein). Although hydrothermal vents at MAR have been known since 1985 (Rona et al., 1986), little is known about population genetic structure of its most abundant shrimp species. Genetic studies conducted to date have focused on clarifying the systematic status of the juvenile morphotype of *R. exoculata* (Creasey et al., 1996; Shank et al., 1998b). The allozyme analyses conducted in these studies showed very low genetic diversity within sites and essentially no genetic differentiation between populations, suggesting that there is high gene flow.

The main goal of this study was to infer the spatial and temporal distribution of genetic diversity of *R. exoculata* from different hydrothermal vents along the Mid-Atlantic Ridge, in order to assess the extent of effective population sizes, rates of dispersal and recent demographic histories at these sites. We used mitochondrial cytochrome c oxidase subunit I (COI) as a molecular marker to estimate the level of genetic variability in natural populations, to test for genetic subdivision and to obtain insights into population history, dynamics and demography at the ridge level. In order to assess the
effect of potential temporal instability of hydrothermal activity and population dynamics of
associated species, two of the five populations studied were analysed at time intervals of one or ten
years.

Materials and Methods

Sampling and DNA extraction

Samples were collected from five locations corresponding to different hydrothermal vent fields
(Rainbow, Logatchev, TAG, Ashadzé and South MAR) along the Mid-Atlantic Ridge (Fig. 1, Table
1). For two locations (Rainbow and Logatchev), samples from different years were used to assess
temporal variability; a time interval of 10 years (Rainbow 1997 and 2007) and 3 consecutive one
year intervals (Logatchev 2005, 2006, 2007) were used. Samples were collected using a slurp-gun
from the ROV (Remotely Operated Vehicle) or the Nautile. Prior to each dive, the bowls used for
collecting the shrimps were aseptically washed with ethanol (96 %) before being filled with sterile
seawater. Once on board, live specimens were either entirely frozen or immediately dissected into
body components under sterile conditions and frozen or stored in 70 % alcohol. DNA extraction was
performed using the CTAB (cetyl trimethyl ammonium bromide) method (Doyle & Doyle, 1990) on
muscle tissue. Sampling size could not be pre-defined to a constant value across all sampling sites
and times due to the overall difficulty in obtaining samples from deep hydrothermal vents; in some
sites sampling size was thus limited to the shrimp samples available from Ifremer research cruises
for each particular location and time, and (for the 4 °S location) some samples kindly provided by
Max Plank Institute (N. Dubilier).
Polymerase chain reaction and sequencing

Mitochondrial COI was amplified using the universal primers LCO1490 and HCO2198 described by Folmer et al. (1994). All polymerase chain reaction (PCR) amplifications were carried out in 50 µL volumes containing 50 ng DNA, 1× reaction buffer (GoTaq, Promega), 0.2 mM of each dNTP, 2.5 mM MgCl₂, 0.4 U Taq DNA polymerase (GoTaq, Promega, Madison, WI, USA) and 0.6 µM of each primer. The PCR amplification was conducted in a Perkin-Elmer Gene Amp System 7200 (Waltham, MA, USA) with the following program: 2 min. at 95 ºC; 35 cycles composed of 1 min denaturation at 95 ºC, 1 min at 52 ºC and 1.5 min elongation at 72 ºC, followed by a final elongation step of 7 min at 72 ºC. PCR products were sent to be purified and sequenced commercially at Macrogen, Inc. (Seoul, Korea). Sequences were proofread and aligned using Geneious software (Drummond et al., 2009). The 18S rDNA (~1.7 kb) was amplified for 4 individuals of all sampled locations and years (except for the Ashadzé population, where only the three individuals available were amplified) using 18S-82F/18S-1498R primers (López-García et al., 2003) and sequenced in an ABI 3130 XL automatic sequencer (Applied Biosystems, Foster City, CA, USA), to provide hints into nuclear variation. The nuclear regions ITS 1 and 2 (~931 bp) were also amplified for four individuals of three populations (Rainbow, Logatchev and South MAR) using the primer pair ITS 1 and 4 (Kumar & Shukla, 2005). The obtained sequences were cloned (PGem-T easy vector, Promega) following the supplier’s protocol; four clones per individual were sequenced using the ABI 3130 XL automatic sequencer (Applied Biosystems).

No diversity was detected across the 4 individuals from the 5 sites sequenced for 1200 bp of 18S rDNA. The amplified regions ITS1 and 2 proved to be too variable to be used: all the sequences obtained were different, including sequences from different clones of the same individual.

Data analysis
ARLEQUIN version 3 (Schneider *et al*., 2000) was used to estimate gene diversity and conduct statistical tests. For each population, the following statistics were computed: haplotype \( h \) (Nei, 1987) and nucleotide diversities \( \pi_2 \) (Nei, 1987), and mean number of pairwise differences \( \pi_1 \) (Tajima, 1983). For populations with significantly different sample sizes, a re-sampling method was applied using R to generate files for 100 random re-samplings of the population haplotype frequencies to a minimum common size. The 100 generated files were then analysed with the batch file function of ARLEQUIN software to re-estimate gene diversity in these sub-samples. To assess population differentiation, pairwise \( F_{ST} \) values were calculated following the method of Hudson *et al*. (1992) and exact tests of differentiation were conducted following the method of Raymond & Rousset (1995).

In order to analyse and illustrate the phylogenetic relationships among haplotypes, i) a median-joining network was constructed using Network v. 4.1.0.9 (Bandelt *et al*., 1999) to infer the most parsimonious branch connections between the sampled haplotypes, and ii) a neighbour-joining phylogram was constructed using the mutation model HKY (Geneious software; Drummond *et al*., 2009) and the bresiliid sister species *Alvinocaris* sp. as an outgroup.

We tested whether the sampled populations fitted mutation drift equilibrium expectations, and assessed which population demographic model was most appropriate, i.e. the size-stationary model or the expansion model. We determined Fu’s \( F_s \) (Fu, 1996) and Tajima’s \( D \) (Tajima, 1989), which are sensitive to departures from both selective neutrality and from population size equilibrium caused by expansions or bottlenecks (Tajima, 1996; Fu, 1997). Both statistics result in negative values after a population expansion (Ray *et al*., 2003) or a selective sweep, whereas positive values are expected under balancing selection of recent bottlenecks. ARLEQUIN was used to perform tests and estimate the associated \( P \)-values. Mismatch curves were drawn for each population and observed values were
compared to the expected curves fitted for the constant population size model and for the population
growth/decline model, using the DnaSP 4.0 program (Rozas et al., 2003).

**Results**

*Genetic diversity and population differentiation*

A total of 47 haplotypes were recovered from the 152 individuals analysed, 30 (63.8 %) of the
haplotypes were “private”, i.e. unique to a single locality (Table 2; Fig. 1; accession numbers:
HM125910 to HM125956). The most common haplotype was present in all population samples and
was the most abundant haplotype across all populations, identified in 49 % of all the sampled
individuals across the entire study region. This main haplotype was also central to all other
haplotypes, most of these represented by a single individual and divergent by a single point mutation,
leading to this haplotype being the core of a star-like cluster topography (Fig. 2). The remaining
haplotypes (36.2 %), although not unique, were mainly (27.7 %) represented by only one individual
in each population. Accordingly, the neighbour-joining phylogram revealed extremely unresolved
clusters supported by poor bootstrap values (see Appendix S1 in Supporting Information).

At a temporal scale of a 10-year sampling interval (Rainbow 1997 and 2007), the unique haplotypes
(present in one year and not on the other) represented 56 % of the total haplotypes found, whereas on
the temporal scale of a one year sampling interval (Logatchev 2005, 2006, 2007) the proportion of
unique haplotypes was higher (88.5 %).

Haplotype diversity ($h$) was high, ranging from 0 to 1 in populations with small sample sizes
(Ashadzé: 3 individuals and 1 haplotype; South MAR: 4 individuals and 4 haplotypes) and 0.69 to
0.82 in populations with larger sample sizes (>11 individuals). Considering these rather high values
of haplotype diversity, nucleotide diversity ($\pi_2$) was low, ranging from 0 to 0.0033 (Table 2). Highest diversities were recorded in the South MAR population, although the sample size was very low (4 individuals), followed by the Rainbow 2007 population. Populations sampled in more than one year presented different diversity indices across years. The Rainbow population sampled in 2007 showed more polymorphic sites, and higher haplotype and nucleotide diversity than when it was sampled in 1997, even though the number of samples was lower in 2007 (26 individual) than in 1997 (32 individual). A random re-sampling method was therefore not needed for Rainbow, as the sample with the fewest individuals already presented the higher diversity indices. Random resampling was, however, performed for Logatchev, where samples had more contrasting sizes (12 to 31 individuals) with no obvious diversity trend through time. In order to dissociate a putative biological effect from a sampling size effect, the random re-sampling (100 times) was therefore performed to a minimum common size of 12 individuals per sampling time, and no significant differences were found in any of the diversity indices over time (Table 3).

No significant population differentiation was detected between any of the sampling sites or years using pairwise $F_{ST}$ estimates (Table 4).

**Historical demography**

Demographic analyses suggested the occurrence of population expansions, as highly significant negative values for Tajima’s $D$ and Fu’s $F_S$ were obtained for all populations (Table 2). Mismatch distribution curves were consistent with models of population expansion, displaying a unimodal distribution (Fig. 3). When comparing the expected and observed mismatch curves under constant population size *versus* the growth/decline model (Fig. 4), there is an excess of rare variants in the observed curves in relation to that expected under constant population size. Furthermore, the observed curves seem to better follow the expected curve for the growth/decline model.
Discussion

The analysis of mtDNA haplotypes from a total of 152 individuals from five discrete vent localities along the Mid-Atlantic Ridge showed an overall high genetic variability within the studied populations. Both the number of haplotypes per population (1 to 16) and gene diversity (0.69 to 0.82) are within the range of values found for other vent organisms studied with the same marker, including tubeworms (e.g. *Riftia pachyptila*, *Alvinella pompejana*), limpets (e.g. *Lepetodrillus elevatus*) and bivalves (e.g. *Bathymodiolus azoricus*, *Calyptogena magnifica*) (e.g. Hurtado *et al.*, 2004; Johnson *et al.*, 2006; Plouviez *et al.*, 2009). However, these results contradict those obtained for *Rimicaris exoculata* with allozymes, where genetic variability was amongst the lowest reported for vent organisms (Creasey *et al.*, 1996). This may be due to the different properties of the distinct markers because allozymes may exhibit lower polymorphism than mtDNA, possibly associated with protein constraints. Mitochondrial DNA polymorphism is classically considered as less influenced by possible selective processes (Ballard & Whitlock, 2004), although it was recently demonstrated that this may not be a systematic rule (Galtier *et al.*, 2009). In any case, DNA sequences retain more polymorphisms than proteins, as polymorphic DNA sequences include synonymous mutations; higher variability can, therefore, be detected by DNA sequences than by allozymes. On the other hand, a differential imprint of demographic events on mitochondrial versus nuclear DNA may be evoked, although lower diversity is usually expected for mtDNA in relation to nuclear DNA, due to the difference in effective population size. Such discrepancies may also be affected by the more restricted sampling that was available for the allozyme study, which included only two sampling locations despite the higher overall number of samples analysed (448 individuals).
The rather unresolved phylogram, with low bootstrap values, shows a generally low divergence without a geographical imprint. Together with the non-significant $F_{ST}$ values obtained in this study, these results indicate a lack of geographic partitioning of the numerous but slightly divergent haplotypes among the sampled populations. This lack of genetic structure may be due to several non-mutually exclusive factors: high levels of effective dispersal (gene flow) between all populations of the Mid-Atlantic ridge, i.e., panmixia at the ridge level; large effective population sizes ($N_e$); and/or recent common population history. The life cycle of *Rimicaris* is poorly understood. Some authors have suggested that *R. exoculata* might feed in the upper water column during larval dispersal (Dixon & Dixon, 1996), and others that the juvenile stages of these shrimp differ physiologically from the adults (Pond *et al.*, 1997; Allen *et al.*, 1998) in traits such as wax-ester reserves and possibly photosynthetically derived lipids. These findings led the authors to hypothesize that the species had a long planktotrophic phase with long-distance migration achieved *via* the surface, so that some of the barriers usually considered, (depth, seafloor topology and transform faults) would not constrain this species. Yet, such a hypothesis implies either a huge risk factor in the case of passive dispersal, or an extremely sophisticated mechanism of active and deterministic migration (that has not been identified so far) allowing a return to hydrothermal habitat at the end of larval development. In any case, whether dispersal occurs *via* the surface or not, the little information available to date on *R. exoculata* suggests a significant dispersal potential either at larval or at adult stage (although no observations of adults away from hydrothermal sites have been reported so far).

The low degree of genetic differentiation among populations may also be attributed to a relatively recent common history together with large effective population sizes, which limit genetic drift (Crow, 1986). The scenario of (1) high levels of gene flow and that of (2) low gene flow since a recent divergence and large $N_e$ may both leave similar signatures of low genetic structure of populations. The hypothesis of large effective population sizes limiting drift is concordant with the
population biology of this species, as *R. exoculata* is reported to have remarkably high population densities of over 1500 individuals / m² (Copley *et al.*, 1997). Such large population sizes may, in the absence of temporal fluctuation or significant variance in reproductive success, result in large effective population sizes, limiting the differentiating effects of drift. Yet, the haplotype phylogram and network, together with the analyses of population history, indicate a recently very low effective population size as they show: (1) a star-like haplotype network where many new haplotypes are derived from one dominant haplotype via single mutation steps, indicating that they are the product of recent mutation events; (2) negative and highly significant values of Tajima’s *D* and Fu’s *F*_S tests, as expected under population bottleneck/expansion scenarios; and (3) the expected and observed mismatch curves that follow a growth/decline model. This profile supports a scenario of relatively young populations sharing a recent common history of bottleneck or founder events, and expansion.

Recent study of epibionts of the *R. exoculata* gill chamber (Petersen *et al.*, 2009) showed spatial clustering of bacterial sequences across the different sites (Rainbow, Logatchev and South MAR). Such geographical segregation of the symbionts compared to host homogeneity may be due either to the influence of environment on symbionts, or to the very recent common history of *R. exoculata* populations with recent divergence only detectable on short generation symbionts but still undetectable on the genetics of their hosts.

The analysis of temporal variability revealed a lack of genetic differentiation among samples separated by time steps of one (Logatchev) and ten years (Rainbow), although within population diversity increased within a decade in Rainbow. In contrast to Logatchev, where no significant differences were observed across three consecutive years, Rainbow exhibited a significantly higher genetic diversity in 2007 compared with ten years earlier, despite the lower sample size and equal number of haplotypes (but more evenly distributed) in 2007. Genetic diversity, quantified both in terms of allelic richness and heterozygosity, increased significantly over the ten year interval in
Rainbow, in agreement with a possible ongoing effect of population expansion (Table 2). Such trends were not detected in the diversity indices for the one year interval samples, possibly indicating that temporal variations in diversity are not sufficiently large to be detected over the shorter time scale of one year intervals. The pattern of variability across temporal samples therefore suggests a contemporary expansion, over one decade, at least in Rainbow. However, population growth tests are known not to perform well when population expansion is very recent (Ramos-Onsins & Rozas, 2002), and any expansion episodes detected using the growth decline model are, therefore, likely to be older than a decade. Altogether, with the star-like cluster of haplotypes revealing the accumulation of mutations over a recent evolutionary scale, these results imply either successive episodes of expansion or one long-term expansion episode affecting the pattern of haplotype diversity at both ecological and evolutionary time scales.

Considering the demographic history of these apparently rather young genetic lineages, which have probably undergone rather recent recolonization and/or expansion events, the lack of genetic differentiation among sites does not allow us to make inferences about the ongoing occurrence of extensive or restricted gene flow among the hydrothermal sites distributed along the Mid-Atlantic Ridge. Yet, in the absence of any selective hypothesis explaining the star-like pattern of COI polymorphism, the hypothesis of such a recent and global (re)colonization also implies a high dispersal potential *per se* in the rather recent past. Recent bottlenecks and/or expansion have also been observed across populations of several hydrothermal invertebrates along other oceanic ridges (Young *et al*., 2008; Faure *et al*., 2009; Plouviez *et al*., 2009) and such profiles of young lineages may be a rule rather than an exception for hydrothermal vent-associated species, due to the dynamic nature of vents subject to recurrent extinction events. In other ridges, such as the East Pacific Rise, most species studied to date also display a clear North/South genetic differentiation and in some cases even highly significant $F_{ST}$ values within the north and south populations (Hurtado *et al*., 2004;
Matabos et al., 2008; Plouviez et al., 2009). This genetic differentiation reported among populations of different vent species may be due to a lower dispersal potential and/or effective population size of these organisms, or to a higher occurrence or strength of physical and abiotic barriers to gene flow, such as transform faults, in the East Pacific Rise than in the Mid-Atlantic Ridge (but see O’Mullan et al., 2001). Other species, however, do not exhibit such cleavage (Won et al., 2003; Hurtado et al., 2004; Plouviez et al., 2009), which may indicate a species-specific characteristic (such as dispersal potential or population size), or might simply reflect different sampling schemes. Large gaps in the sampling were indeed observed for most species where a north/south cleavage was shown and such a sampling scheme was demonstrated by Audzijonyte & Vrijenhoek (2010) to have the potential to result in artificially inflated $F_{ST}$ values. Yet, in the study of the sister mussel species Bathymodiolus azoricus and B. puteoserpentis and their hybrids (O’Mullan et al., 2001), the clear distinction in species distribution between north (Rainbow, Lucky strike and Menez Gwen vents) and south (Snake Pit and Logatchev vents) along the MAR, suggests the existence of an ancient vicariance event that apparently does not affect Rimicaris exoculata, possibly due to a more recent history, higher effective population sizes, or higher dispersal potential in this shrimp species.

As few species have been studied so far in the MAR compared with the more extensively studied East Pacific Rise (EPR), a generalized comparison is not yet possible, but a species-specific hypothesis may be put forward to explain the lack of structure observed for $R$. exoculata. High dispersal potential, including long-distance migration as a bet-hedging strategy enabling escape from extinction, is thought to be a factor of risk in a fragmented environment due to the possible loss of progeny in non suitable habitats. In $R$. exoculata, this risk may be compensated by a high fecundity, as suggested for other species (Llodra et al., 2000), and/or by an environmentally driven delay in larval development, suggested by O’Connor et al. (2007) to be a possible adaptation associated with large scale dispersal. Moreover, it has been suggested that $R$. exoculata may detect black-body
radiation associated with high-temperature vents (Van Dover et al., 1989, Herring et al., 1999); this may be a specific adaptation of this species to locate its exceptional habitat during large scale dispersal, allowing the benefit of the large scale dispersal to be kept while minimizing the associated risk of loss at the time of settlement. However, empirical data have shown that the detection of temperature in seawater may only be possible within a small area around the chimneys where the seawater is actually warmer (Segonzac et al., 1993). The putative mechanism that may allow *R. exoculata* larvae to settle in their habitat after this hypothetical long-distance dispersal is therefore not elucidated, it may be temperature-associated or based on chemical signals linked to the presence of adults, as has been demonstrated for other marine invertebrates such as the oyster *Ostrea puelchana*, where preferential settlement of spat occurs close to the adults due to a chemical stimulus (Pascual & Zampatti, 1995).

This study makes a novel contribution to our understanding of the biogeography of the Mid-Atlantic Ridge by providing new insight into the population biology and biogeographic structure of a key species of shrimp from the unique, extreme and remote marine habitats of hydrothermal vents. Our results indicate that, across this very remote biogeographic region, all vent populations of this shrimp have a recent common history derived from a bottleneck or founder event, followed by a demographic expansion toward apparently large effective population sizes. Associated with high dispersal potential, such history possibly results in the absence of significant drift effects. Interestingly, these results may indicate a possible common source or centre of origin of a (re)colonization event at the origin of the lineages sampled from the MAR in our study. The identification of such a potential source population is, unfortunately, rendered particularly difficult in these ecosystems because the sampling of hydrothermal vents is necessarily as fragmented in space and time as the ecosystems themselves are. Finally, more species should be studied on the MAR to compare results with the more extensively studied EPR (East Pacific Rise) in order to test for the
possible influence of the shape of ridges and transform faults on the pattern of evolution and genetic differentiation of species associated with hydrothermal vents.
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**Supporting information**
Additional Supporting Information may be found in the online version of this article:

Appendix S1 Neighbour-joining phylogram constructed with 152 COI mitochondrial sequences of *Rimicaris exoculata*

As a service to our authors and readers, this journal provides supporting information supplied by the authors. Such materials are peer-reviewed and may be re-organized for online delivery, but are not copy-edited or typeset. Technical support issues arising from supporting information (other than missing files) should be addressed to the authors.

Biosketches

The researchers involved in this study have a long-standing interest in Biogeography, Ecology and Evolution of marine organisms and especially deep-sea hydrothermal vent communities (D.D.) and their symbiotic relationships (M.A-C.B). Author contributions: S.A-H. and D.D. conceived the ideas; M-A.C-B. collected most of the samples, S.T. collected the data; S.T., S.A.-H. and E.S. analysed the data; and all authors contributed to the writing.

Editor: Craig McClain
## Tables

**Table 1.** Geographic coordinates, depth and cruise details for the populations of organisms sampled along the Mid-Atlantic Ridge.

<table>
<thead>
<tr>
<th>Population</th>
<th>Year</th>
<th>GPS coordinates</th>
<th>Depth</th>
<th>Cruise</th>
</tr>
</thead>
<tbody>
<tr>
<td>Rainbow</td>
<td>2007</td>
<td>036°08’44”N; 034°00’02”W</td>
<td>2200 m</td>
<td>MoMAR; ROV Victor6000; RV Pourquoi pas?</td>
</tr>
<tr>
<td>TAG</td>
<td>2005</td>
<td>26°02’34”N; 044°54’00”W</td>
<td>3650 m</td>
<td>EXOMAR; ROV Victor6000; RV L’Atalante</td>
</tr>
<tr>
<td>Logatchev (Irina2)</td>
<td>2007</td>
<td>14°44’62”N; 046°35’59”W</td>
<td>2860 m</td>
<td>SERPENTINE; ROV Victor6000; RV Pourquoi pas?</td>
</tr>
<tr>
<td>Ashadzé (SE1)</td>
<td>2007</td>
<td>12°50’71”N; 044°54’47”W</td>
<td>3200 m</td>
<td>SERPENTINE; ROV Victor6000; RV Pourquoi pas?</td>
</tr>
<tr>
<td>SouthMAR</td>
<td>2006</td>
<td>4°47’S; 12°22’W</td>
<td>3048 m</td>
<td>M64/1; ROV Meteor/Quest</td>
</tr>
</tbody>
</table>
Table 2. Genetic diversity indices calculated for each sampled population and year, of the deep-sea shrimp *Rimicaris exoculata* sampled along the Mid-Atlantic Ridge. (*n*) sample size; (*Nh*) number of haplotypes; (*Nph*) number of private haplotypes, i.e. the number of haplotypes that only appeared in one population; (*k*) number of polymorphic sites; (*h*) haplotype diversity; (*π₁*) mean number of pairwise differences; (*π₂*) nucleotide diversity. Neutrality and population expansion tests: $D=\text{Tajima’s D-test;} \; F_{S}=\text{Fu’s } F_{S} \text{ test. All values obtained for the neutrality tests were significant at the 5}\% \text{ level.}$

<table>
<thead>
<tr>
<th>Population</th>
<th>Year</th>
<th><em>n</em></th>
<th><em>k</em></th>
<th><em>Nh</em></th>
<th><em>h</em></th>
<th><em>π₁</em></th>
<th><em>π₂</em></th>
<th><em>D</em></th>
<th><em>F_{S}</em></th>
<th><em>Nph</em></th>
</tr>
</thead>
<tbody>
<tr>
<td>Rainbow</td>
<td>2007</td>
<td>26</td>
<td>18</td>
<td>14</td>
<td>0.82±0.07</td>
<td>1.67</td>
<td>0.0033</td>
<td>-2.292(0.002)</td>
<td>-11.01(0)</td>
<td>3</td>
</tr>
<tr>
<td></td>
<td>1997</td>
<td>32</td>
<td>14</td>
<td>14</td>
<td>0.69±0.09</td>
<td>1.05</td>
<td>0.0021</td>
<td>-2.298(0.001)</td>
<td>-13.87(0)</td>
<td>7</td>
</tr>
<tr>
<td>Logatchev</td>
<td>2007</td>
<td>31</td>
<td>20</td>
<td>16</td>
<td>0.77±0.08</td>
<td>1.41</td>
<td>0.0028</td>
<td>-2.487(0)</td>
<td>-15.38(0)</td>
<td>8</td>
</tr>
<tr>
<td></td>
<td>2006</td>
<td>12</td>
<td>9</td>
<td>7</td>
<td>0.77±0.13</td>
<td>1.50</td>
<td>0.0030</td>
<td>-2.016(0.004)</td>
<td>-3.52(0.003)</td>
<td>3</td>
</tr>
<tr>
<td></td>
<td>2005</td>
<td>13</td>
<td>8</td>
<td>8</td>
<td>0.81±0.11</td>
<td>1.36</td>
<td>0.0027</td>
<td>-1.831(0.012)</td>
<td>-5.25(0)</td>
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</tr>
<tr>
<td>TAG</td>
<td>2005</td>
<td>31</td>
<td>16</td>
<td>16</td>
<td>0.77±0.08</td>
<td>1.09</td>
<td>0.0022</td>
<td>-2.454(0)</td>
<td>-18.32(0)</td>
<td>6</td>
</tr>
<tr>
<td>Ashadzé</td>
<td>2007</td>
<td>3</td>
<td>0</td>
<td>1</td>
<td>0</td>
<td>0</td>
<td>0</td>
<td>0</td>
<td>0</td>
<td>0</td>
</tr>
<tr>
<td>SouthMAR</td>
<td>2006</td>
<td>4</td>
<td>3</td>
<td>4</td>
<td>1.00±0.18</td>
<td>1.50</td>
<td>0.0030</td>
<td>-0.754(0.23)</td>
<td>-2.37(0.008)</td>
<td>0</td>
</tr>
</tbody>
</table>
Table 3. Genetic diversity indices for the temporal samples of the population of *Rimicaris exoculata* from Logatchev. Diversity indices for the sampling years of 2005 and 2007 are given as averages (±SD) due to the random re-sampling (100 times) to a minimum common size of 12 individuals per population. (Nh) number of haplotypes; (k) number of polymorphic sites; (h) haplotype diversity; (\(\pi_2\)) nucleotide diversity.

<table>
<thead>
<tr>
<th>Year</th>
<th>Nh</th>
<th>k</th>
<th>h</th>
<th>(\pi_2)</th>
</tr>
</thead>
<tbody>
<tr>
<td>2005</td>
<td>5.26±1.1</td>
<td>5.1±1.46</td>
<td>0.74±0.12</td>
<td>0.0024</td>
</tr>
<tr>
<td>2006</td>
<td>7</td>
<td>9</td>
<td>0.77±0.13</td>
<td>0.0030</td>
</tr>
<tr>
<td>2007</td>
<td>5.9±1.51</td>
<td>6.78±2.25</td>
<td>0.73±0.15</td>
<td>0.0026</td>
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</tbody>
</table>
Table 4. Pairwise $F_{ST}$ values based on haplotype frequencies obtained for the species *Rimicaris exoculata* sampled along the Mid-Atlantic Ridge. None of the $F_{ST}$ values were significant at the 5% level.

<table>
<thead>
<tr>
<th>Pops</th>
<th>Rainbow07</th>
<th>Rainbow97</th>
<th>Logatchev07</th>
<th>Logatchev06</th>
<th>Logatchev05</th>
<th>TAG</th>
<th>Ashadzé</th>
<th>South MAR</th>
</tr>
</thead>
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<tr>
<td>Rainbow07</td>
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<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
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<tr>
<td>Rainbow97</td>
<td></td>
<td></td>
<td>-0.0115</td>
<td>-0.01243</td>
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<tr>
<td>Logatchev07</td>
<td>-0.0115</td>
<td>-0.01243</td>
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<tr>
<td>Logatchev06</td>
<td>-0.0184</td>
<td>-0.0221</td>
<td>-0.0298</td>
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<tr>
<td>Logatchev05</td>
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<td>-0.0227</td>
<td>-0.0360</td>
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</tr>
<tr>
<td>TAG</td>
<td>-0.0147</td>
<td>-0.0138</td>
<td>-0.0122</td>
<td>-0.0262</td>
<td>-0.0260</td>
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</tr>
<tr>
<td>Ashadzé</td>
<td>0.0480</td>
<td>-0.0230</td>
<td>0.0140</td>
<td>0.0145</td>
<td>0.0331</td>
<td>0.014</td>
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</tr>
<tr>
<td>South MAR</td>
<td>-0.0589</td>
<td>0.0452</td>
<td>0.0031</td>
<td>-0.0147</td>
<td>-0.0495</td>
<td>0.003</td>
<td>0.250</td>
<td></td>
</tr>
</tbody>
</table>
Figures

Figure 1. Location of the *Rimicaris exoculata* populations sampled along the Mid-Atlantic Ridge, and the respective mtDNA haplotype distribution (pie charts) per population. The size of the pie charts is proportional to the number of individuals sampled from each population, and each colour in the pie charts represents a different haplotype; the singletons (haplotypes that appear only once in one population) are coded as white, the same haplotype found in different populations is represented, by the same colour in the corresponding pie charts. a) Key of the locations analysed along the MAR; b) colour key for the haplotypes found.
Figure 2. Haplotype network of the obtained mtDNA haplotypes of *Rimicaris exoculata* sampled along the Mid-Atlantic Ridge. Each circle represents a different haplotype, with the size of each circle proportional to the number of individuals displaying that particular haplotype. The white circles represent singletons (haplotypes that occur only once, represented by a single individual), the haplotypes found in more than one individual are identified and represented by pie charts, the colours used represent the locations where that haplotype was found. Segment size is proportional to the relative frequency of a haplotype in each population where it is present. a) Colour key to the locations where each haplotype was present.
Figure 3. Mismatch distribution of nucleotide pairwise differences for all the mitochondrial COI sequences analysed (152) for *Rimicaris exoculata* sampled along the Mid-Atlantic Ridge.

Unweighted mean pairwise difference: 1.277
Figure 4. Observed and simulated mismatch curves of nucleotide pairwise differences of mitochondrial COI sequences of *Rimicaris exoculata* sampled along the Mid-Atlantic Ridge. The graphs represent a comparison of the observed curve of nucleotide pairwise differences to (a) a simulated curve assuming constant population size and (b) to a simulated curve assuming a population growth/decline model.
Supporting information

Fig. S1. Neighbour-joining phylogram constructed under the HKY mutation model with 152 COI mitochondrial sequences of *Rimicaris exoculata*. The outgroup used was the bresiliid shrimp *Alvinocaris* sp. All bootstrap values are shown. a) Colour key to the geographic location where the sequences were present.